
Biophysics of Human Neutrophil Haptokinesis

Steven J. Henry

Funding:

NIH HL18208 to DAH

NSF GRFP to SJH

Committee:

Daniel A. Hammer, PhD (advisor)

Scott L. Diamond, PhD (chair)

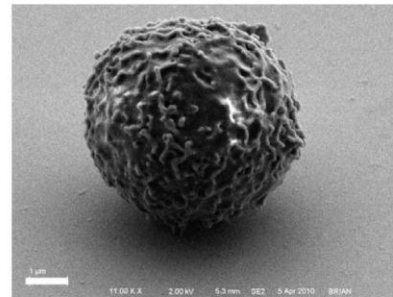
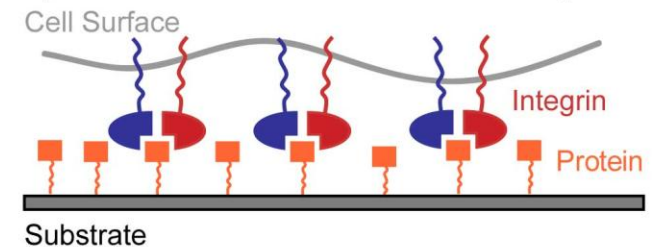
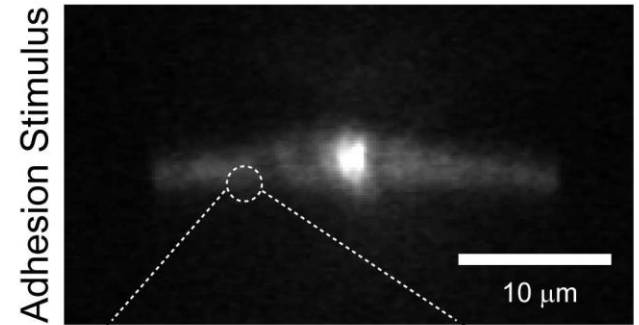
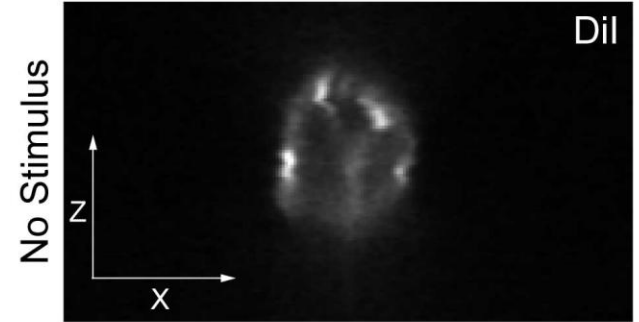
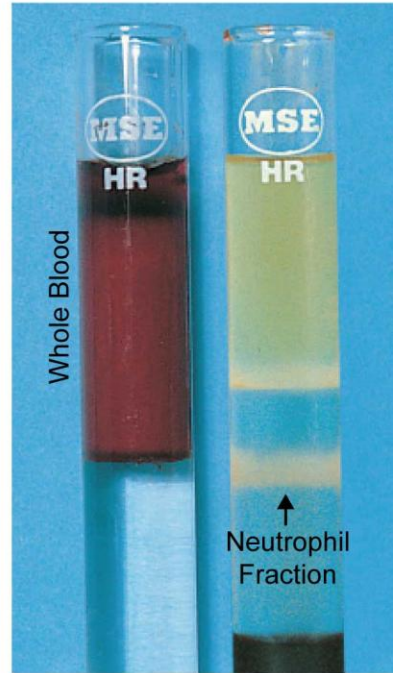
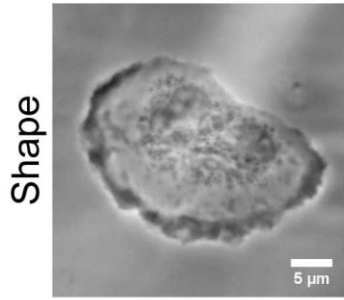
John C. Crocker, PhD

Donggeun Huh, PhD

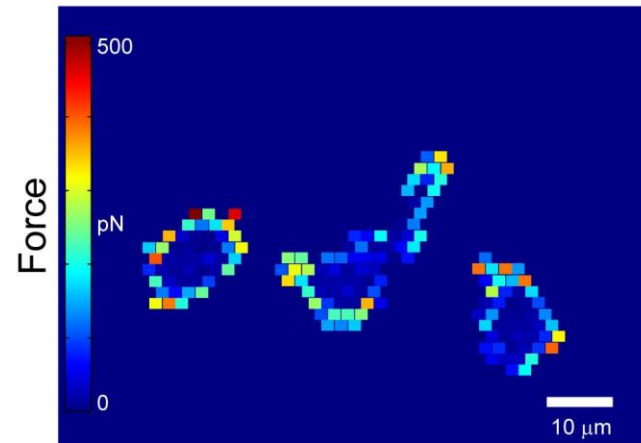
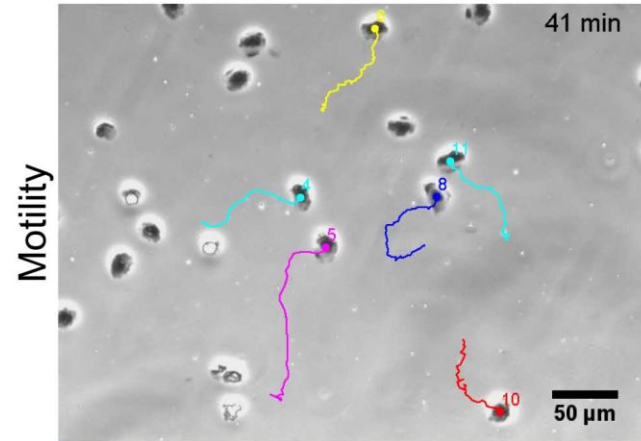


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Biophysics of Human Neutrophil Haptokinesis

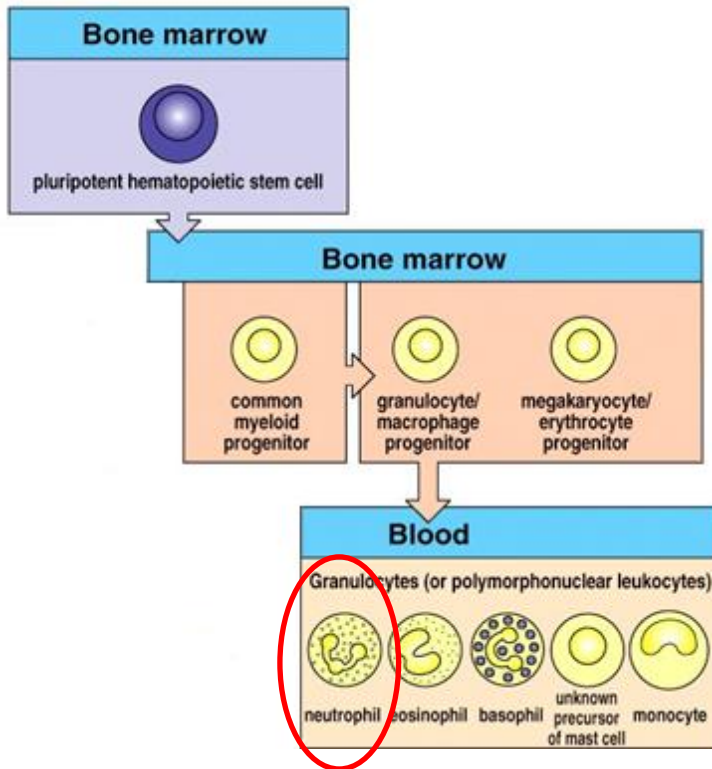


Lomakina et al. 2014. *Biophys J.*



Neutrophils: first responders to trauma and infection

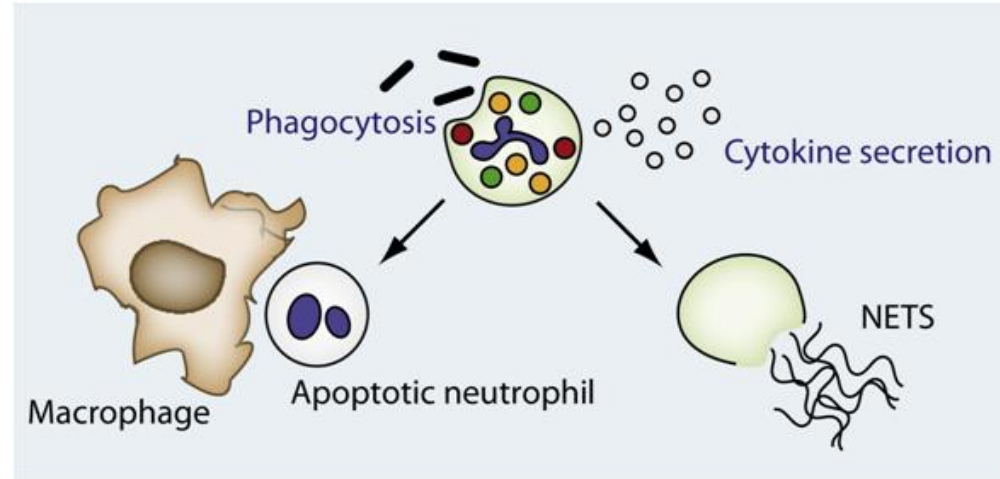
White Blood Cell



Janeway et al. *Immunobiology*. 6th Ed.

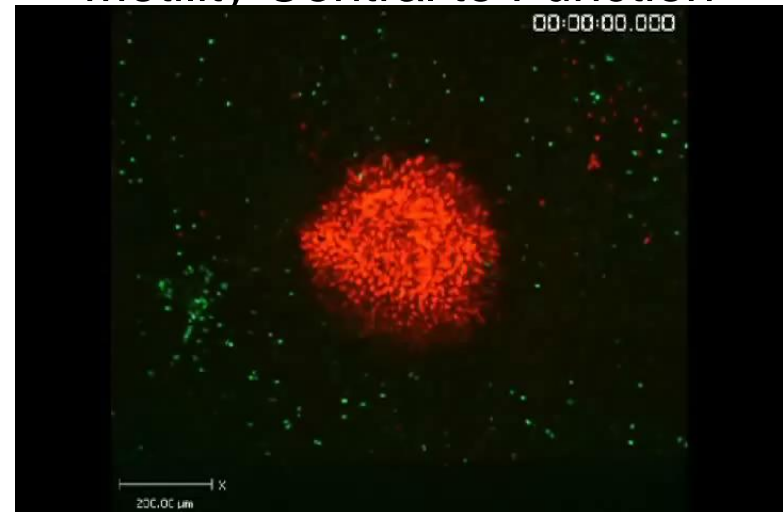
66% marrow production = neutrophils
10¹¹ neutrophils/day

Fast (sec-min) Response Times



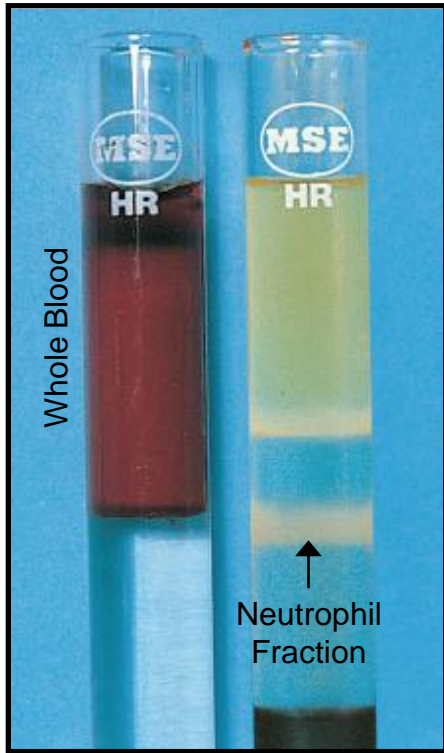
Borregaard. 2010. *Immunity*.

Motility Central to Function



McDonald et al. 2010. *Science*.

Neutrophils: a model cell type



Axis-Shield

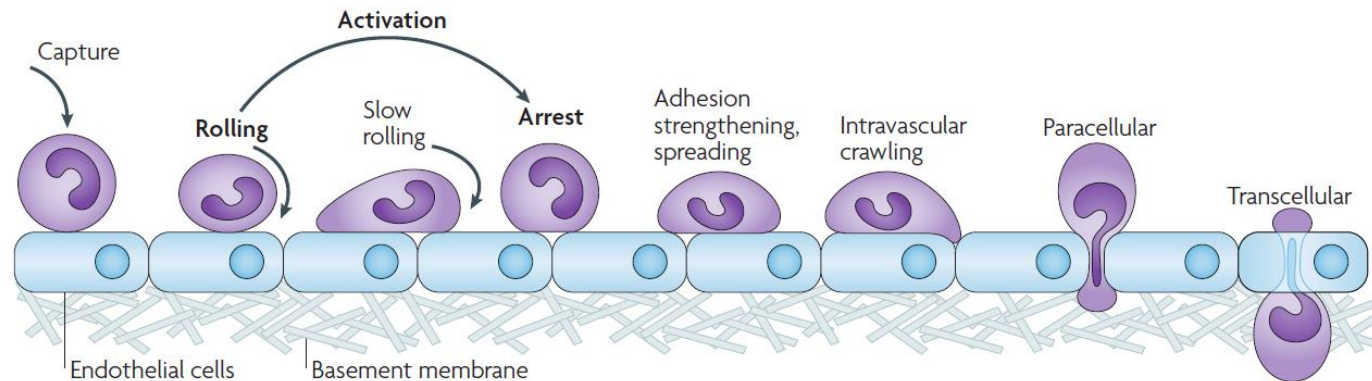
Minimally invasive: venipuncture

Ubiquitous: $\sim 10^6$ cells/mL whole blood

Fast-acting : sec-min

Highly motile: ~ 10 μ m/min

Leukocyte Adhesion Cascade

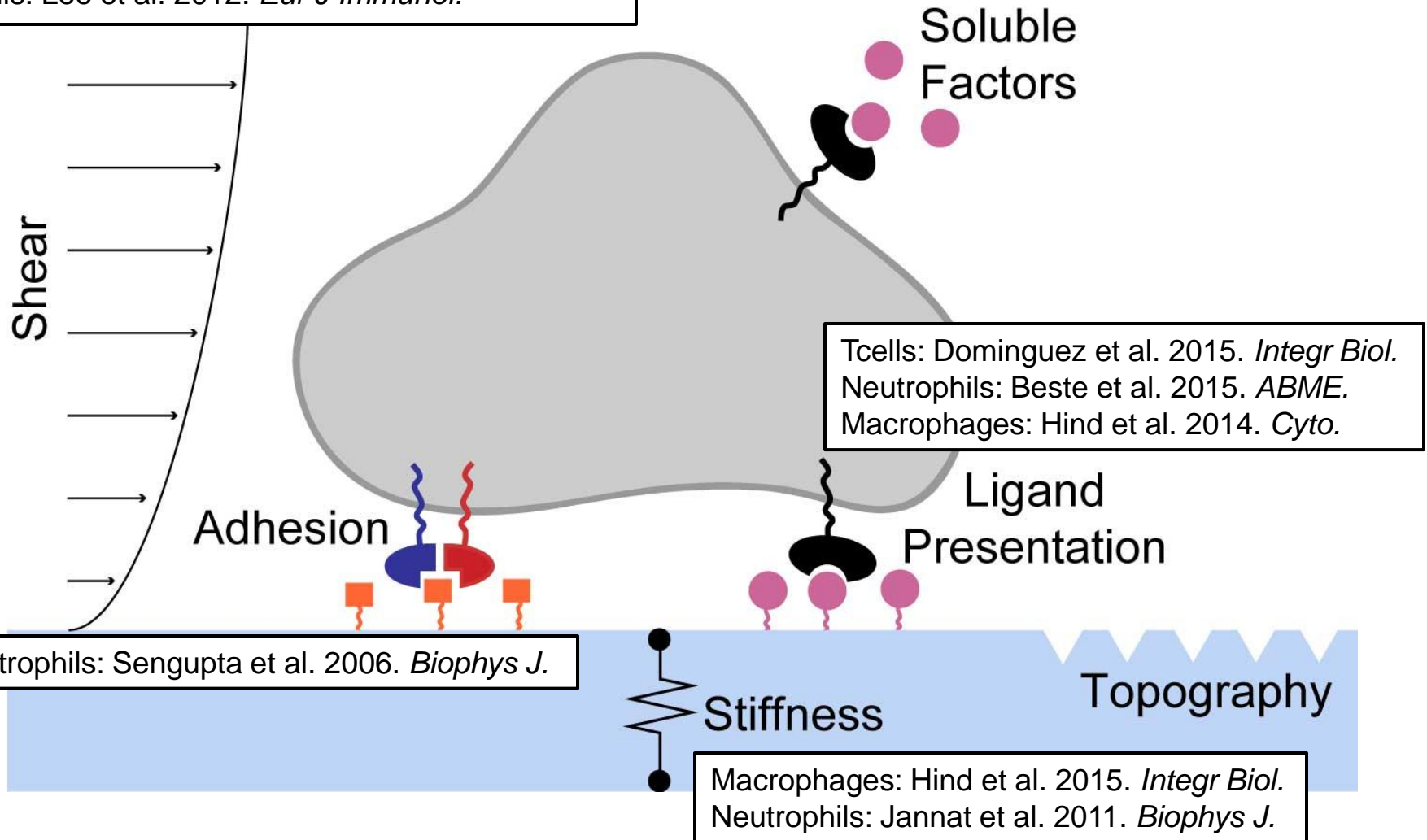


Ley. 2007. *Nat Rev Immunol.*

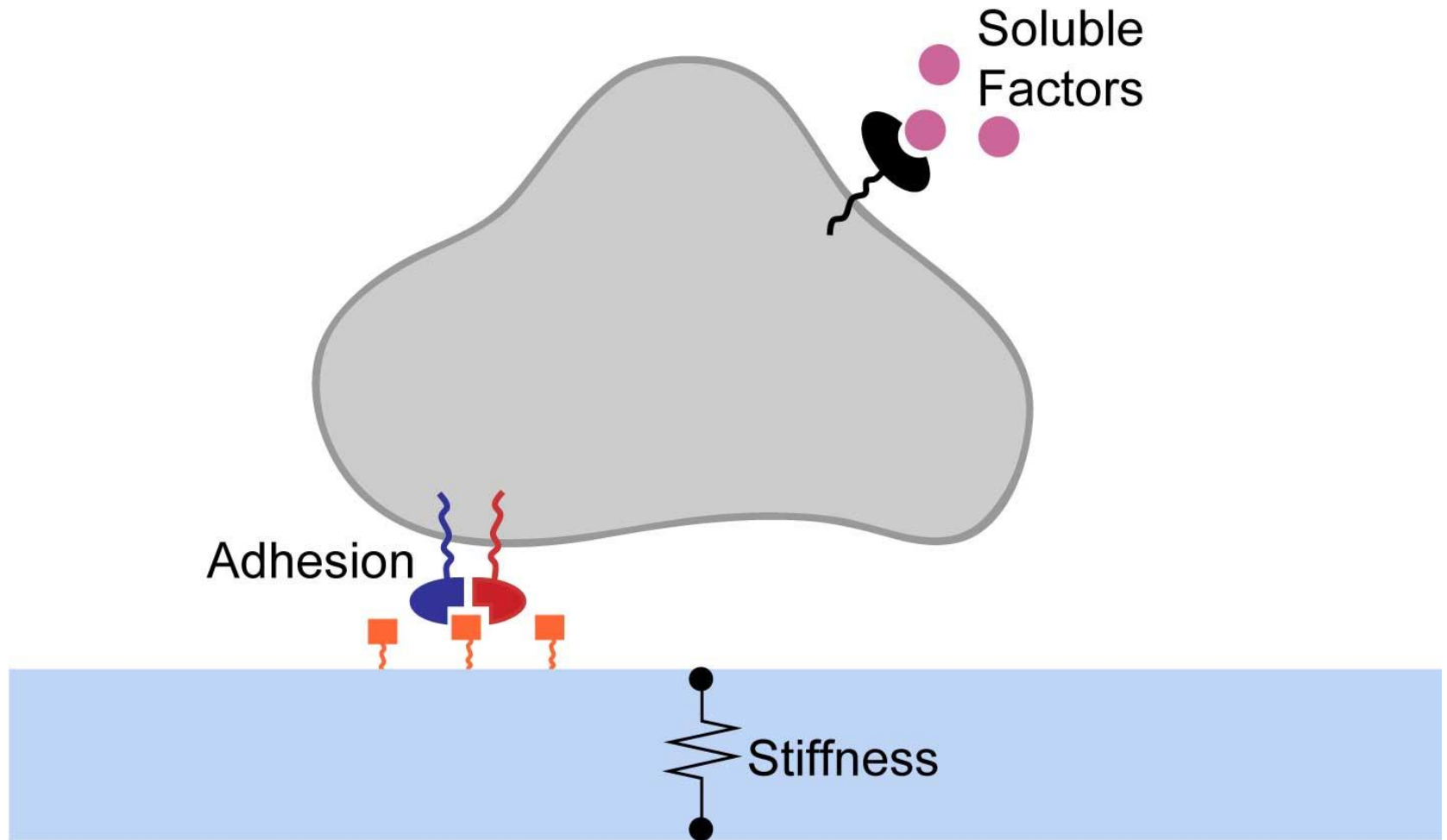
Cell environments are complex (multi-stimulatory)

Tcells: Dominguez et al. 2015. *Integr Biol.*
Neutrophils: Pepper et al. 2013. *Prot Eng Des Sel.*
Tcells: Lee et al. 2012. *Eur J Immunol.*

DCs: Ricart et al. 2011. *J Immunol.*
Neutrophils: Smith et al. 2007. *Biophys J.*

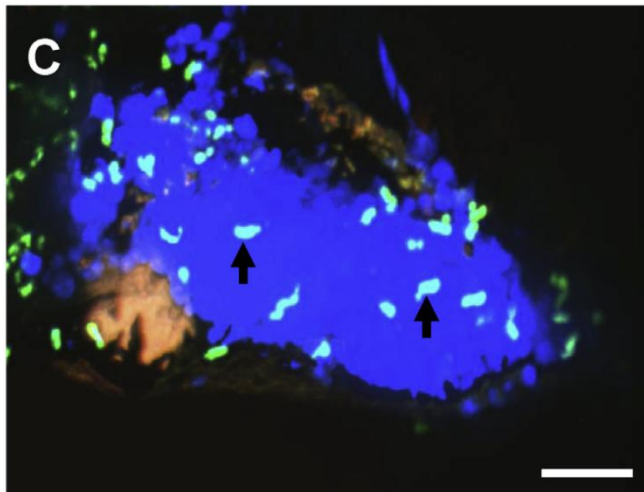


Today, neutrophil responses to:

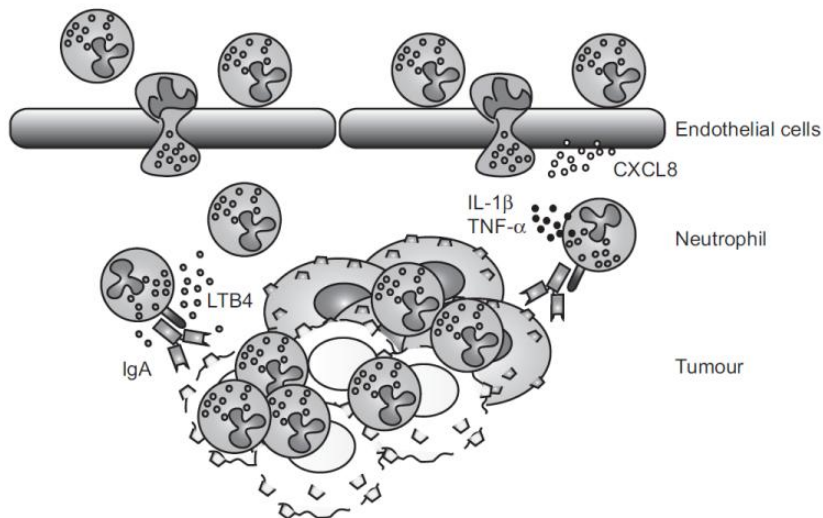


Why we should care ... therapies of ~~tomorrow~~ Today!

Neutrophils Infiltrate Tumors



Tazzyman. 2013. *Sem Canc Bio*.



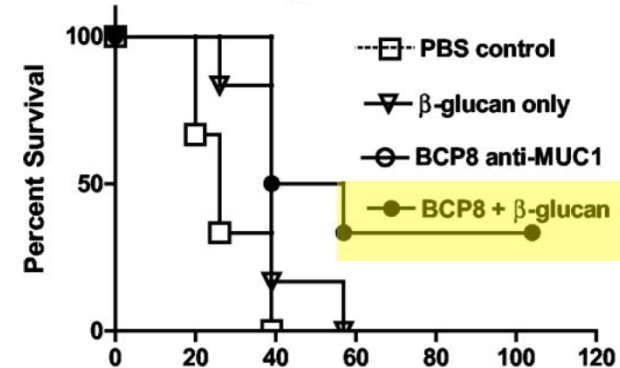
van Egmond et al. 2013. *Sem Canc Bio*.

Imprime PGG®
(β -glucan as
Leukocyte
Adjuvant)

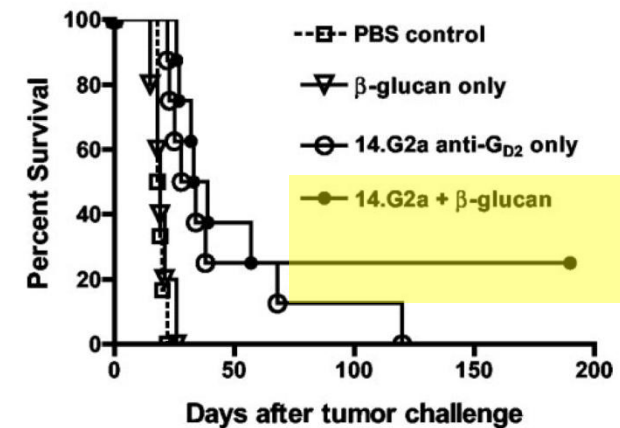


www.biothera.com

Liver Tumors



Lung Tumors



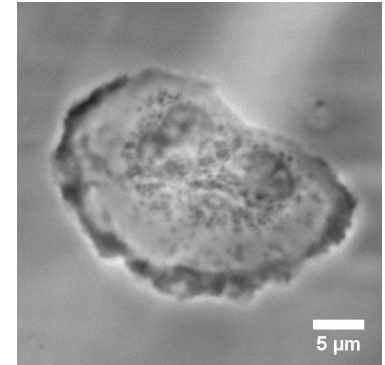
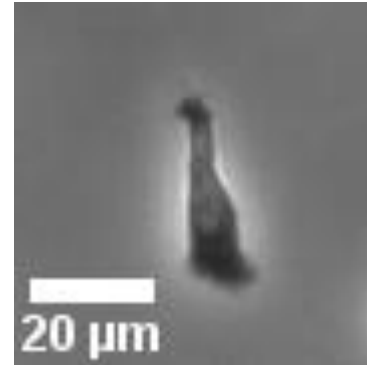
Hong et al. 2003. *Canc Res*.

Outline

Shape and Motility

Ligand density elicits phenotypic switch in human neutrophils

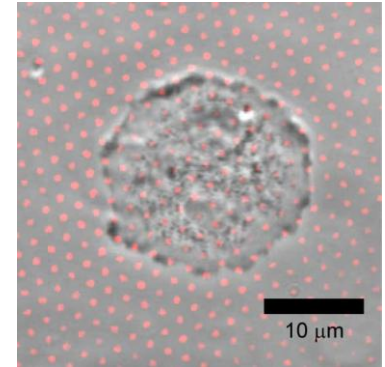
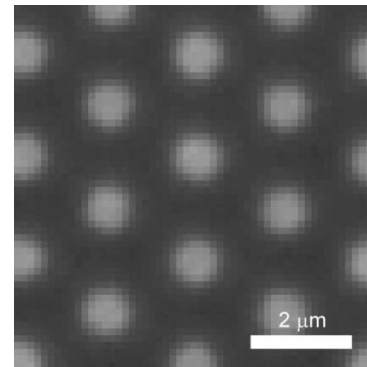
Henry, Crocker, Hammer. 2014. *Integr Biol.*



Density Sensing

Dynamic traction forces of spreading and adherent human neutrophils

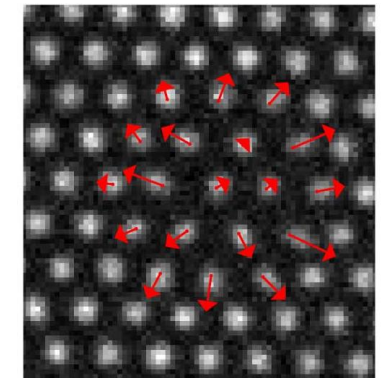
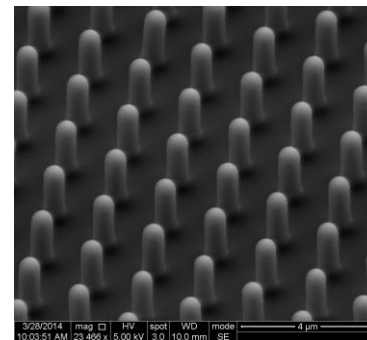
Henry, Crocker, Hammer. 2015. *ABME* (In Prep)



Spreading Mechanics

Dynamic traction forces of spreading and adherent human neutrophils

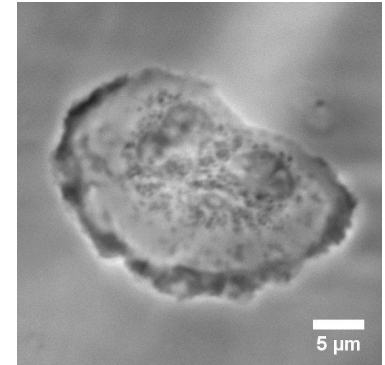
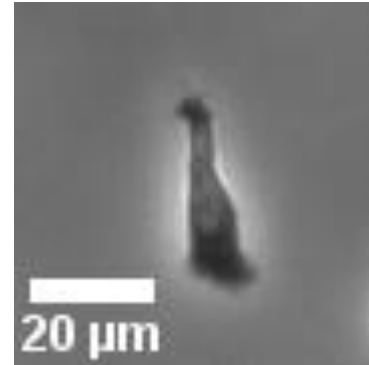
Henry, Chen, Crocker, Hammer. 2015. *Biophys J.* (Under Revision)



Shape and Motility

Ligand density elicits phenotypic switch in human neutrophils

Henry, Crocker, Hammer. 2014. *Integr Biol.*



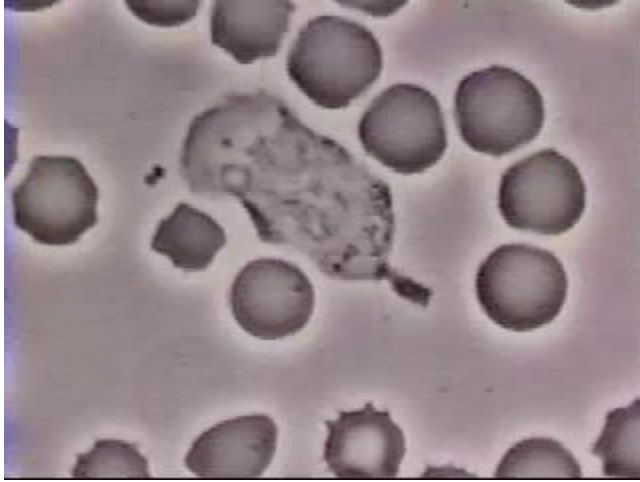
Aim:

Quantify effect of adhesion density on neutrophil shape and motility

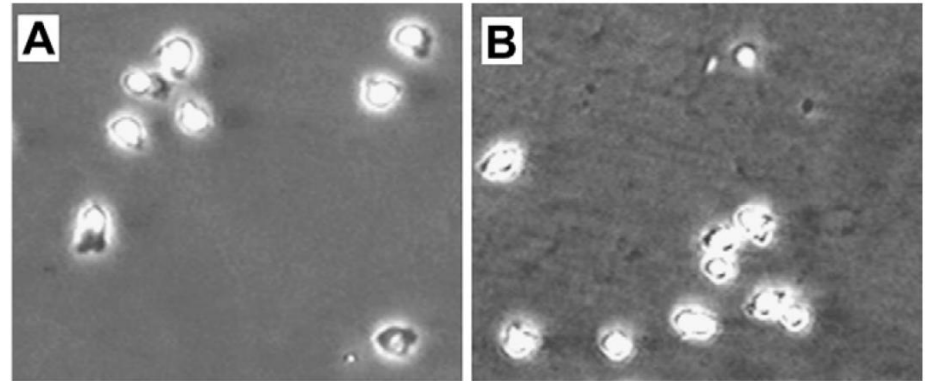
Hypotheses:

Neutrophil shape and motility are adhesion-sensitive
Integrin receptors will mediate this adhesion

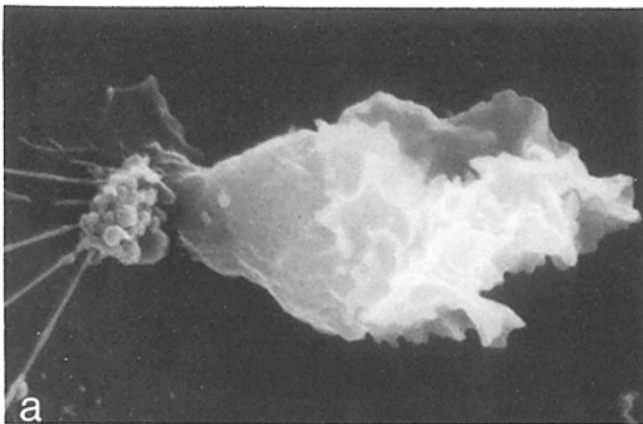
Canonical amoeboid phenotype of neutrophils



David Rogers, 1950s

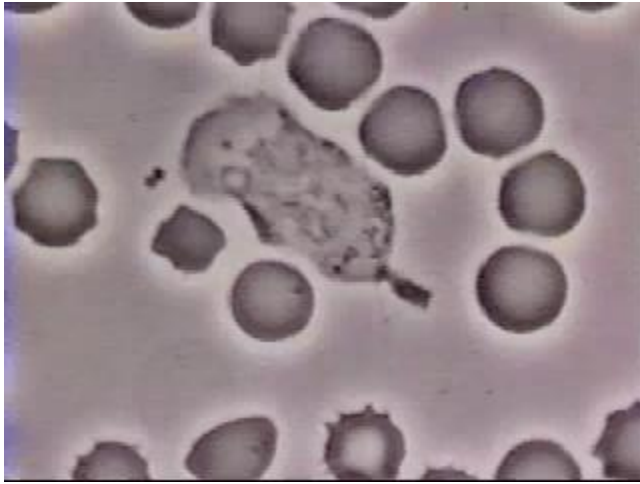


Butler et al. 2008. *Cell Immunol.*

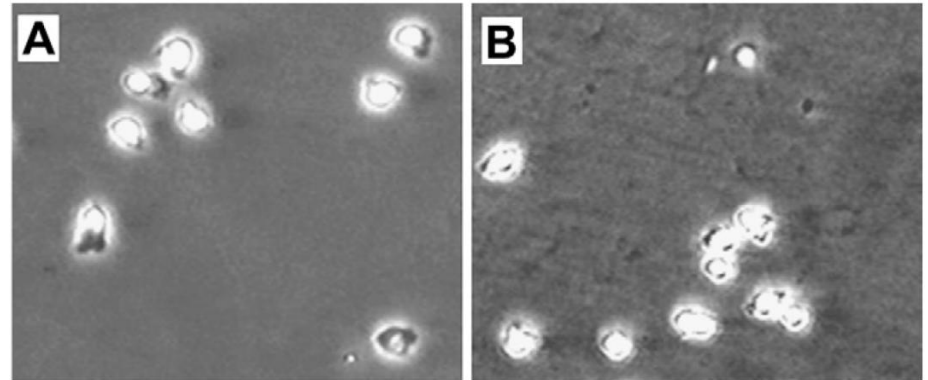


Cassimeris et al. 1990. *JCB.*

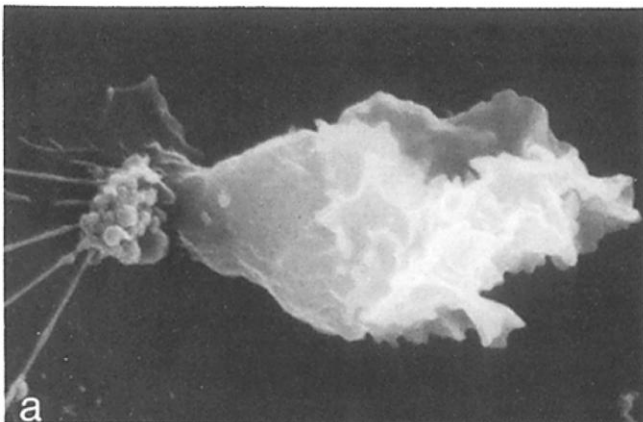
Can adhesivity reconcile these conflicting observations?



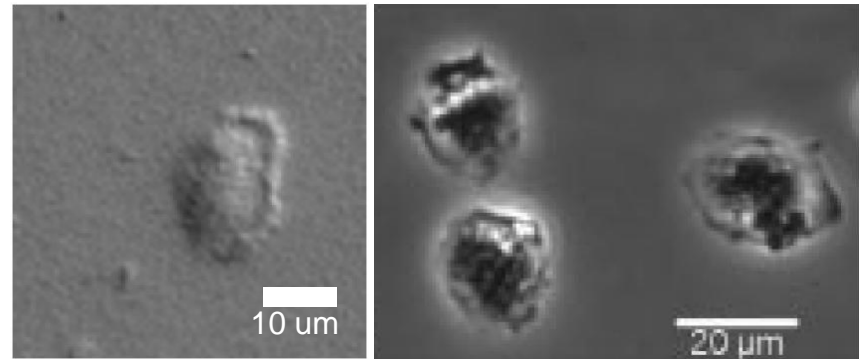
David Rogers, 1950s



Butler et al. 2008. *Cell Immunol.*



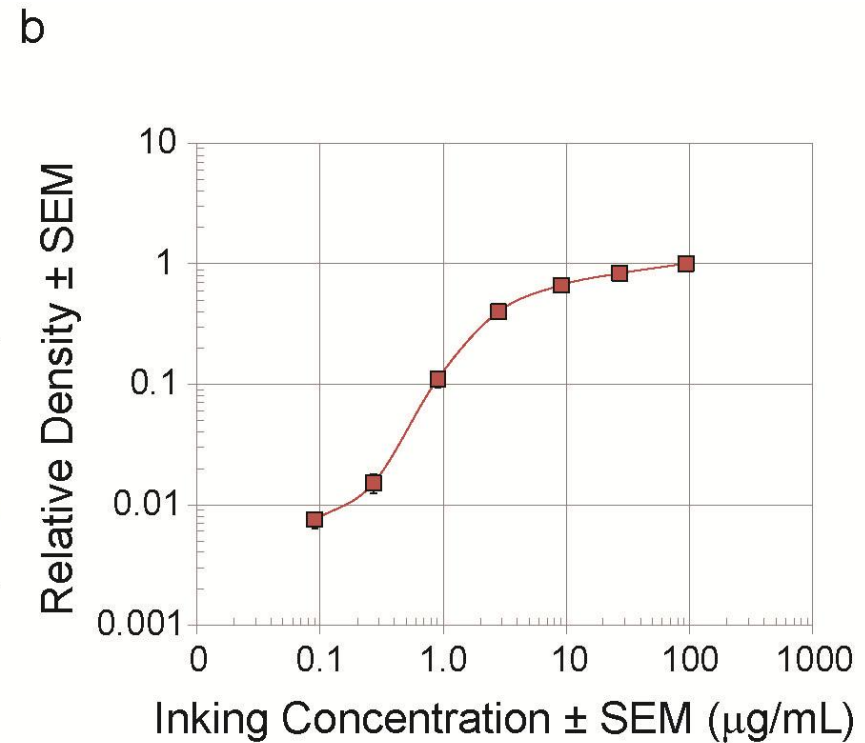
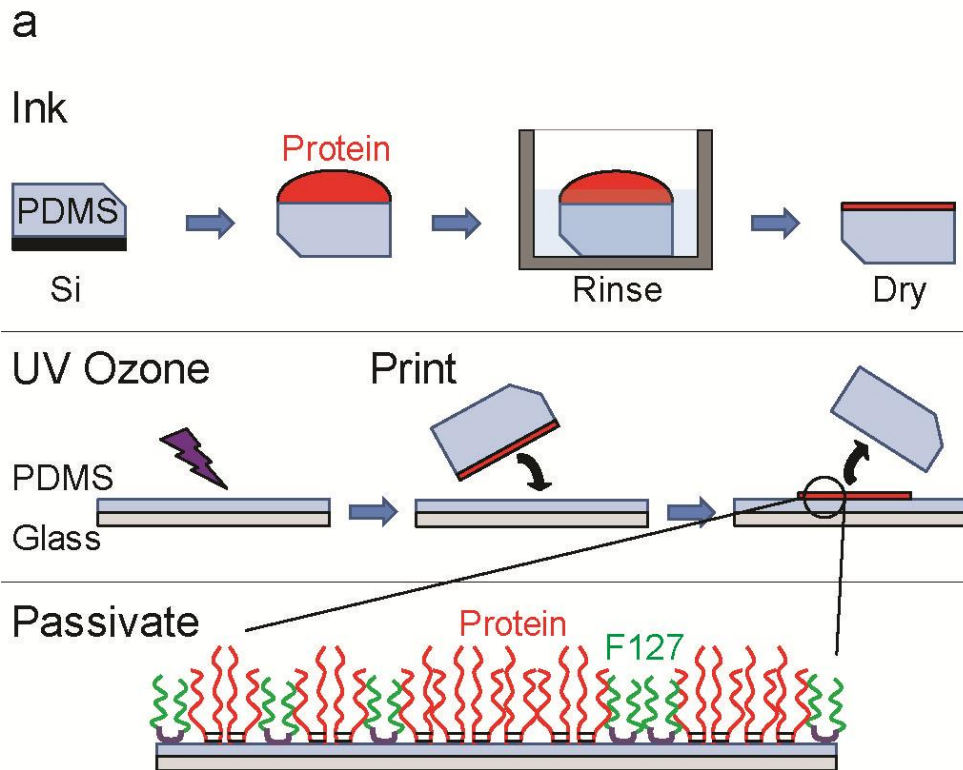
Cassimeris et al. 1990. *JCB.*



Oakes et al. 2009.
Blood.

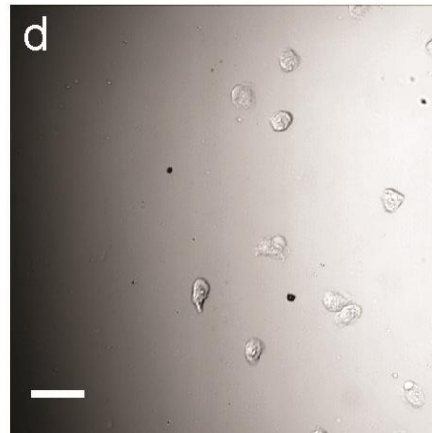
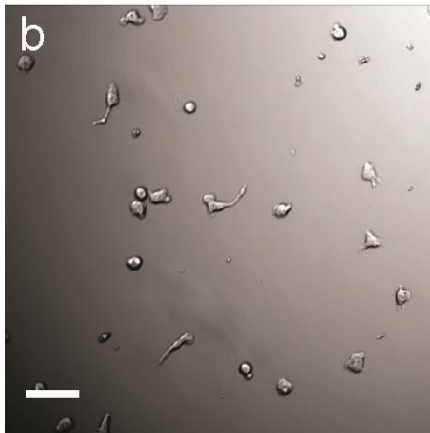
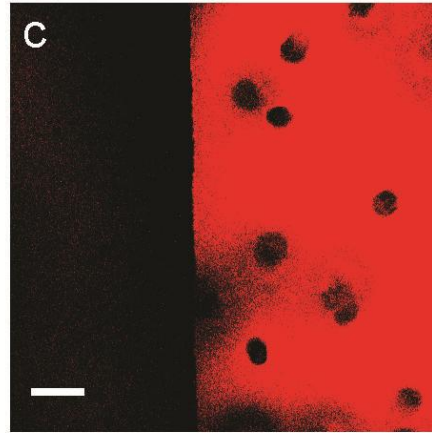
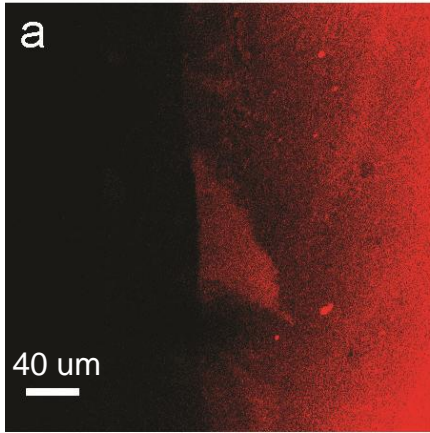
Stroka et al. 2009.
Cyto.

Tuning Adhesivity via Microcontact Printing

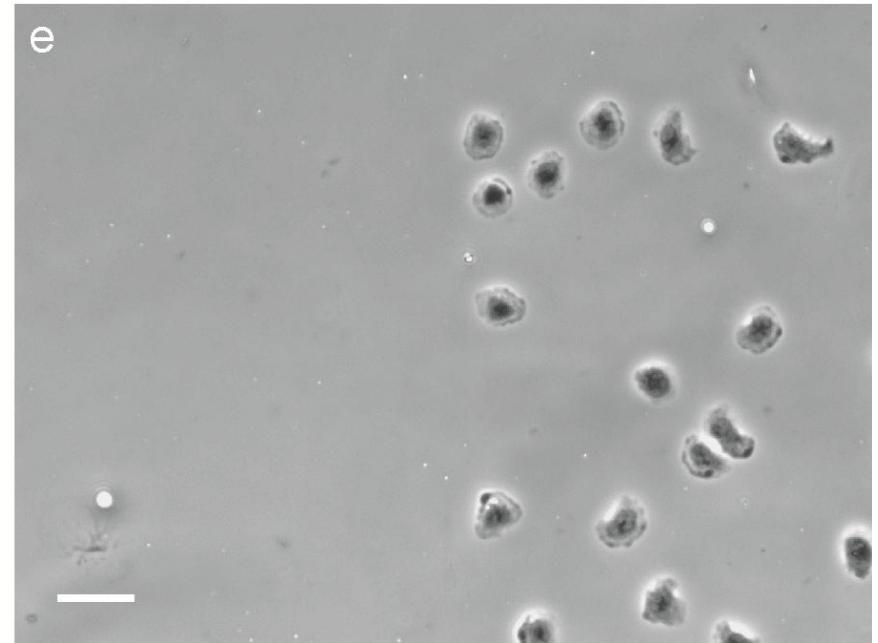


Exquisite cell-ligand specificity

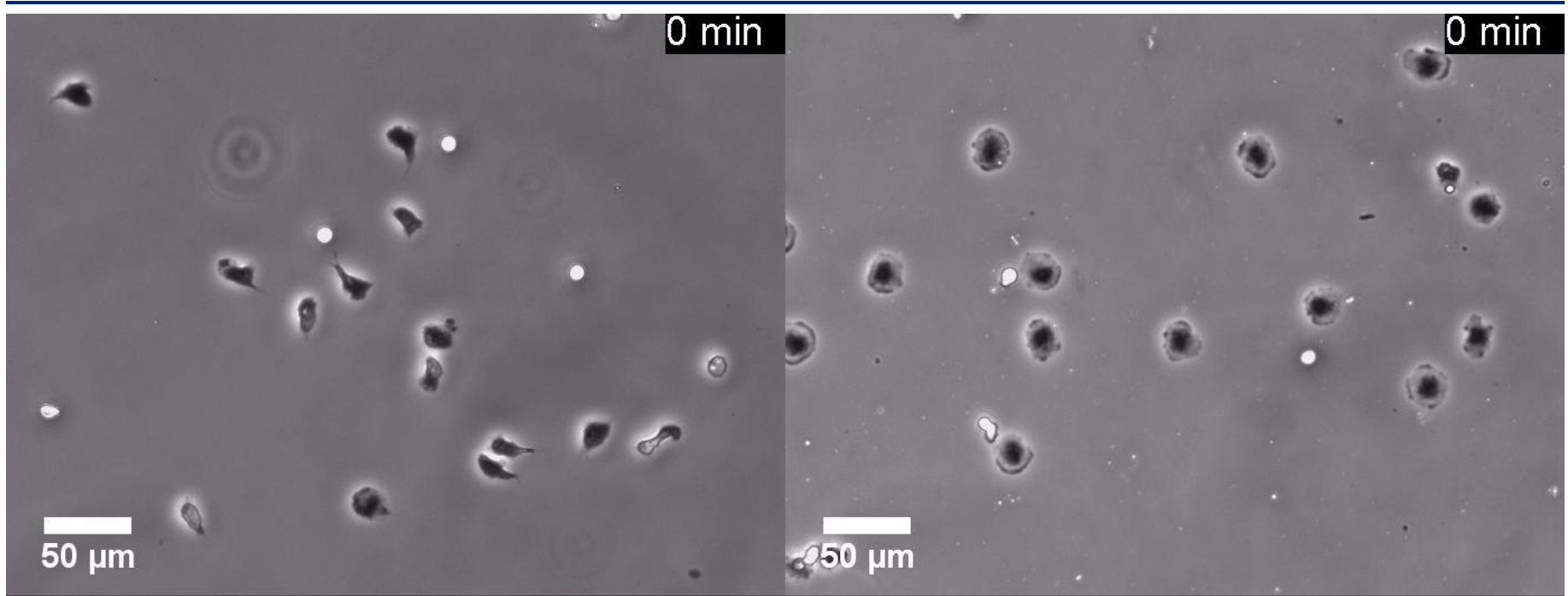
BSA Blocked



Pluronic Blocked



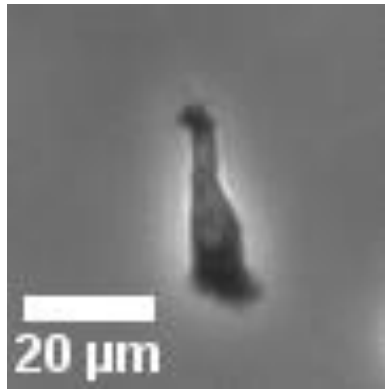
Two dramatically different modes of motility



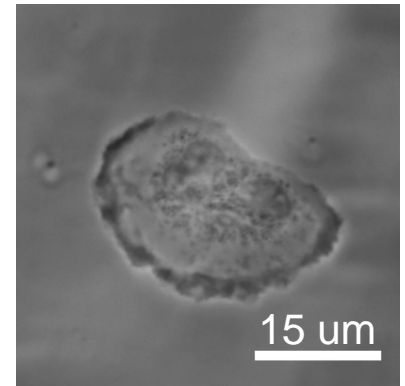
Amoeboid

“Keratocyte-Like”

Highly
Adhesive
Surface

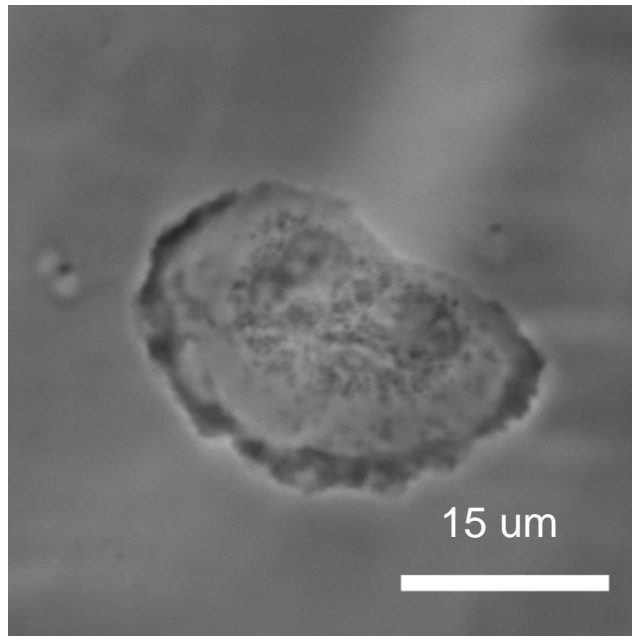


Moderately
Adhesive
Surface



“Keratocyte-like” morphology

Neutrophils

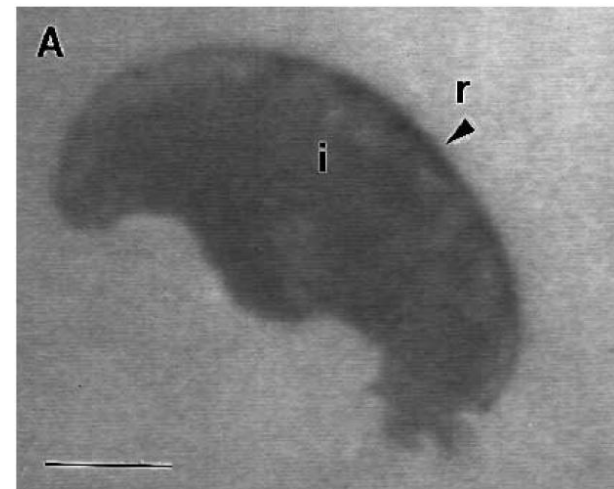


Henry et al. 2014. *Integr Biol.*

Epithelial Keratocytes

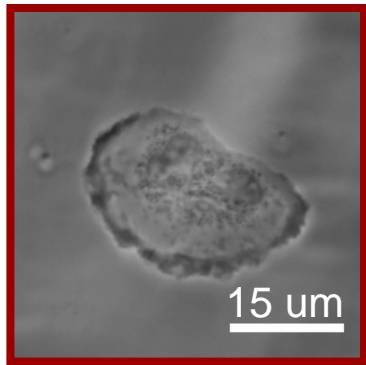


Keren et al. 2008. *Nature.*

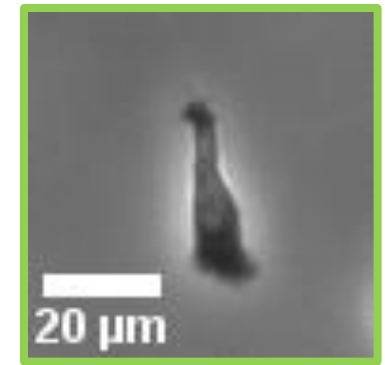
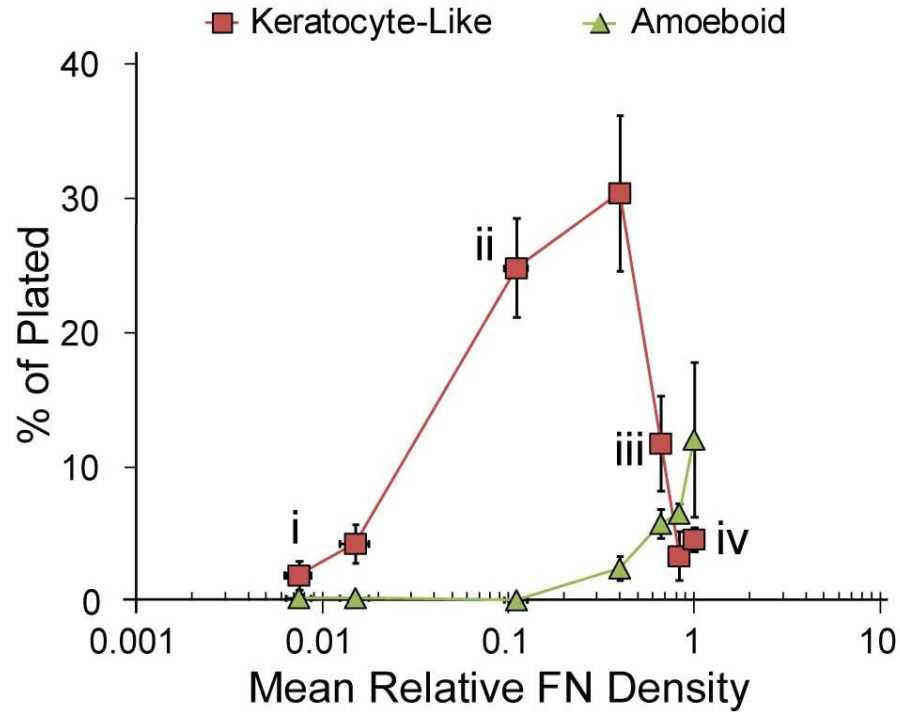


Lee et al. 1997. *JCS.*

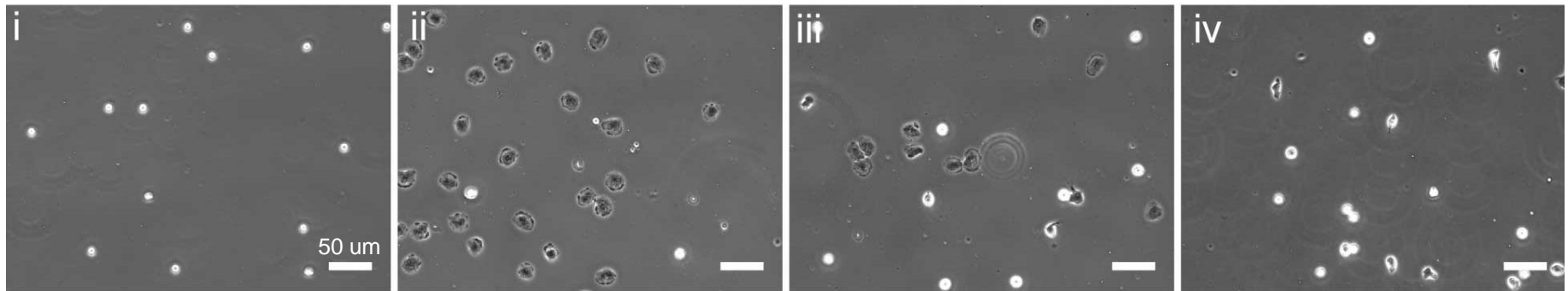
Fibronectin density as controller of shape



Keratocyte-Like



Amoeboid

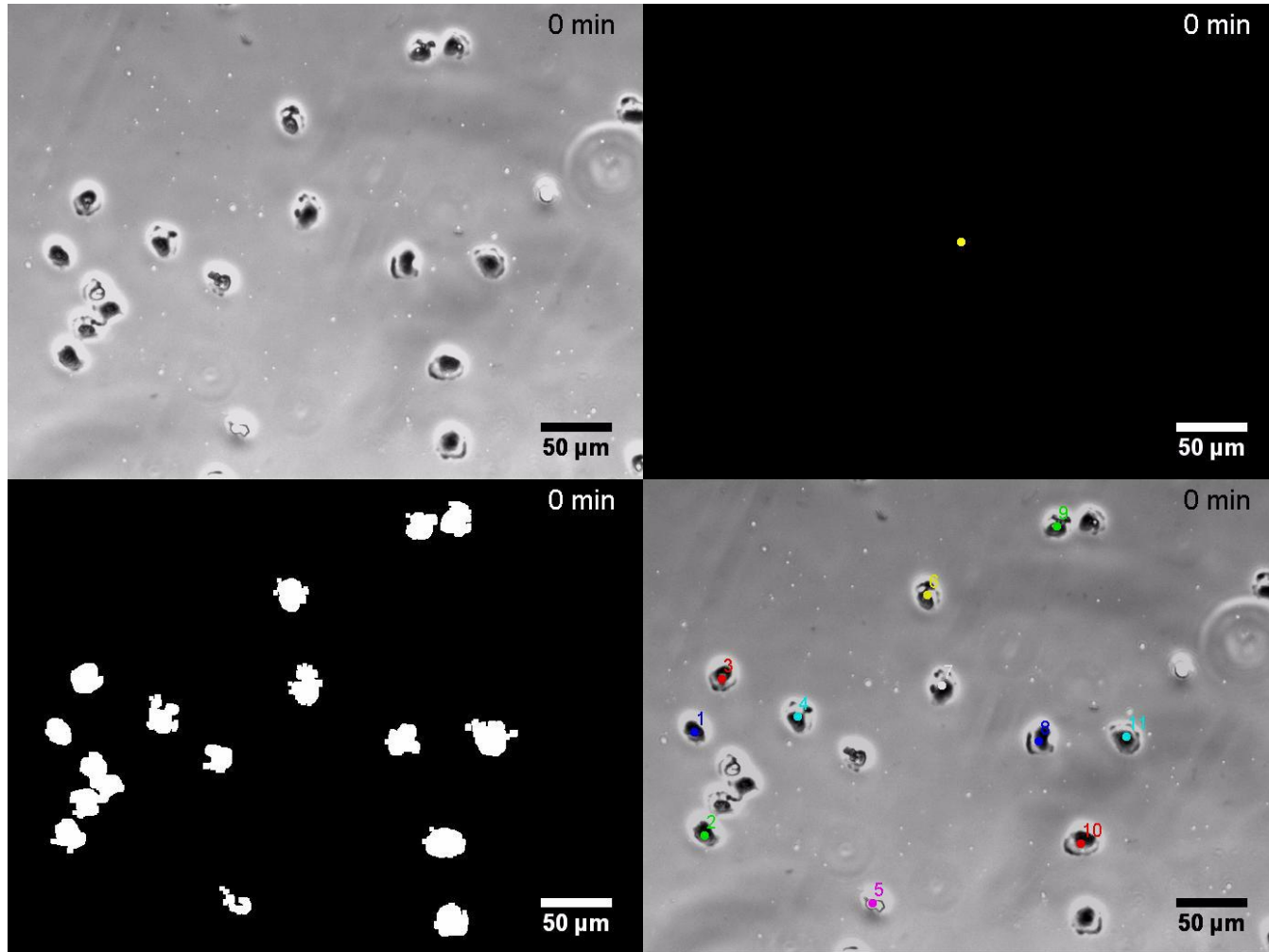


Increasing FN



Objective and reproducible cell tracking

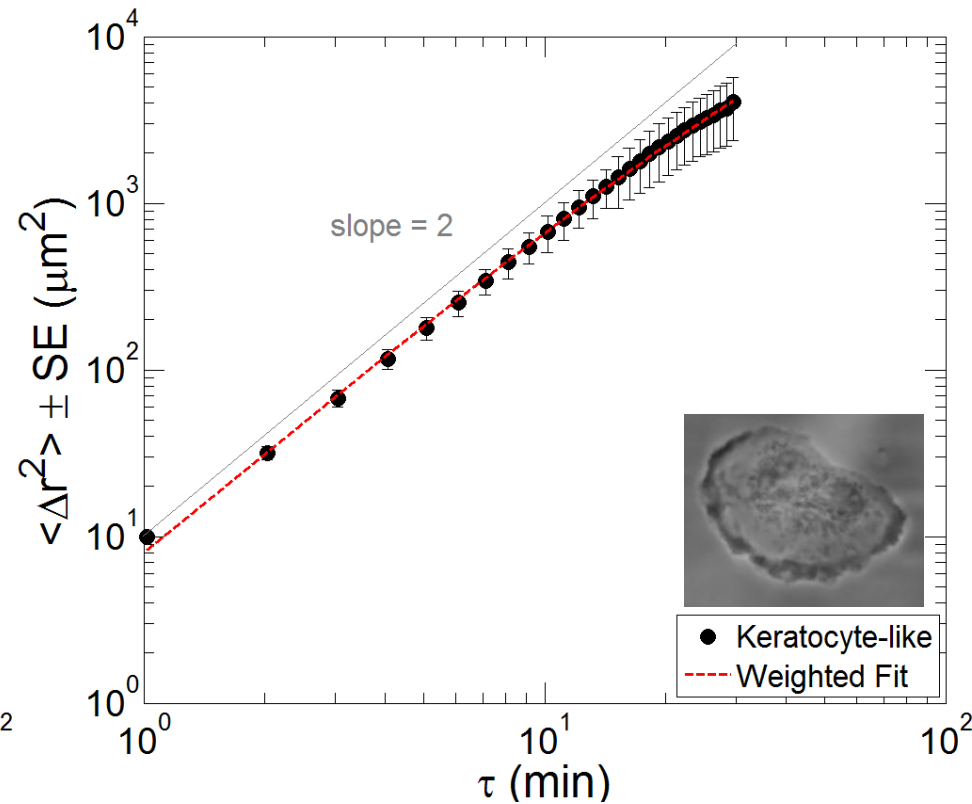
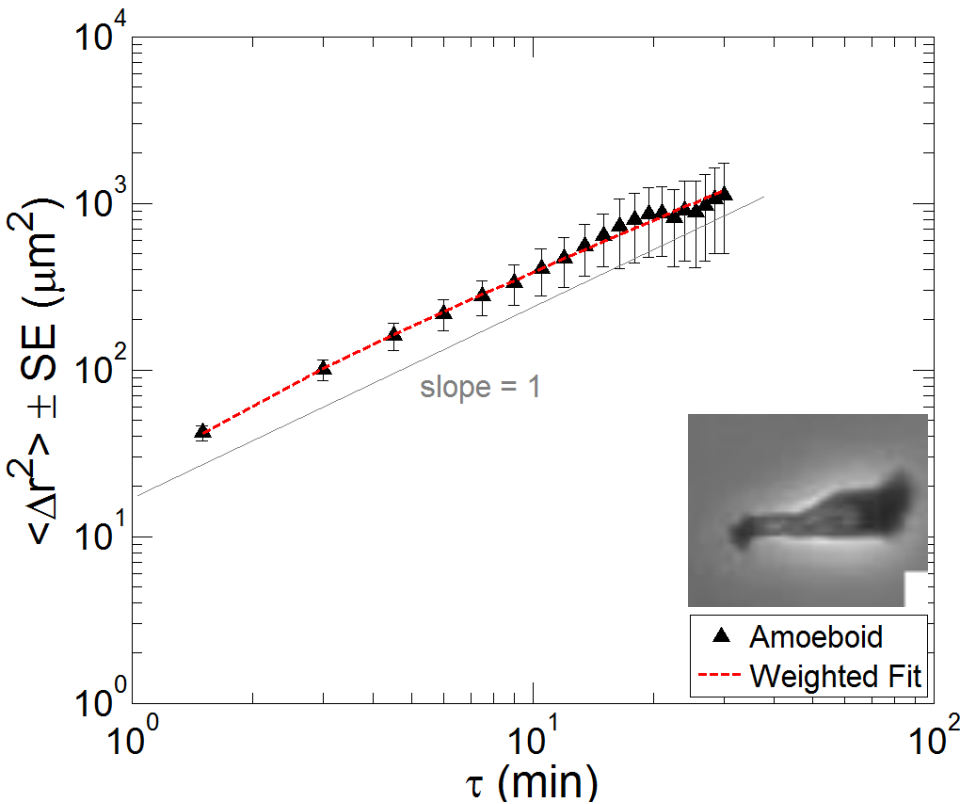
Segment



Merge

Link Centroids

Motility as a persistent random walk



$$\langle \Delta r^2(\tau) \rangle = 2S^2P[\tau - P(1 - \exp(-\tau/P))]$$

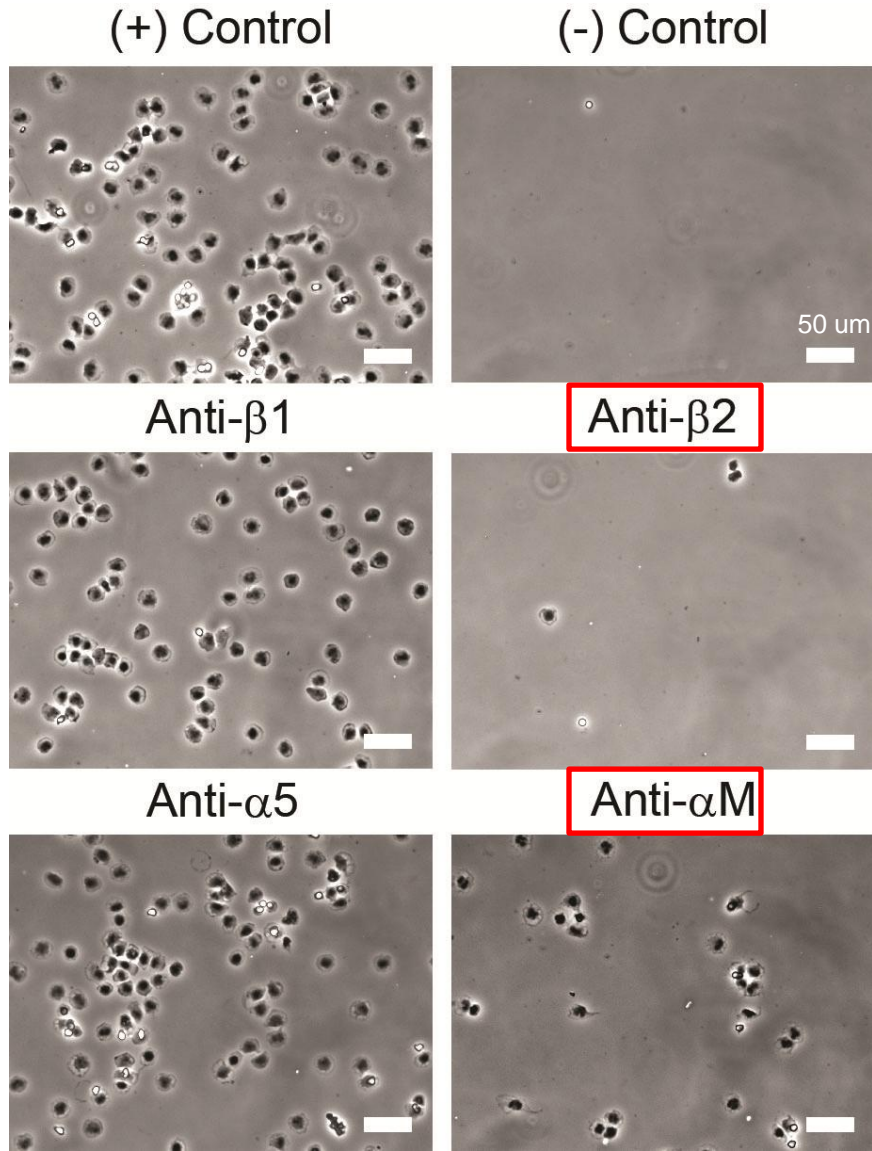
$$S = 6 \mu\text{m}/\text{min}$$

$$P = 0.5 \text{ min}$$

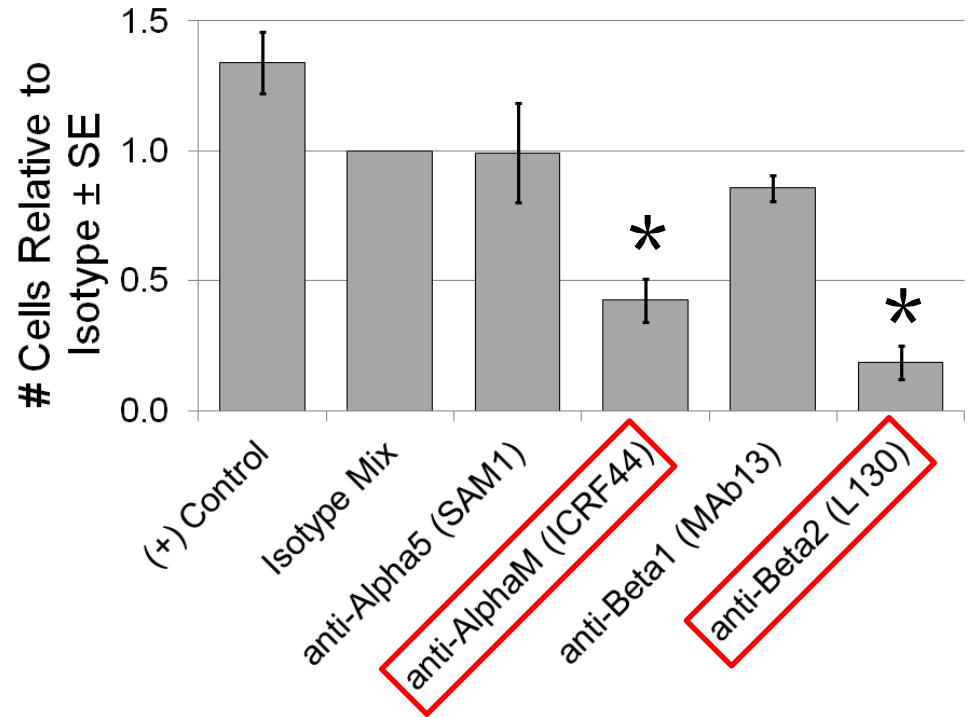
$$S = 3 \mu\text{m}/\text{min}$$

$$P = 15 \text{ min}$$

Hypothesis: integrins mediate adhesion



$\alpha_M\beta_2$ (Mac-1)

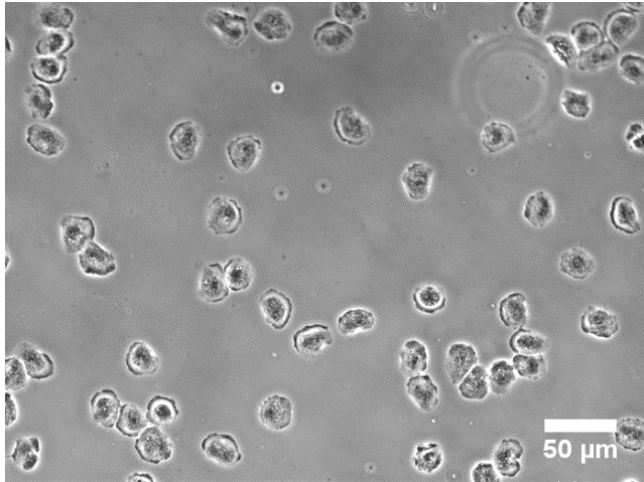


* p < 0.05, Dunnet's One Way ANOVA

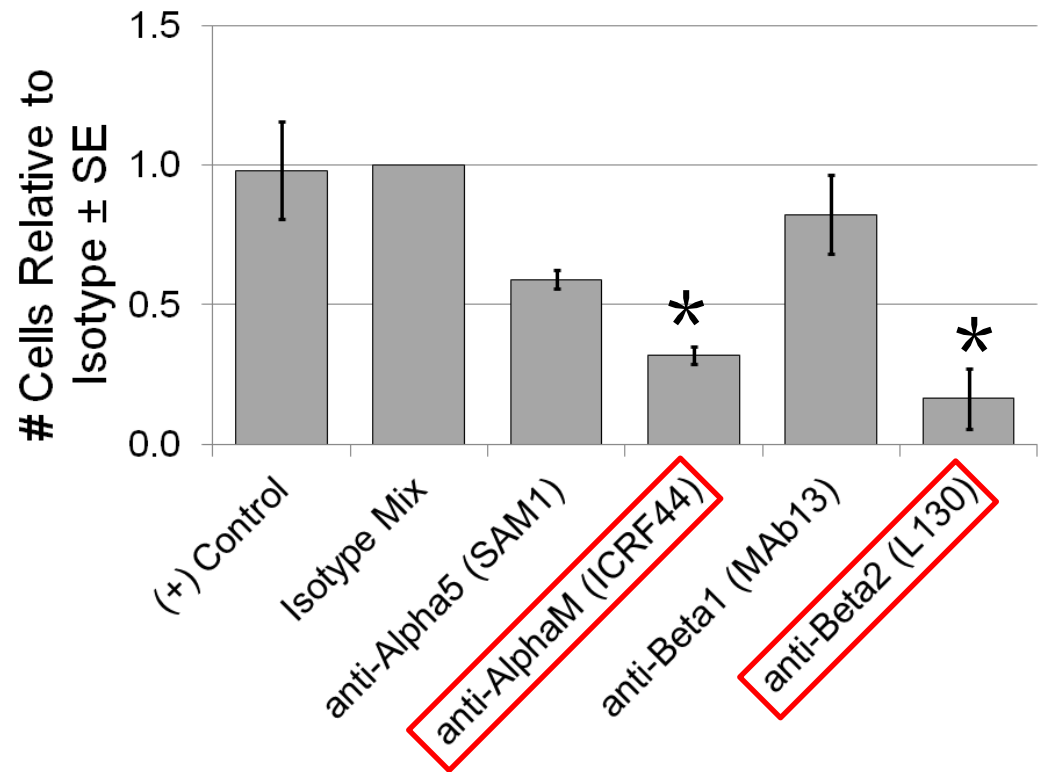
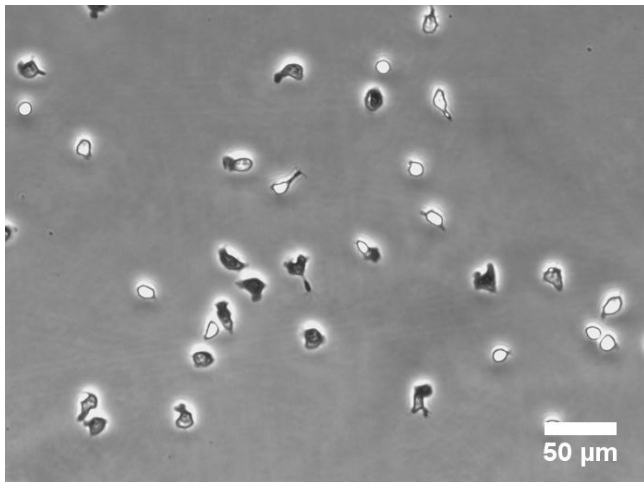
$\alpha_M\beta_2$ (Mac-1) is a promiscuous integrin

Hypothesis: density sensitivity is not FN specific

Intermediate density BSA ← Be careful about choice of “blocking” agent!



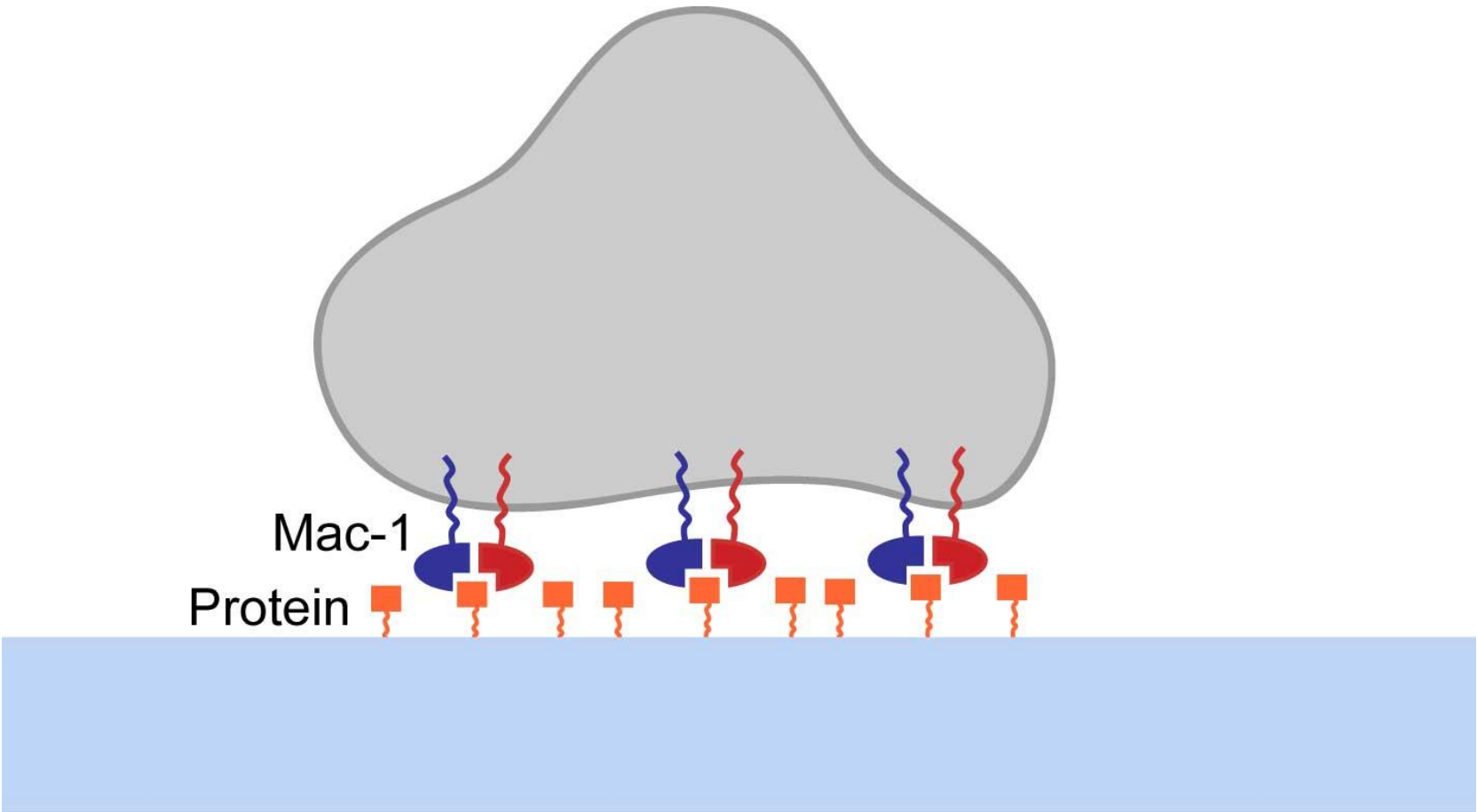
High density BSA



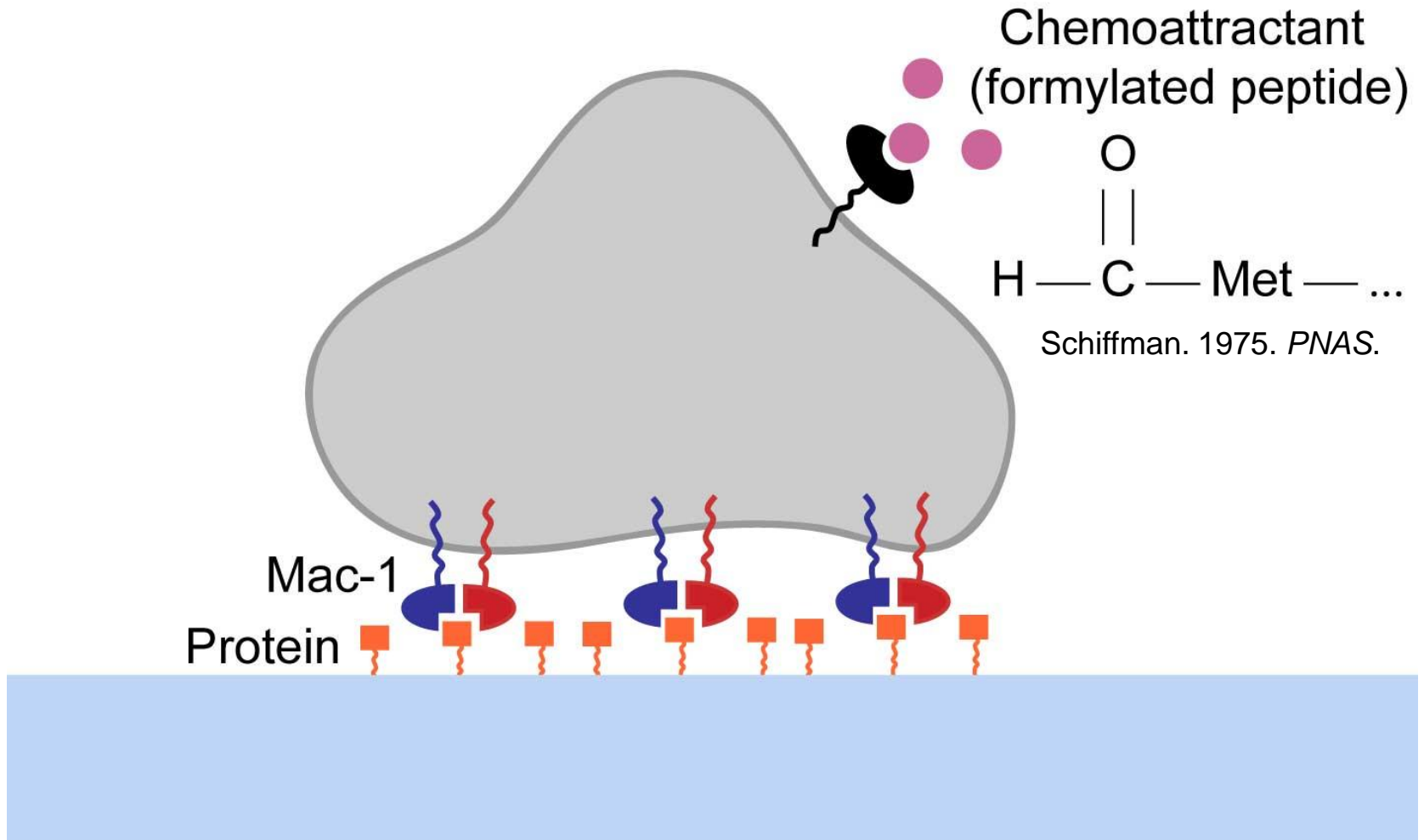
$\alpha_M\beta_2$ (Mac-1)

* p < 0.05, Dunnet's One Way ANOVA

So far, response to adhesive ligand alone (haptokinesis)



Response to adhesive ligand **and** chemoattractant?



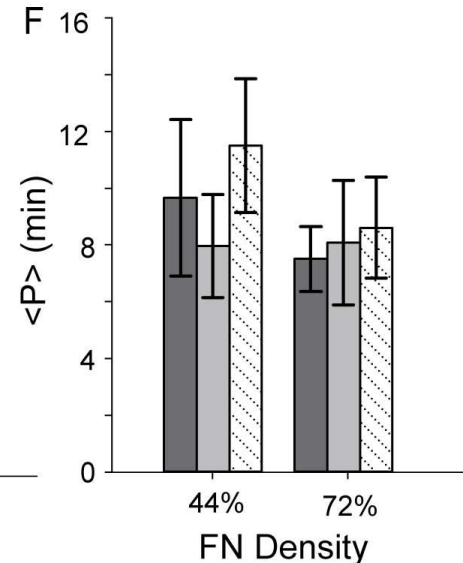
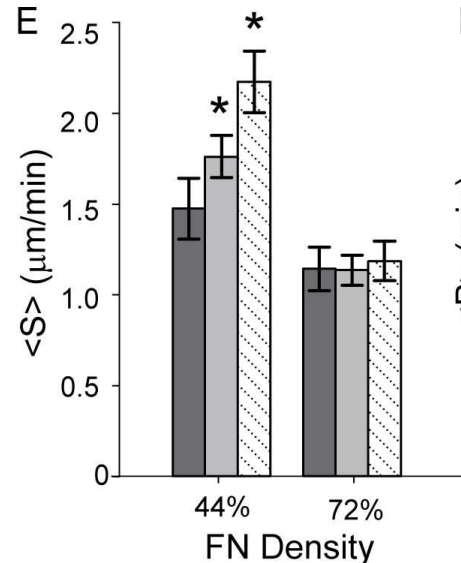
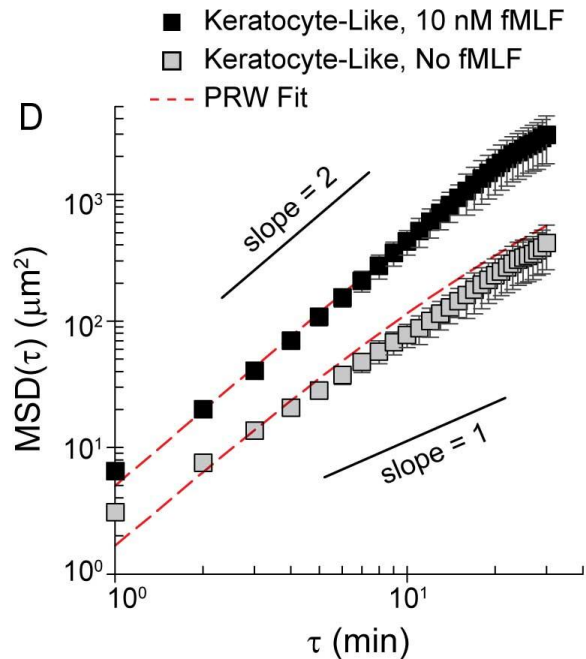
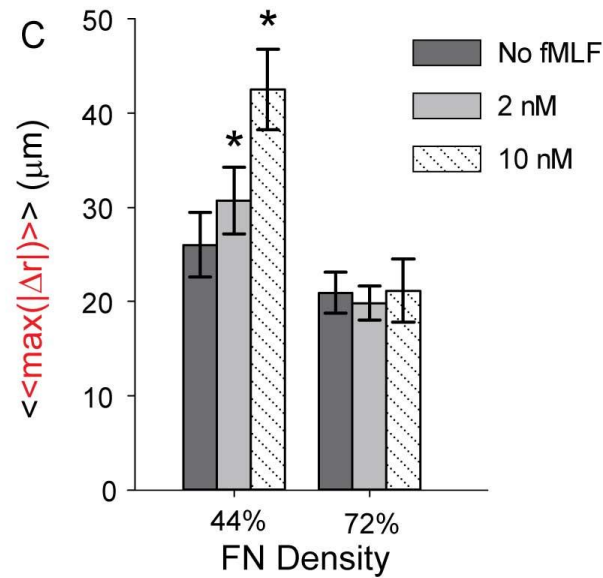
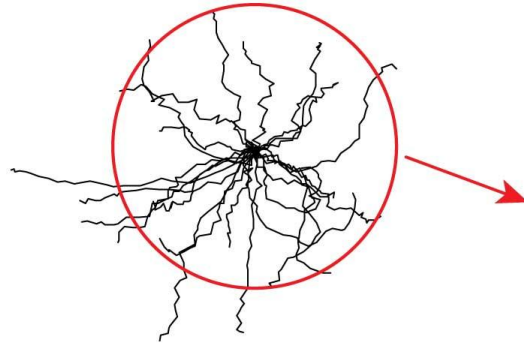
Haptokinesis (surface stim.) → chemokinesis (soluble stim.) of keratocyte-like phenotype

Haptokinesis

Chemokinesis

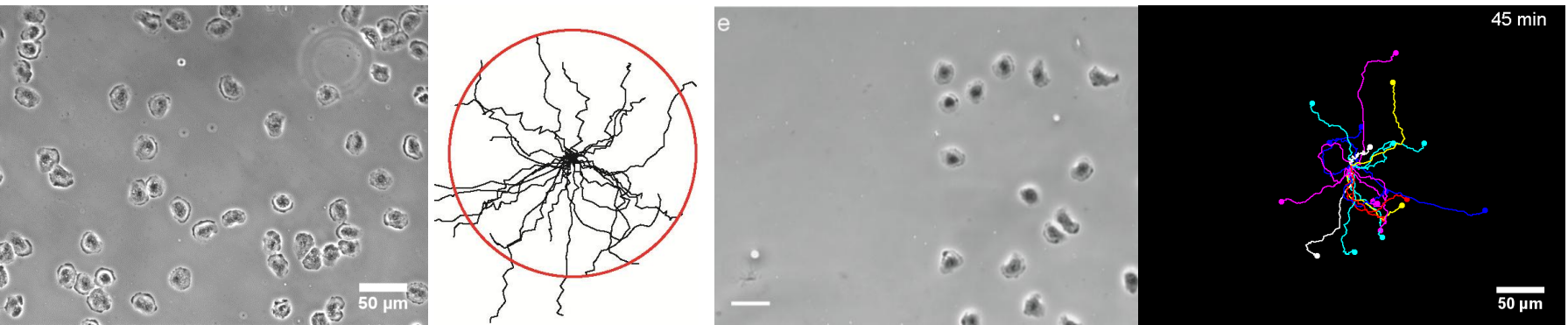


50 μm

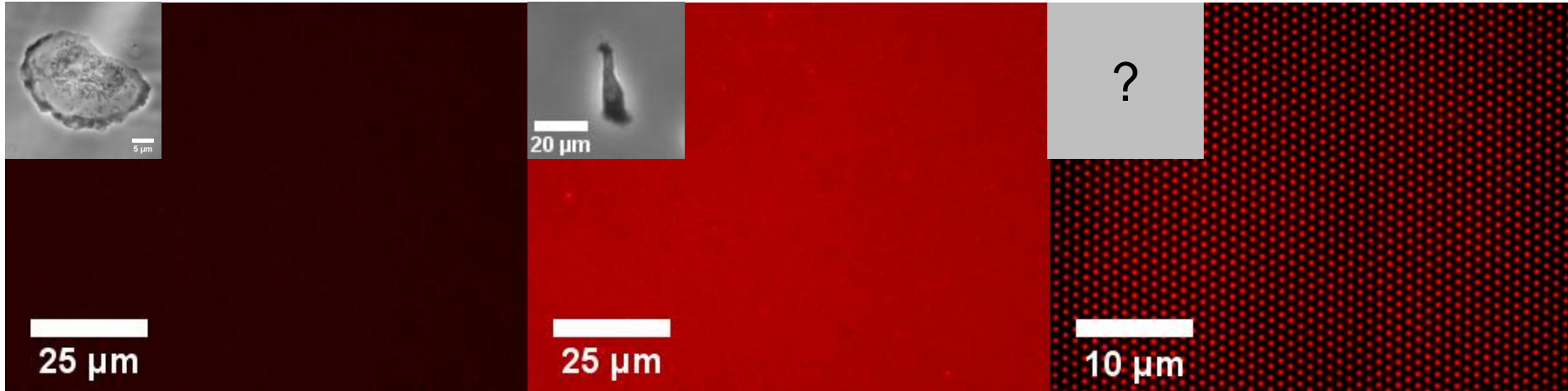


Neutrophils are capable of an adhesion-driven phenotypic switch with respect to shape and motility.

Promiscuous Mac-1 mediates this sensitivity.



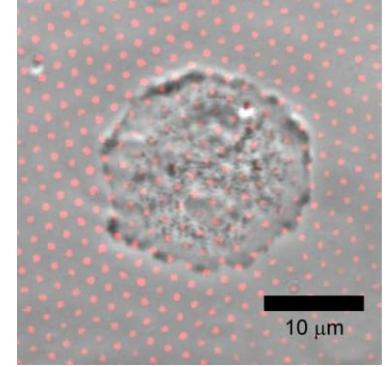
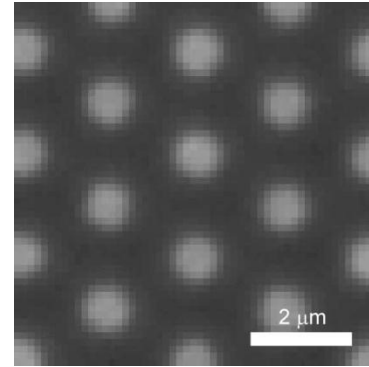
Length scale of density sensing?



Density Sensing

Dynamic traction forces of spreading and adherent human neutrophils

Henry, Crocker, Hammer. 2015. *ABME* (In Prep)



Aim:

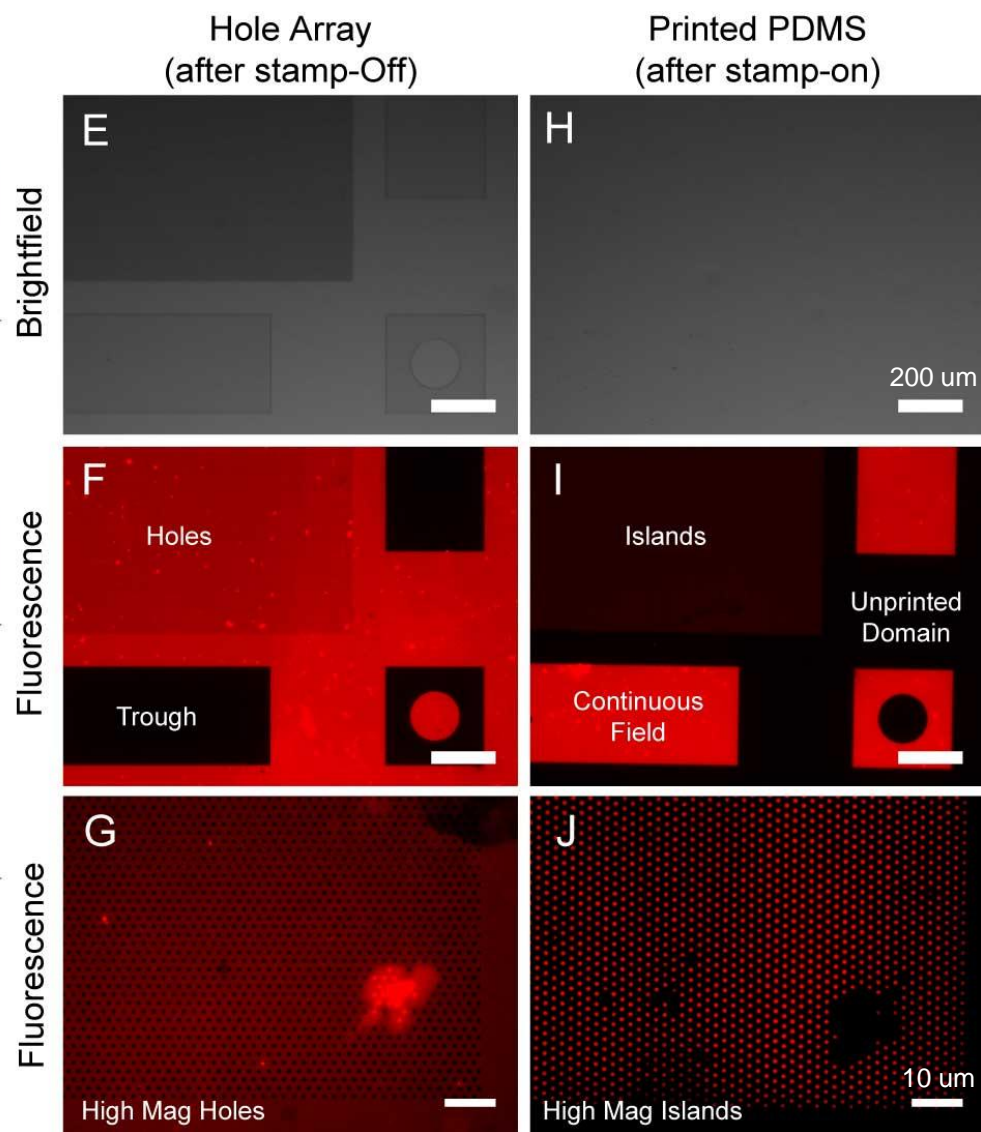
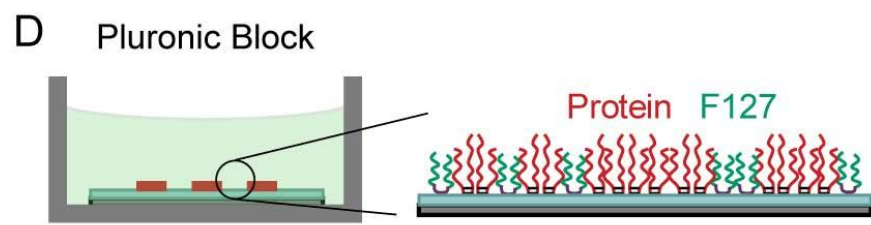
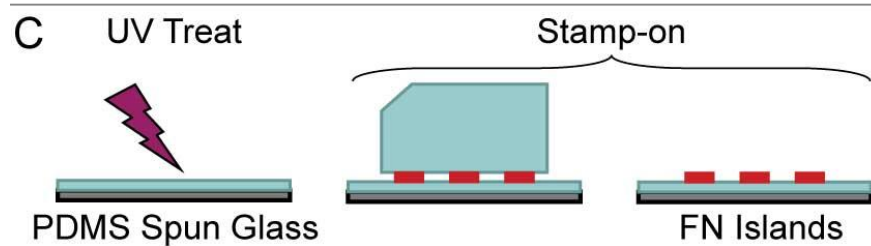
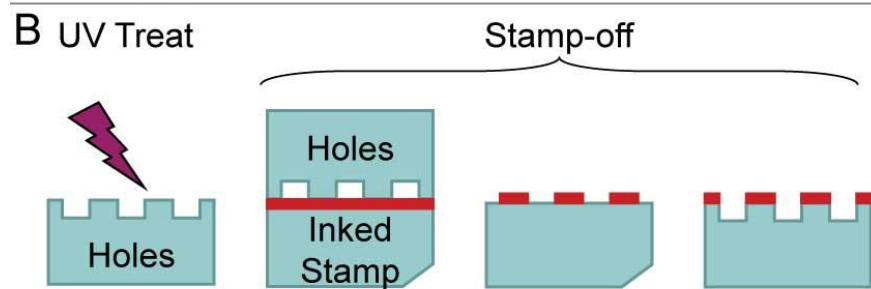
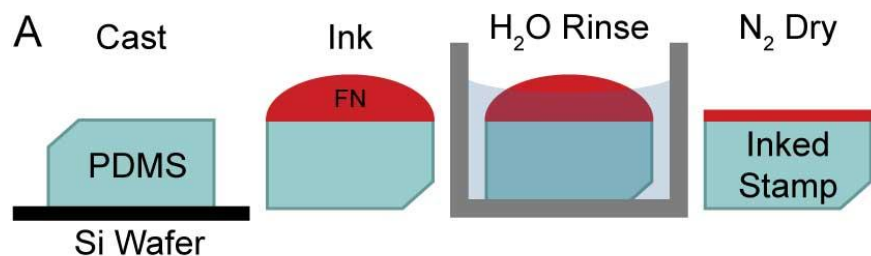
Elucidate length scale of density sensitivity

Hypotheses: (on dual adhesive environments)

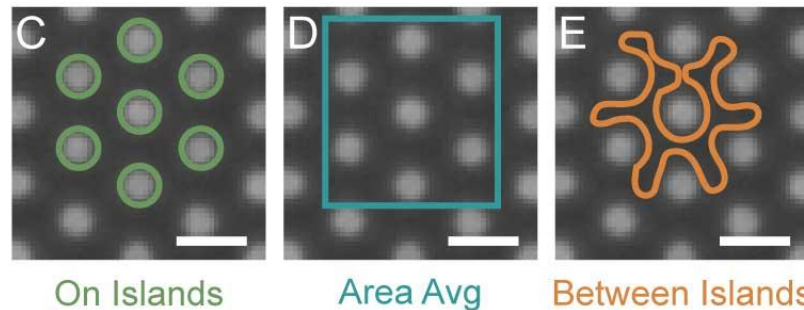
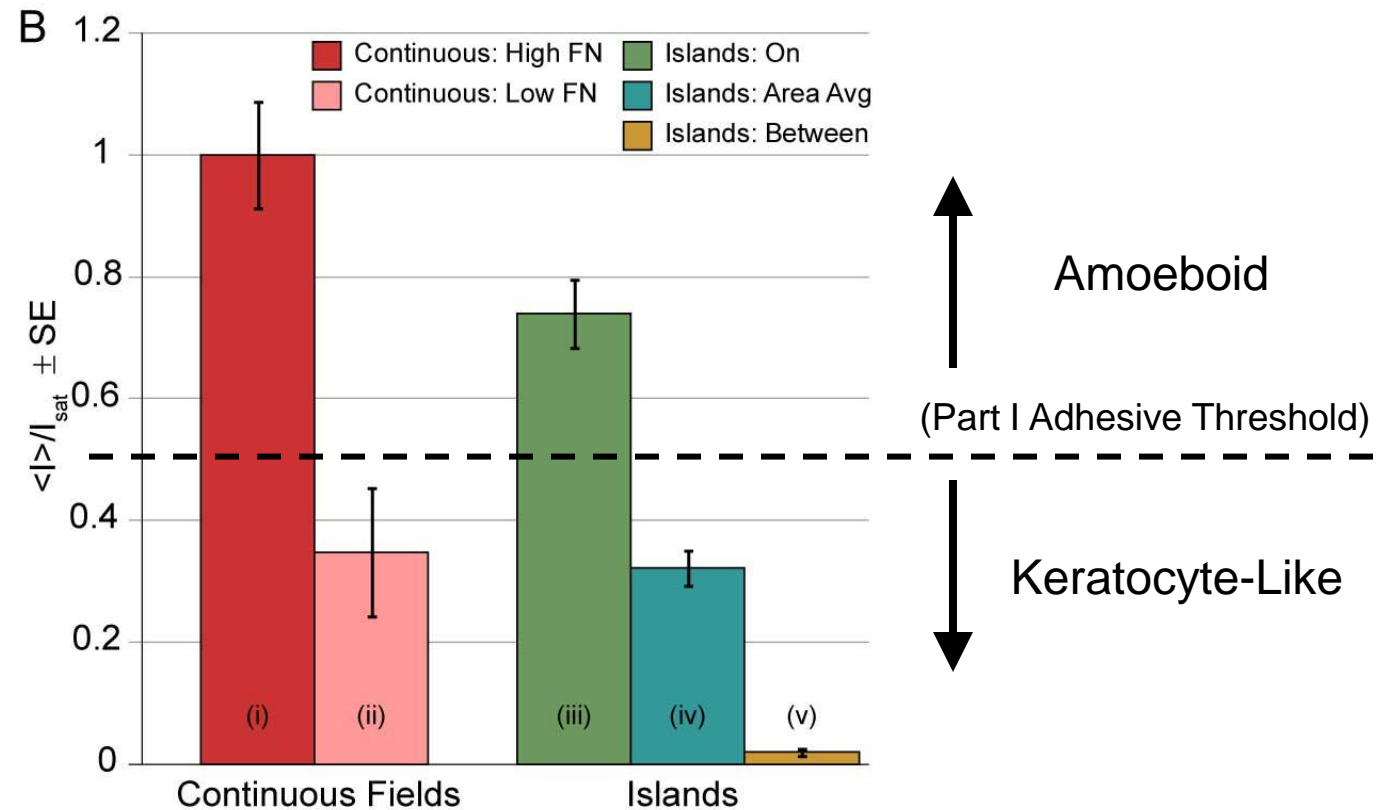
Local (submicron) sensitivity → amoeboid

Global (whole cell) sensitivity → keratocyte-like

Arrays of discrete islands via “stamp-off”



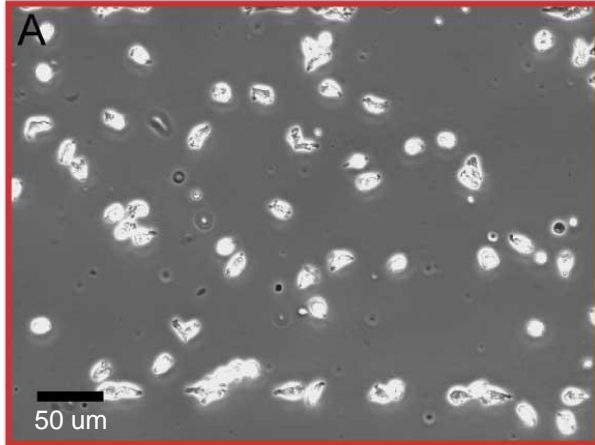
Engineering dual adhesive length scales



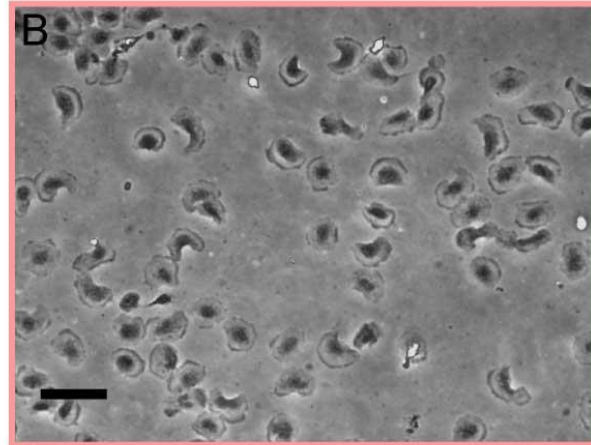
Neutrophil phenotype on islands?

Keratocyte-Like!

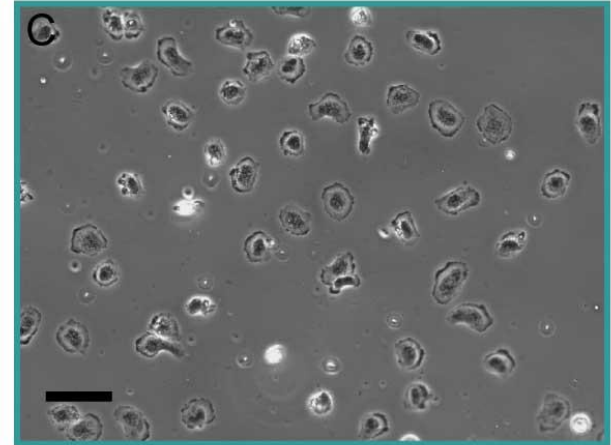
High Density Continuous Field



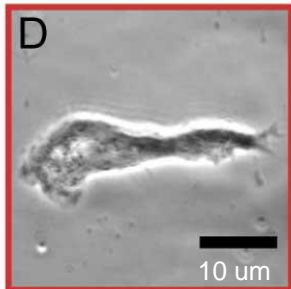
Low Density Continuous Field



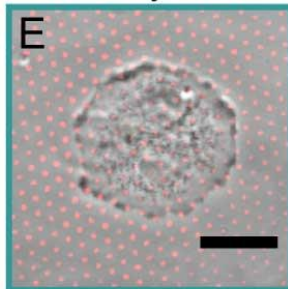
Islands



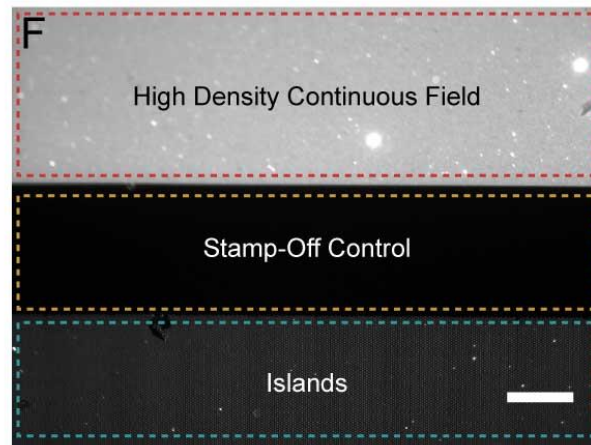
Amoeboid



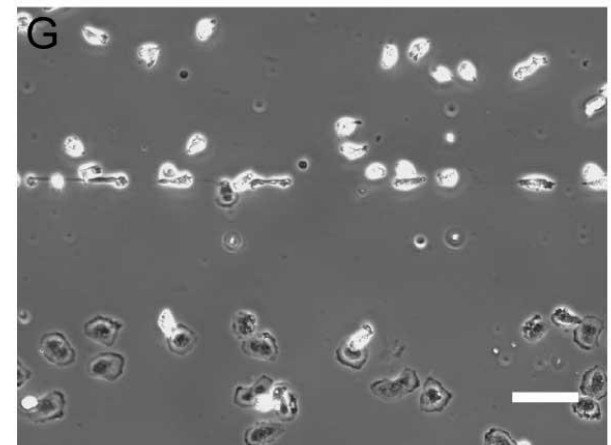
Keratocyte-Like



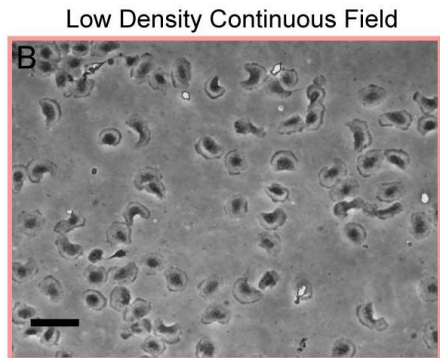
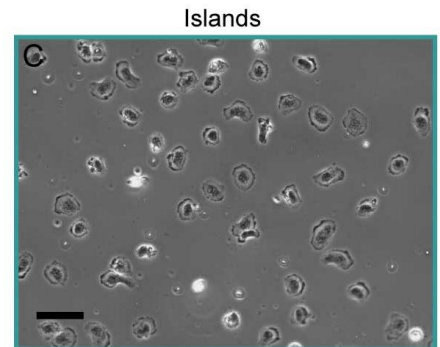
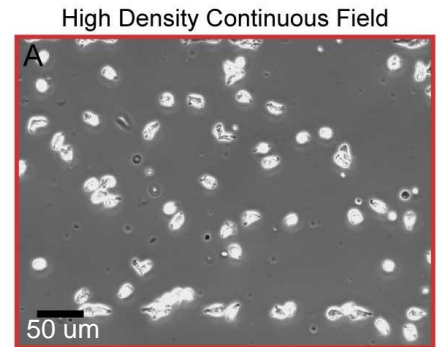
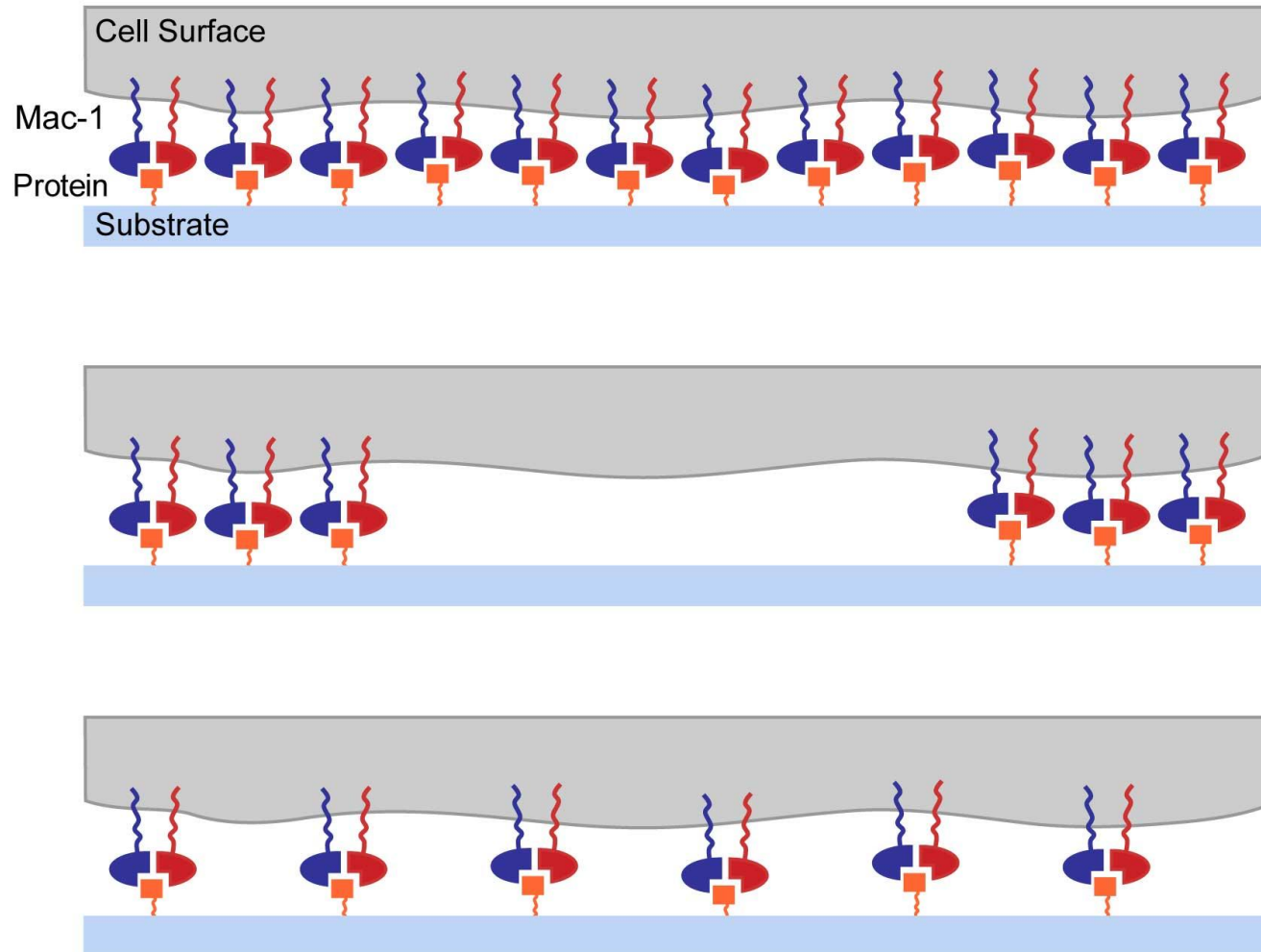
Fluorescence



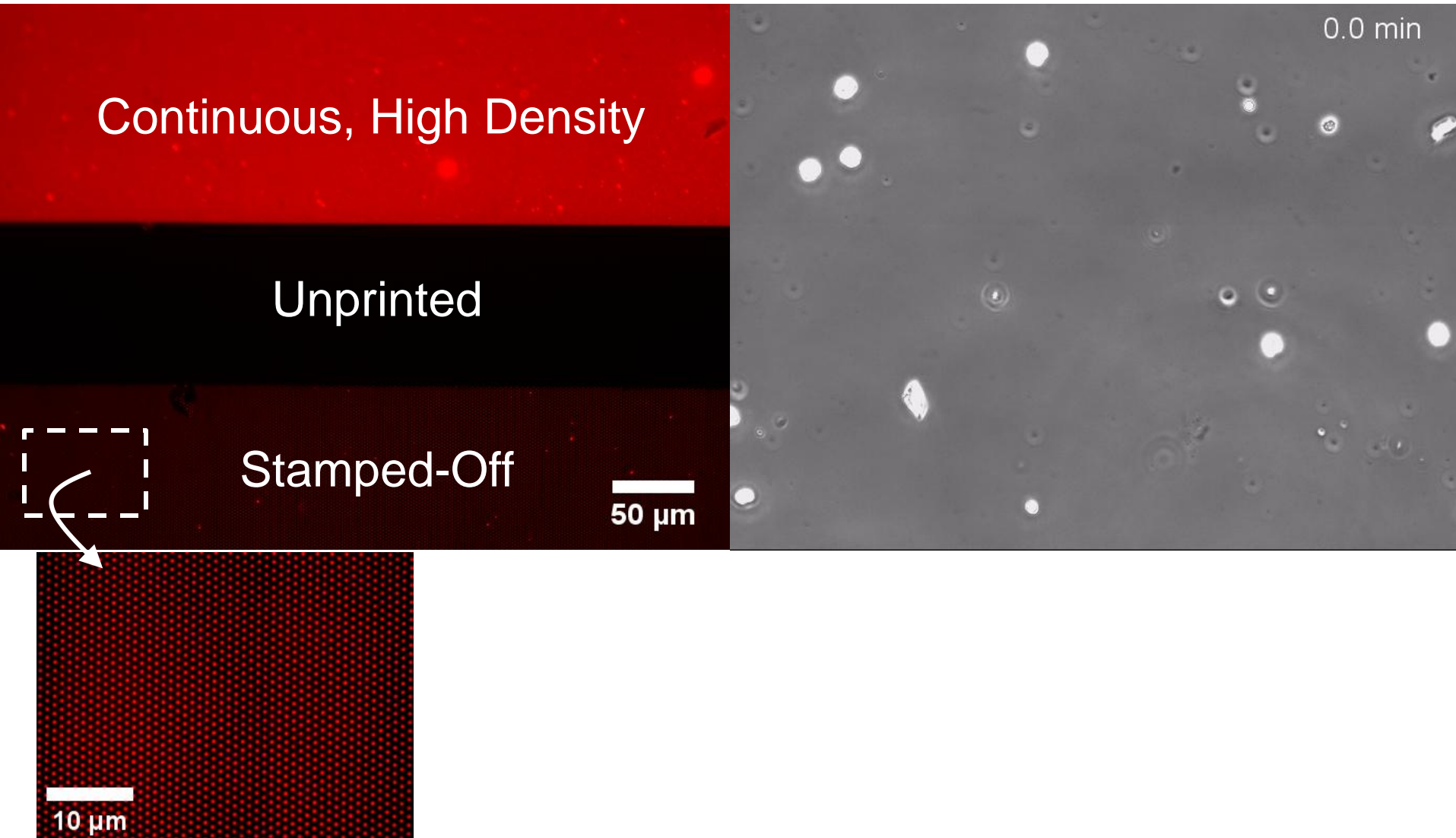
Phase Contrast



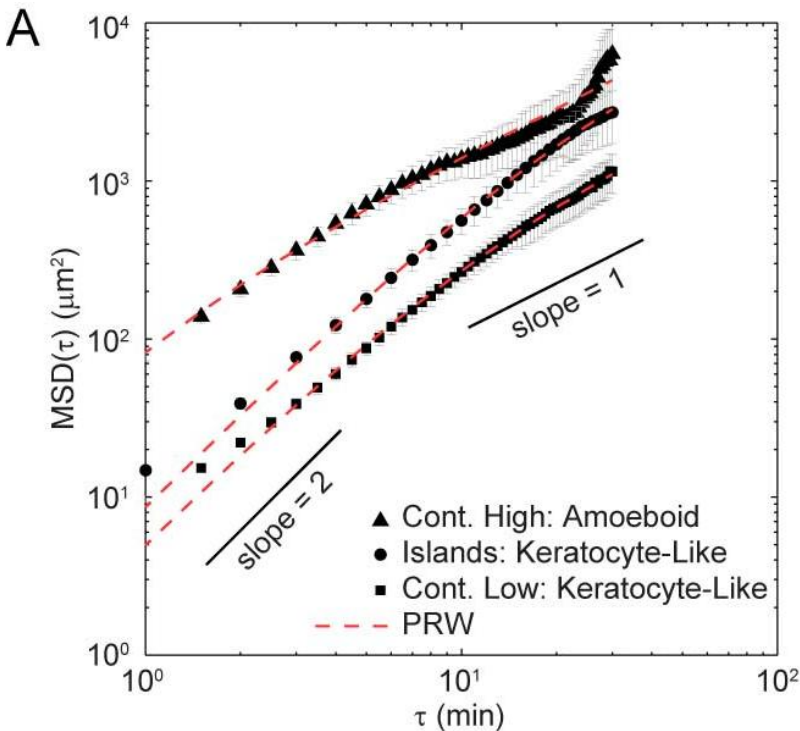
Neutrophils **integrate** adhesive stimulation



Neutrophils integrate adhesive stimulation Rapid amoeboid \rightarrow keratocyte-like transitions

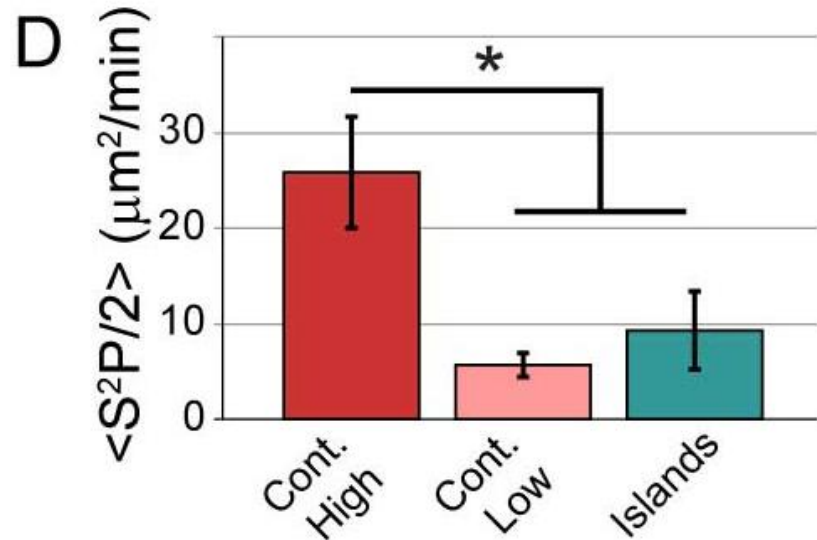


Motility on islands \approx moderate adhesivity continuous field



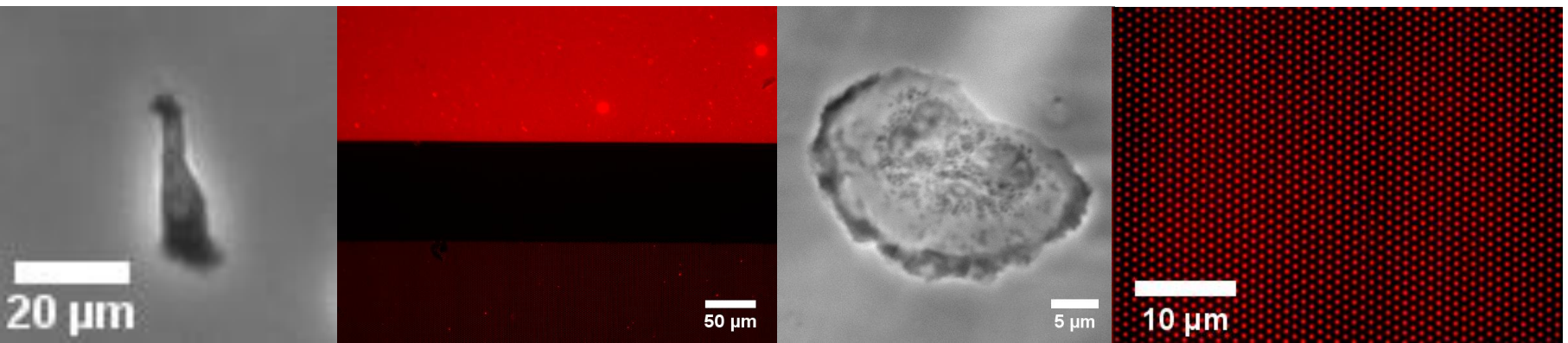
$$\langle \Delta r^2(\tau) \rangle = 2S^2P[\tau - P(1 - \exp(-\tau/P))]$$

random motility coeff. = $S^2P/2$



Part II Summary

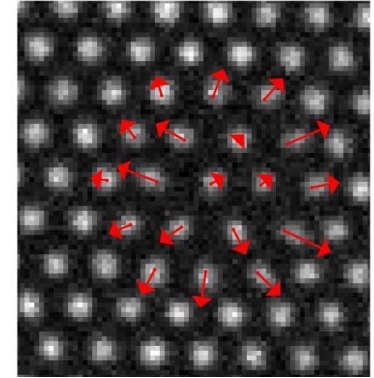
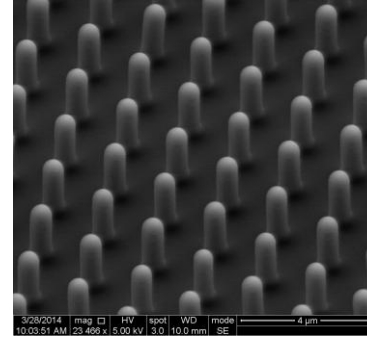
Neutrophils integrate local (submicron) adhesive stimuli and coordinate a global (whole cell) phenotypic response.



Spreading Mechanics

Dynamic traction forces of spreading and adherent human neutrophils

Henry, Chen, Crocker, Hammer. 2015. *Biophys J.*
(Under Revision)



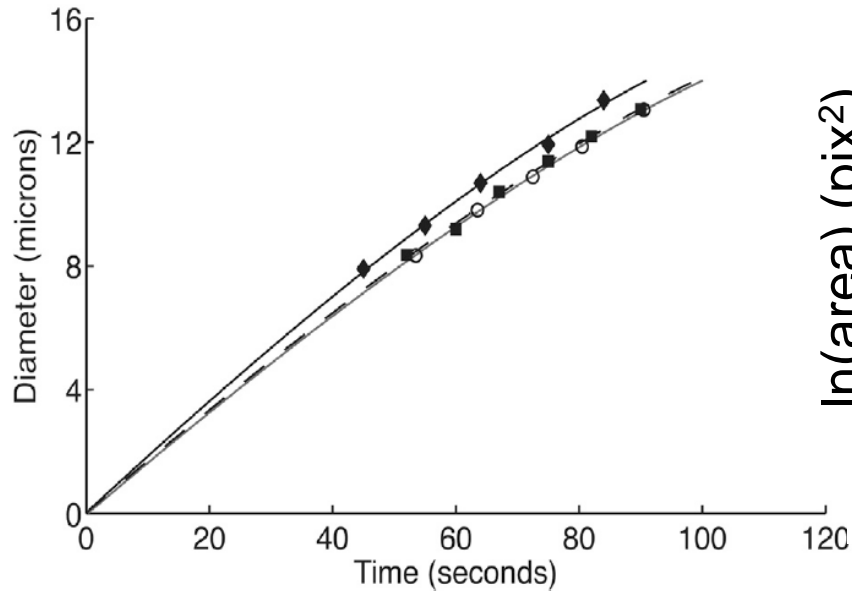
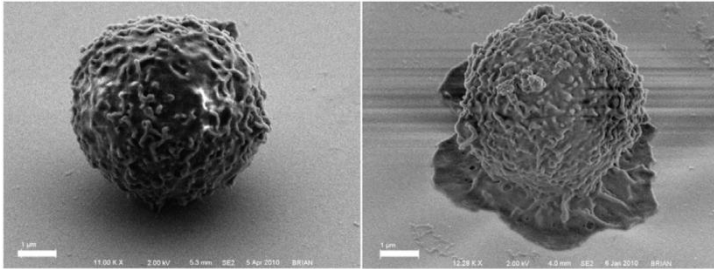
Aim:

Measure forces of adhesion-driven spreading

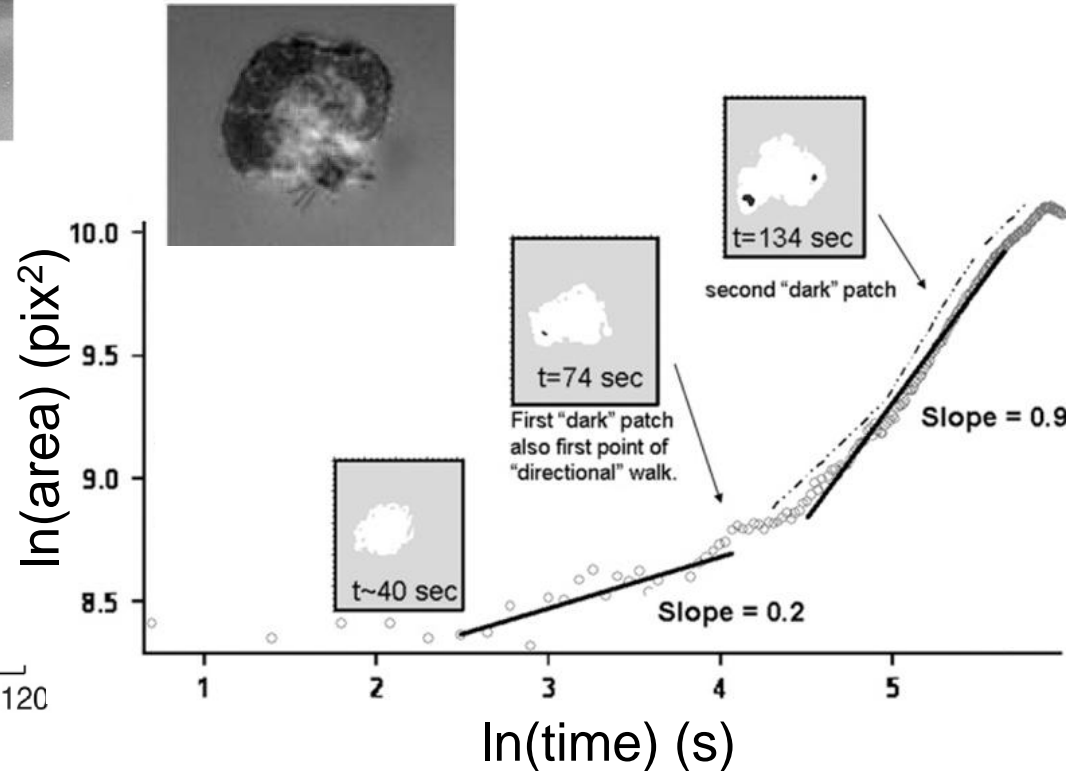
Hypothesis:

Spreading is an active process analogous to lamellipodium formation

Neutrophil spreading is fast. Can we measure the associated forces?

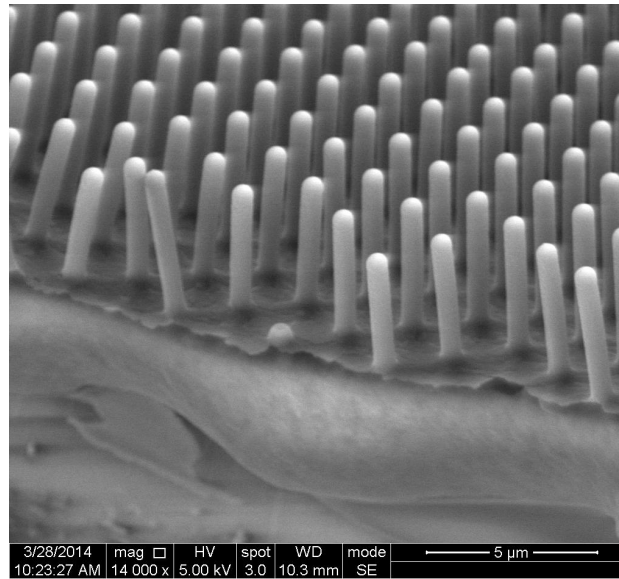
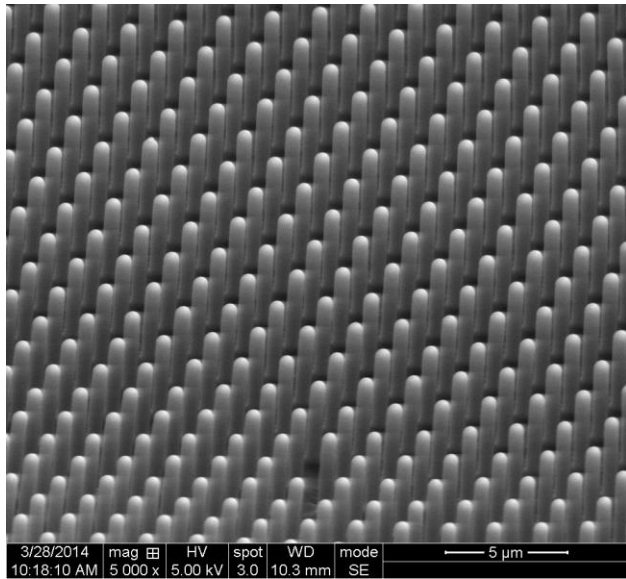


Lomakina et al. 2014. *Biophys J.*



Sengupta et al. 2006. *Biophys J.*

mPADs (microfabricated Post-Array-Detectors):



$$k_{\text{spring}} = 0.28 \pm 0.07 \text{ nN/um}$$

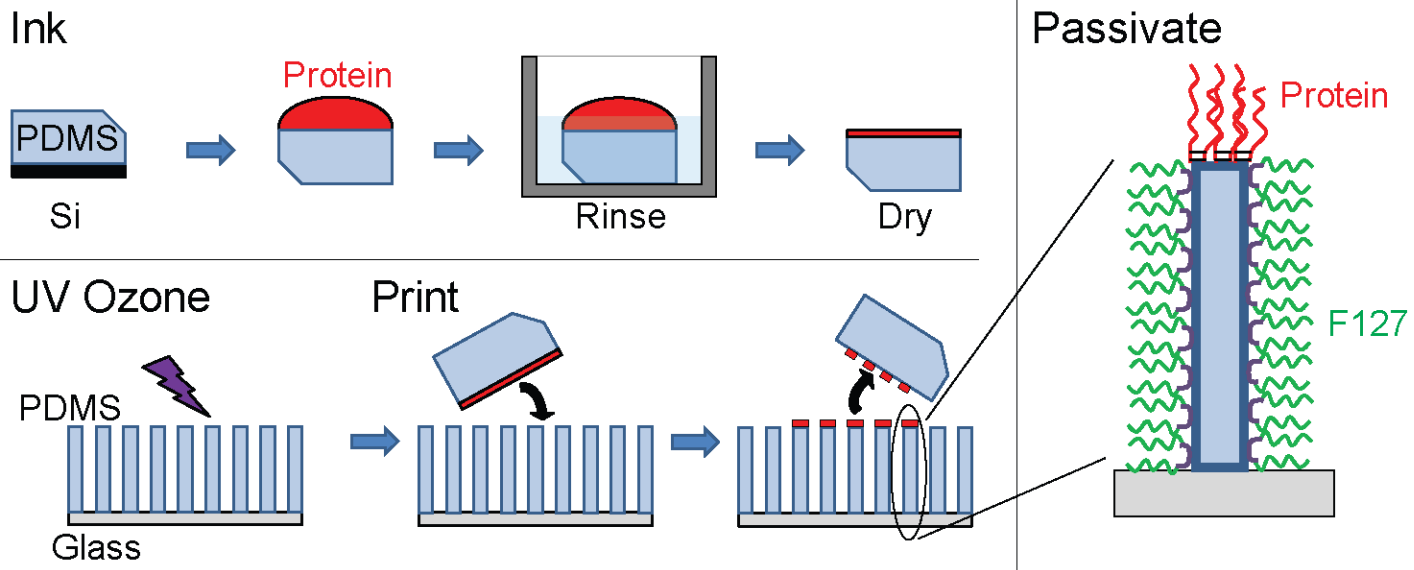
$$G \sim 5 \text{ kPa}$$

Schoen correction = 0.93

$$k_{\text{spring}}^* = (0.93)(k_{\text{spring}})$$

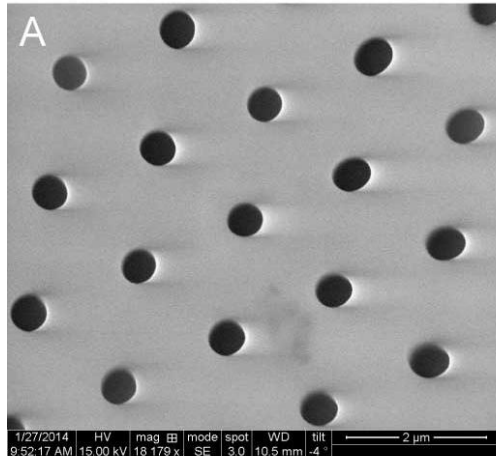
$$k_{\text{spring}}^* = 0.26 \text{ nN/um}$$

Schoen et al. 2010. *NanoLett.*

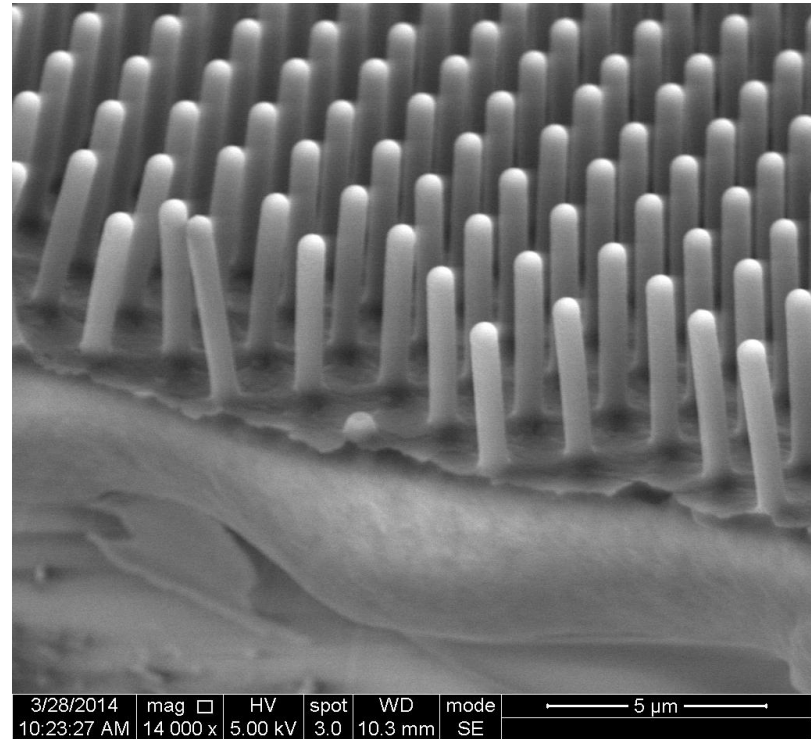
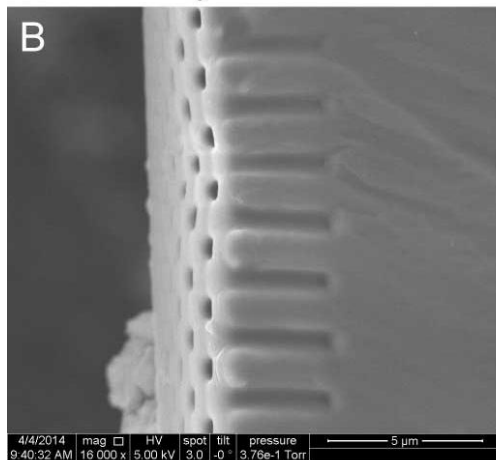


Array geometry preserved from Part II

Hole Arrays: Plan View



Hole Arrays: Cross-Section



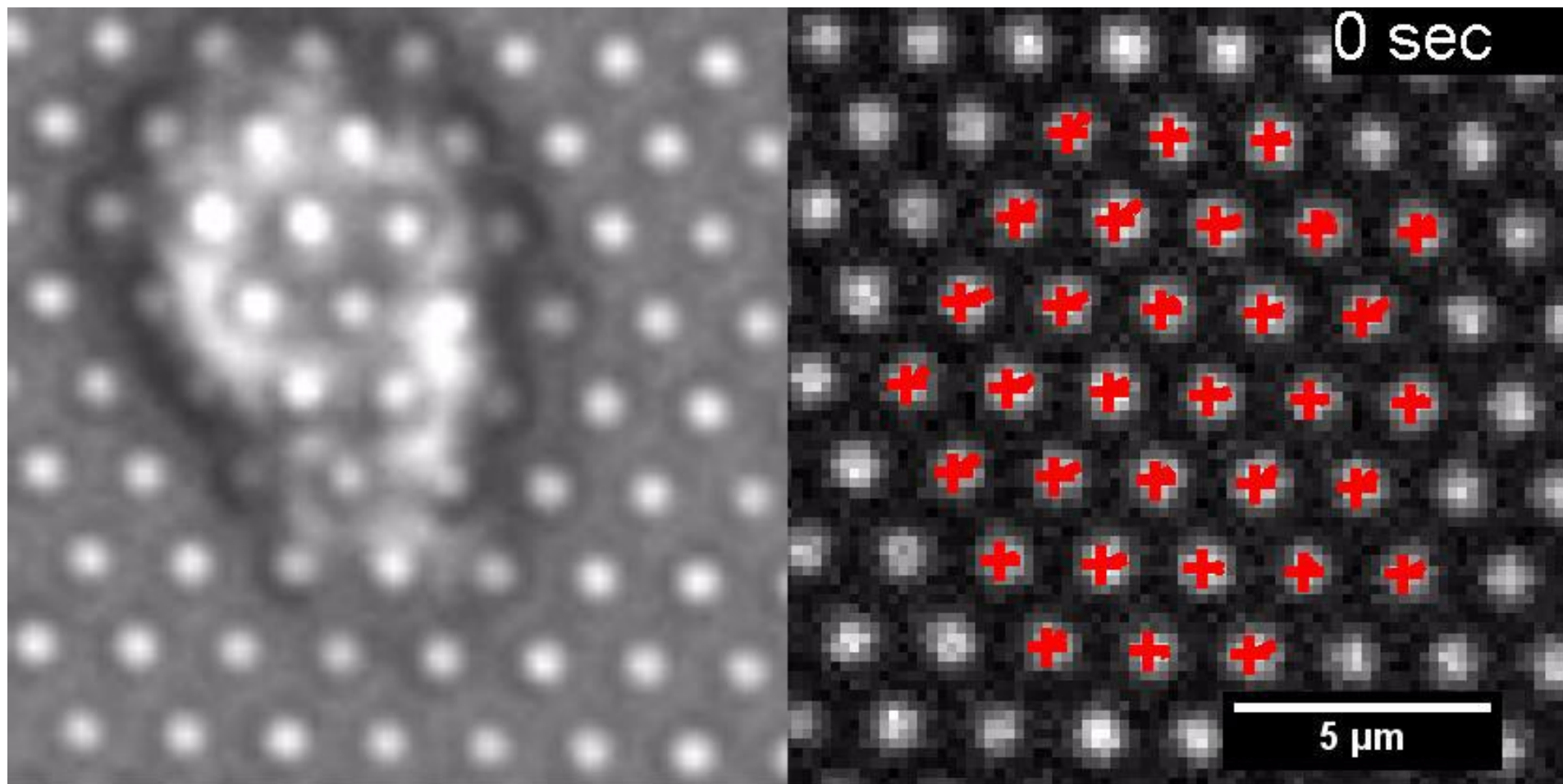
Henry et al. 2015. *Biophys J.* (Under Revision.)

Henry et al. 2015. *ABME.* (In Prep)

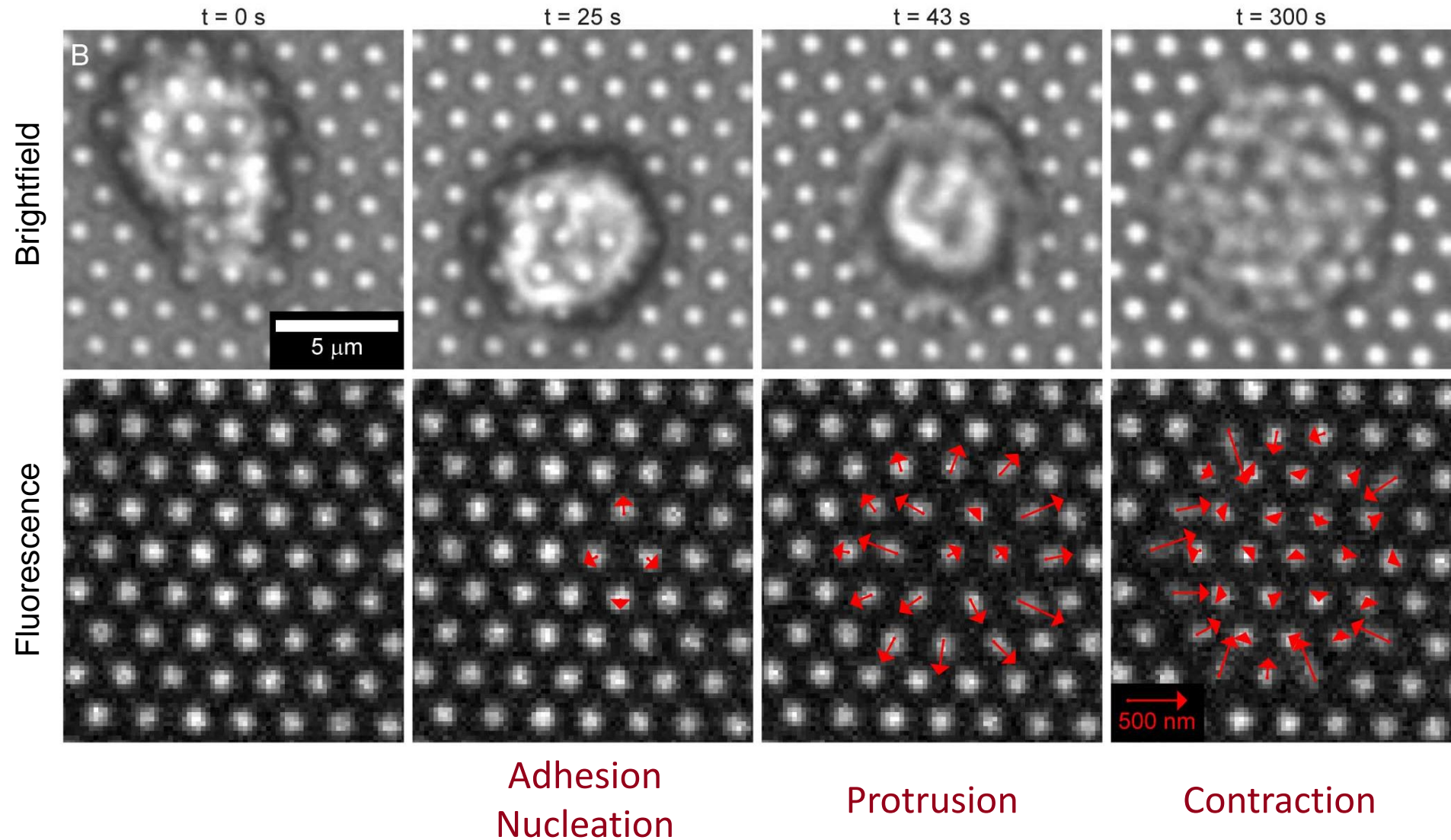
Neutrophil spreading on mPADs: raw data



Neutrophil spreading on mPADs: force annotation

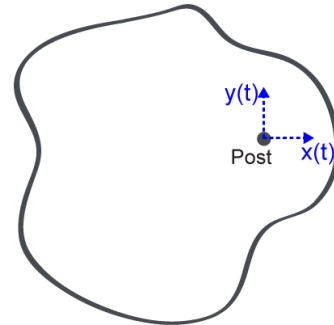


Neutrophil spreading on mPADs

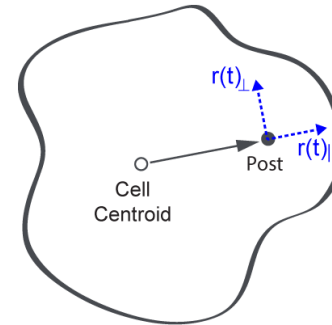


Plotting force trajectories in the cell reference frame

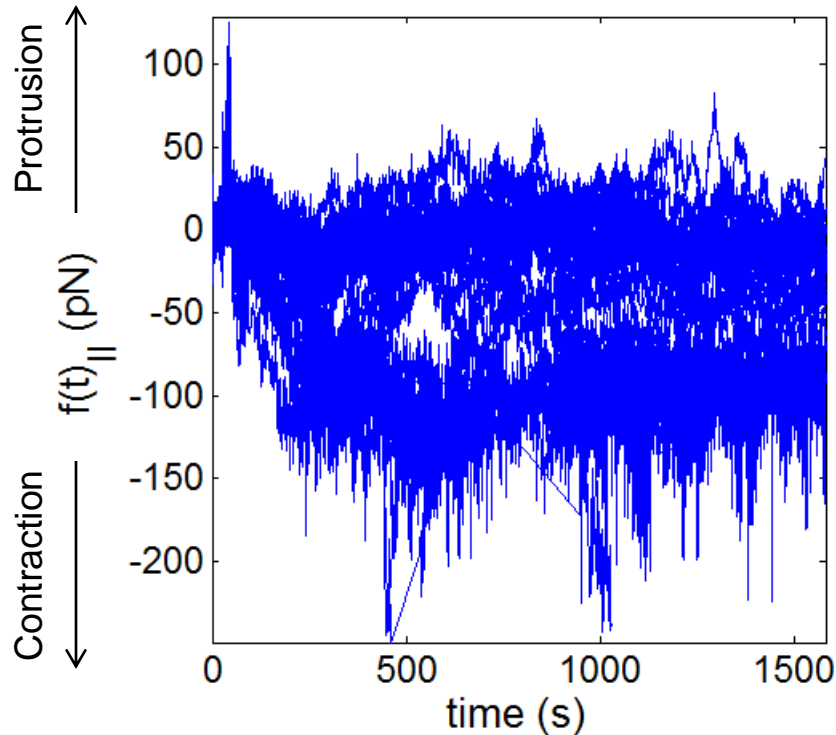
Lab Reference Frame



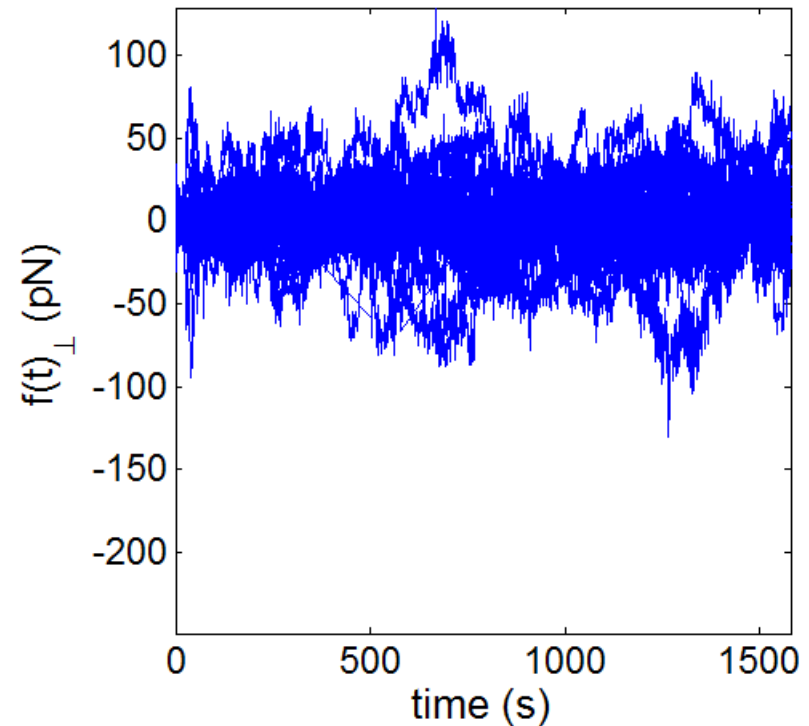
Cell Reference Frame



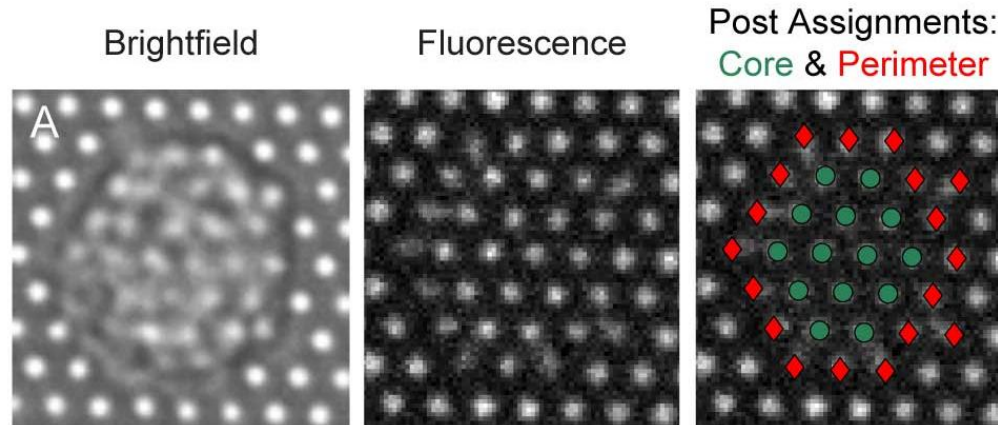
Radial



Tangential

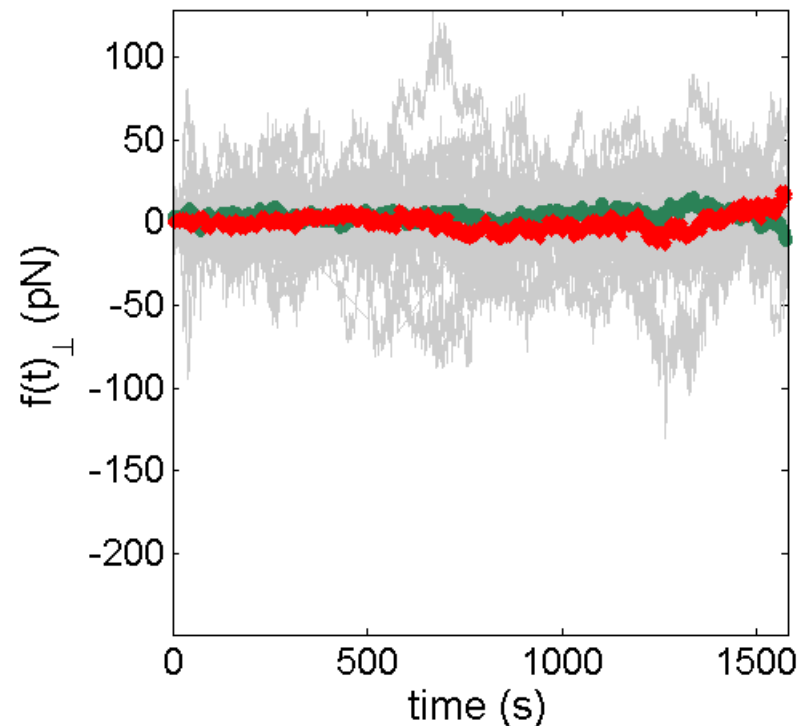
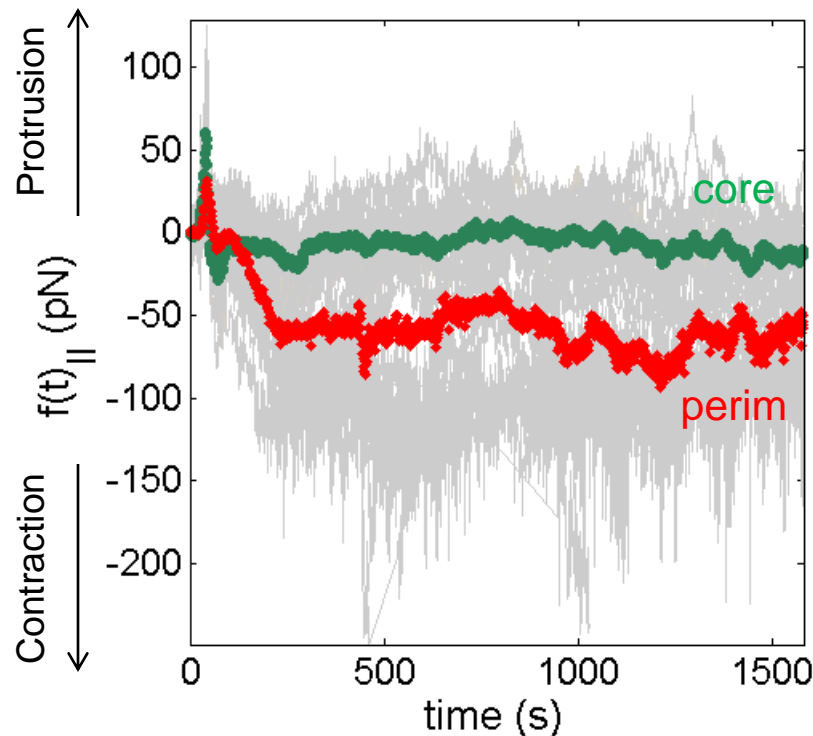


Dichotomizing data on geometric location



Radial

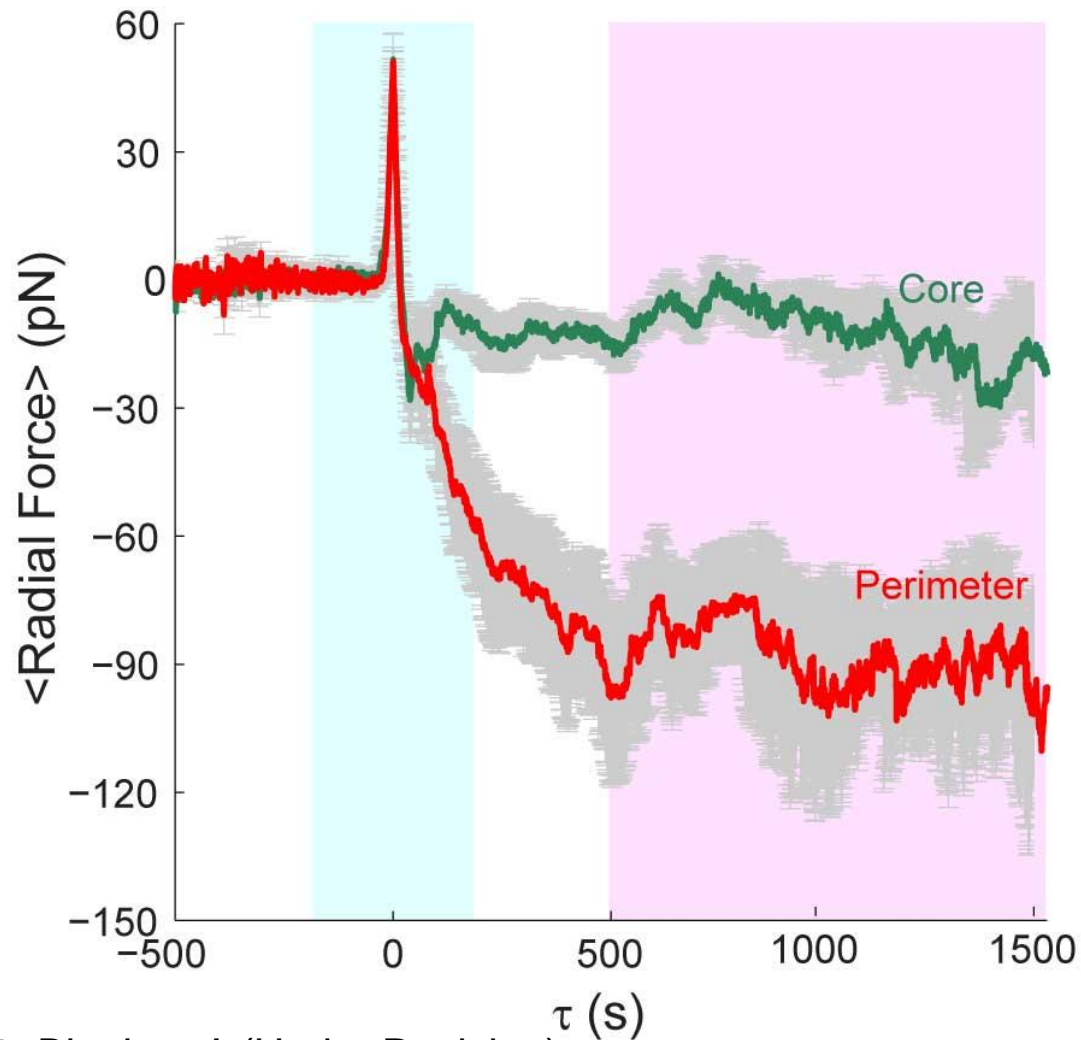
Tangential



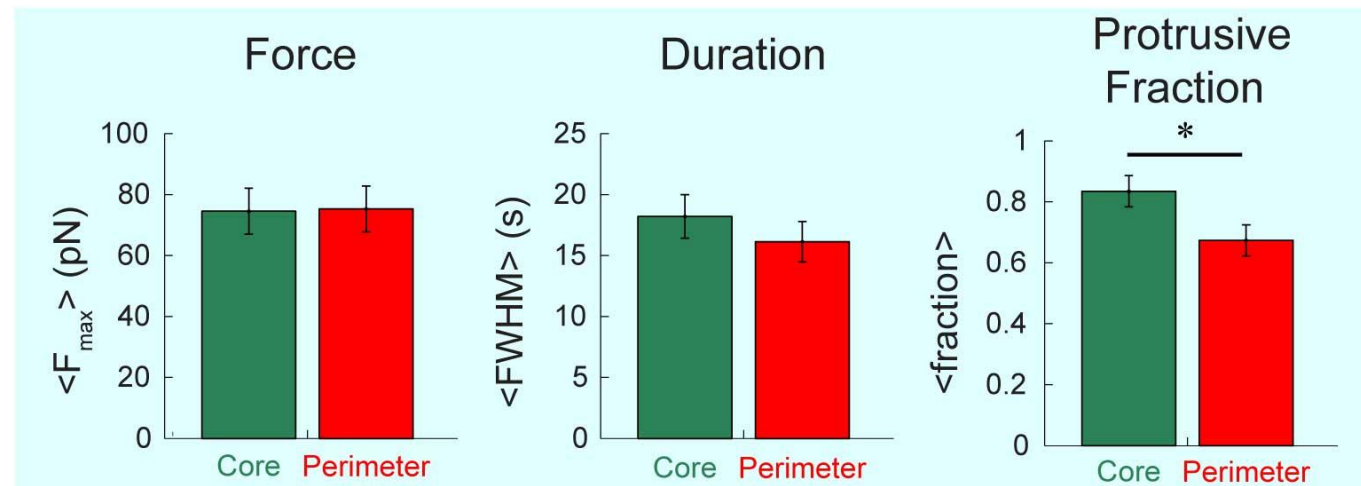
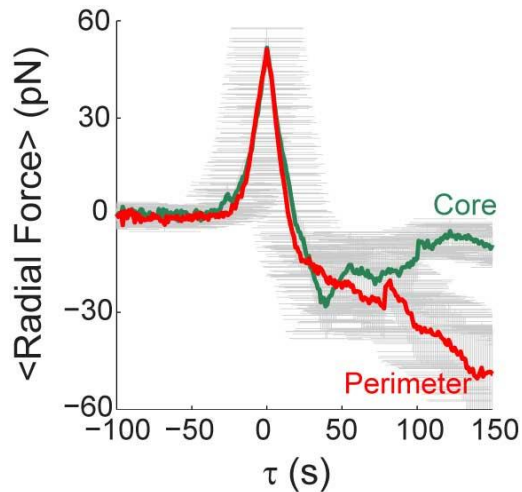
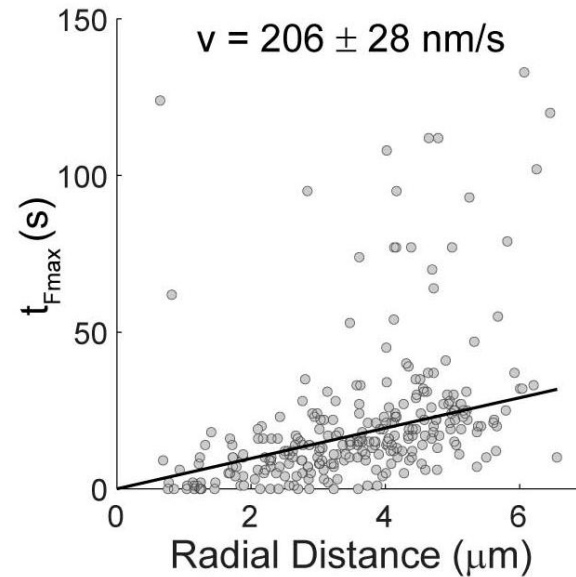
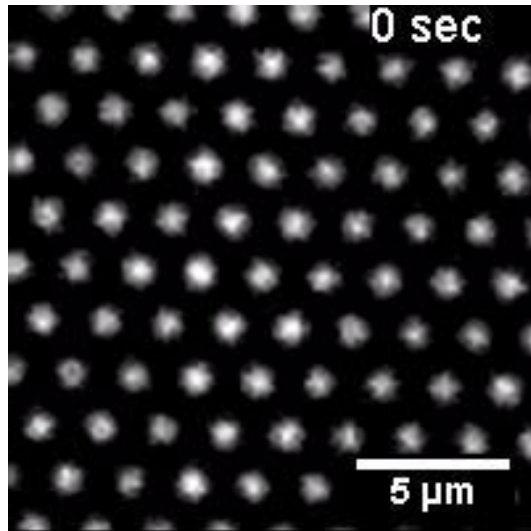
Ensemble avg makes mechanical regimes apparent

Transient Protrusion

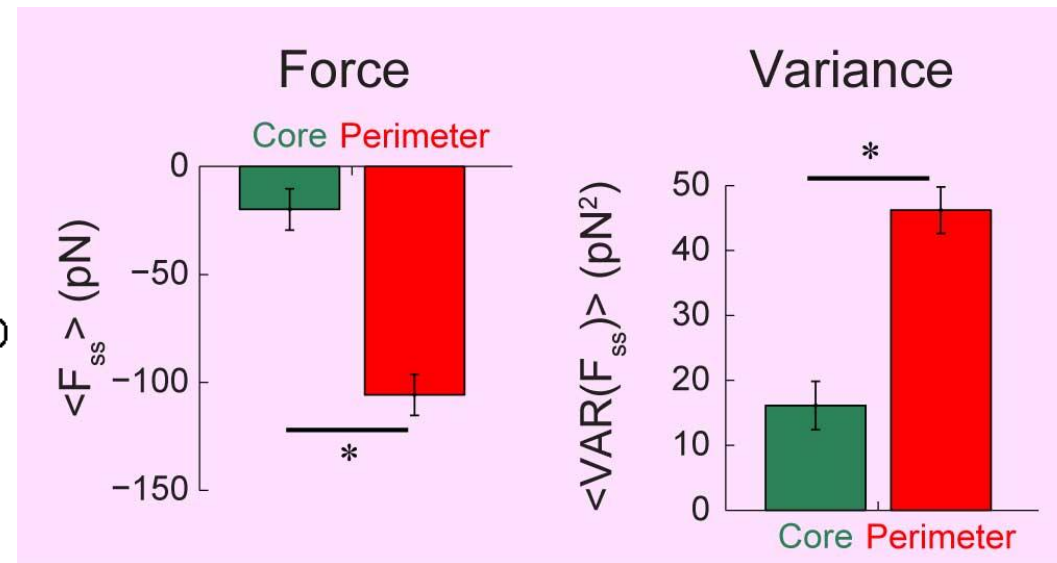
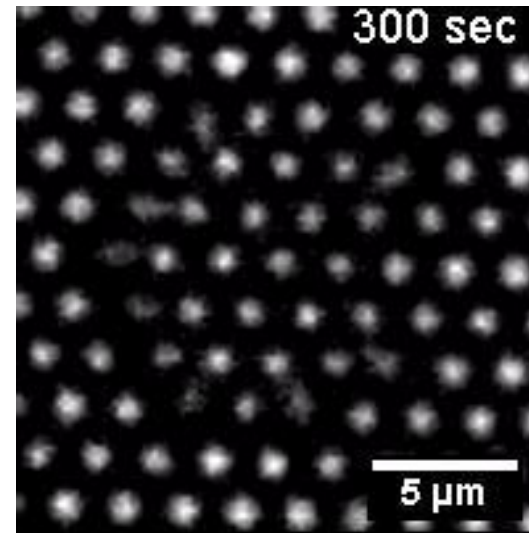
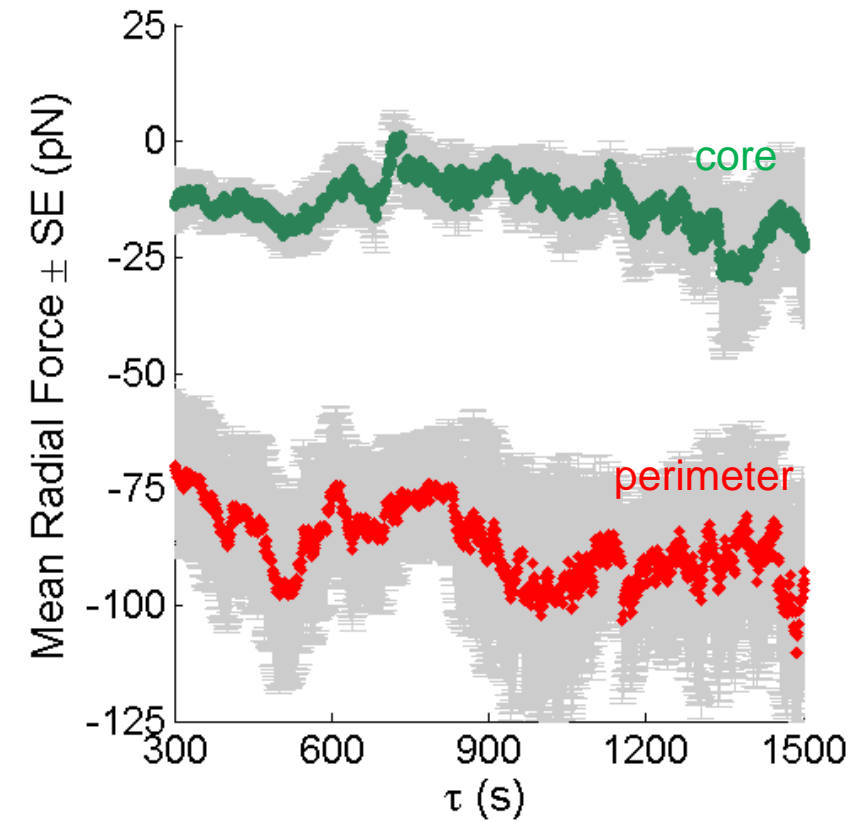
Steady State Contraction



Characterizing the protrusive wave



Characterizing the Steady State Contractile Regime



Are protrusion and contraction biochemically distinct?

Hypothesis: Contraction is RhoA/Rock and Myosin Mediated

Hypothesis: Protrusion is lamellipodium formation

Y27632 (1 μ M)

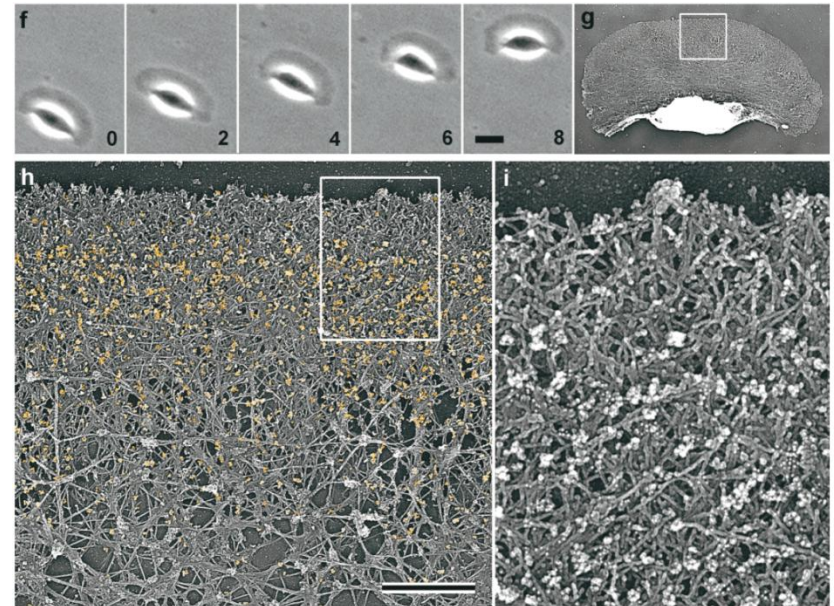
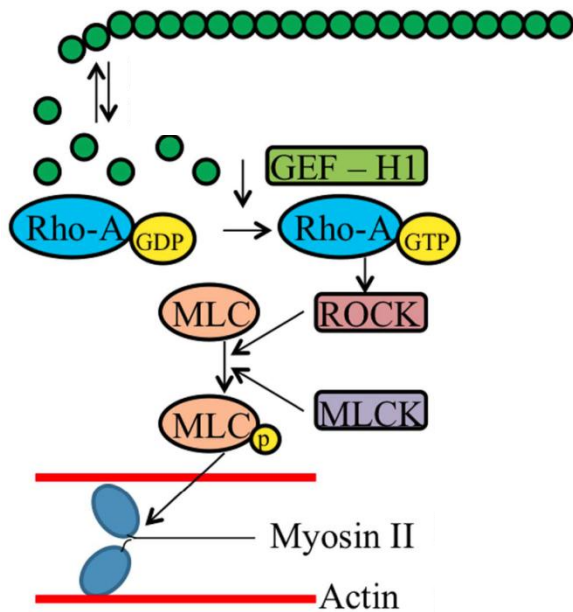
Blebbistatin (5 μ M)

CK666 (1 μ M)

p160ROCK

NM Myosin II

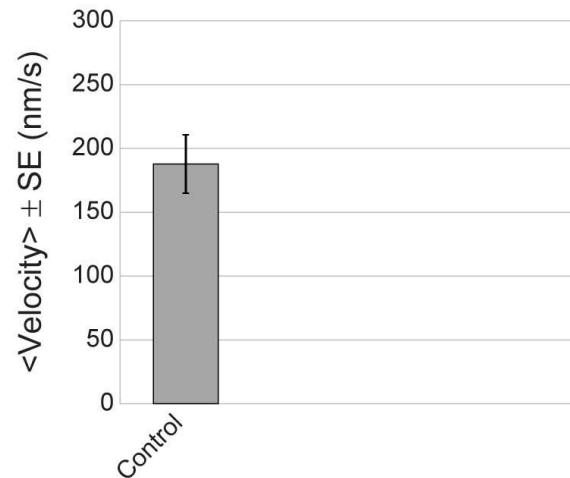
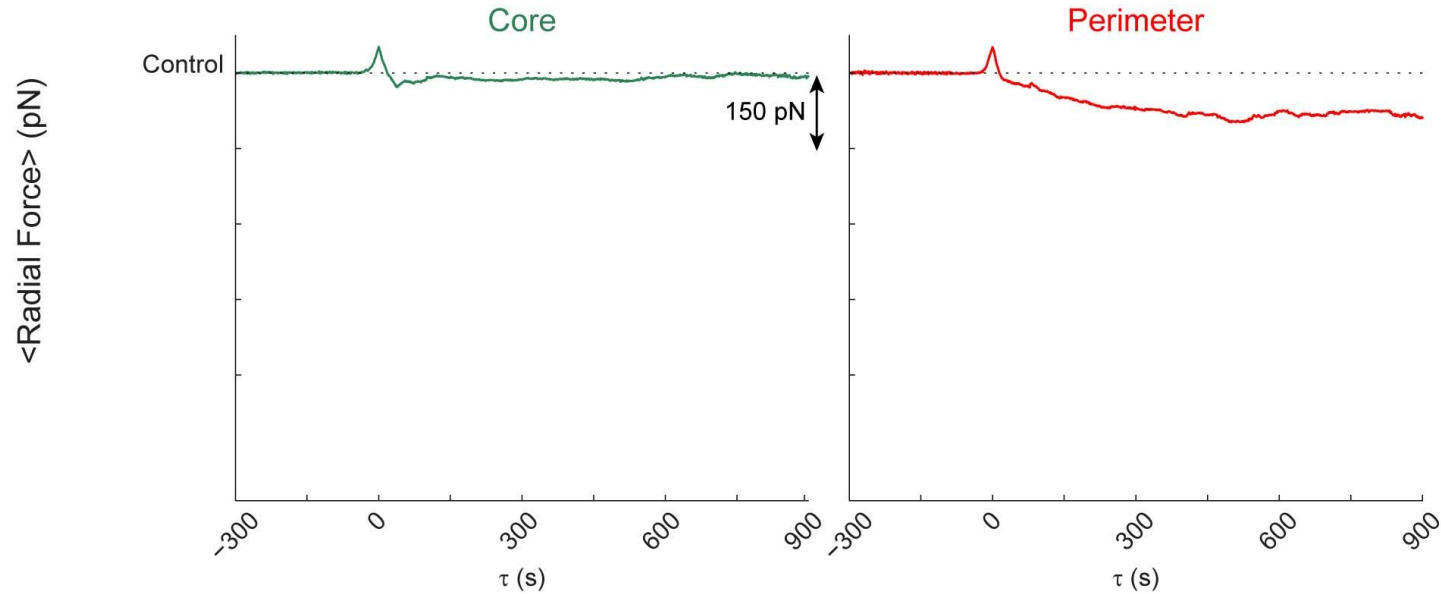
Arp2/3



Modified from Stroka. 2013. *PLOS ONE*.

Svitkina. 1999. *JCB*.

Looking for inhibitor effects



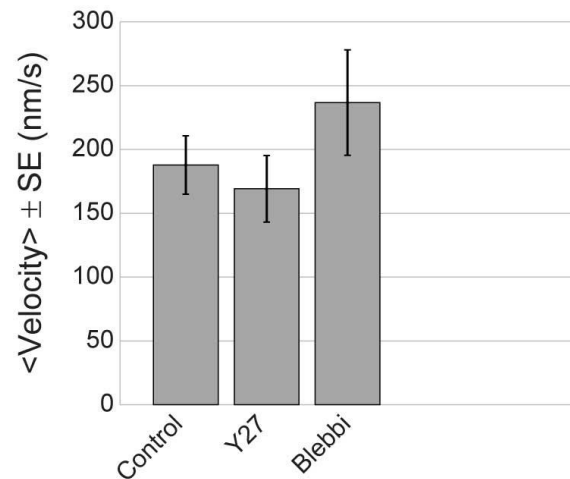
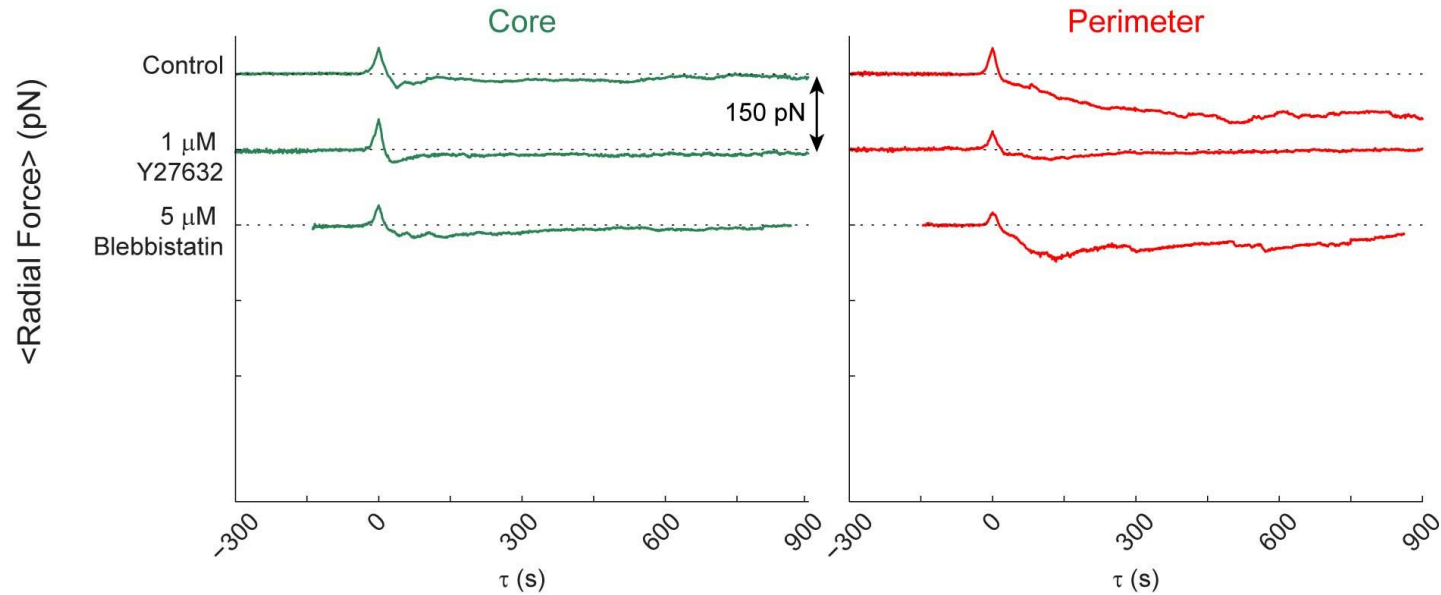
		n cells			
		n posts			
Protrusion	Force	Core			
		Perimeter			
	Duration	Core			
		Perimeter			
	Variance	Core			
		Perimeter			
	Participation	Core			
	Velocity	All Posts			
Contraction	Force	Core			
		Perimeter			
	Variance	Core			
		Perimeter			

* $p < 0.05$, Tukey-Kramer multi. comp.

Henry et al. 2015. *Biophys J.* (Under Revision)

= no sig. diff.

Sustained contractility is ROCK and Myosin II mediated



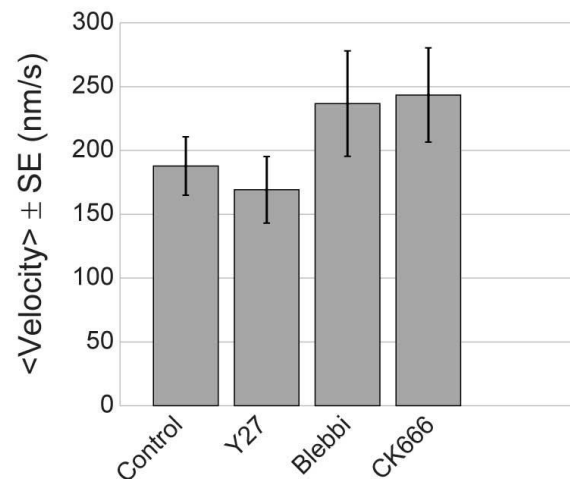
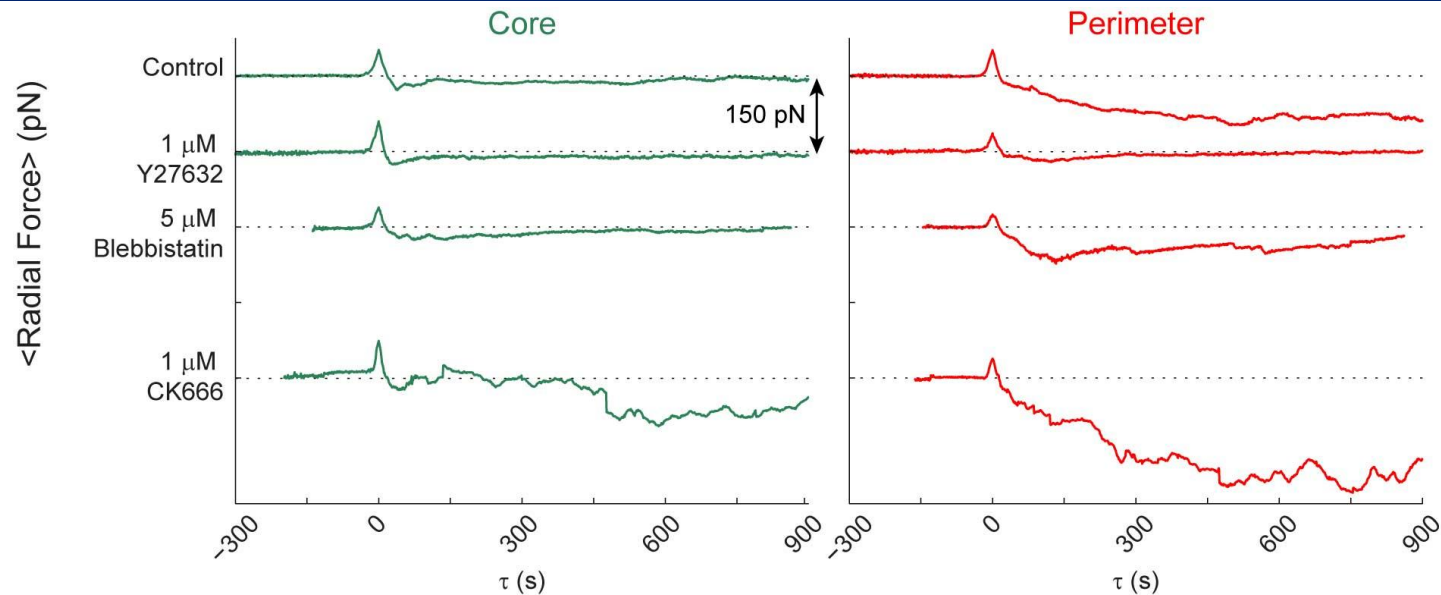
		Y27	Blebbi		
		n cells	10	4	
		n posts	325	117	
Protrusion	Force	Core			
		Perimeter			
	Duration	Core			
		Perimeter			
	Variance	Core			
		Perimeter			
	Velocity	All Posts			
Contraction	Force	Core			
		Perimeter	* ↓	* ↓	
	Variance	Core			
		Perimeter	* ↓		

☐ = no sig. diff.

* $p < 0.05$, Tukey-Kramer multi. comp.

Henry et al. 2015. *Biophys J.* {Under Revision}

Spreading is **not** actin-branching liable



		Y27	Blebbi	CK666	
		n cells	10	4	5
		n posts	325	117	161
Protrusion	Force	Core			
	Force	Perimeter			
	Duration	Core			
	Duration	Perimeter			
	Variance	Core			
	Variance	Perimeter			
Contraction	Participation	Core			
	Participation	Perimeter			
	Velocity	All Posts			
	Force	Core			
Contraction	Force	Perimeter	* ↓	* ↓	
	Variance	Core			* ↑
	Variance	Perimeter	* ↓		

☐ = no sig. diff.

* $p < 0.05$, Tukey-Kramer multi. comp.

Henry et al. 2015. *Biophys J.* (Under Revision)

Spreading is not analogous to lamellipodium formation

Hypothesis: Contraction is canonical
RhoA/Rock and Myosin Mediated

Y27632 (1 μ M)



p160ROCK

Blebbistatin (5 μ M)



NM Myosin II

Hypothesis: Protrusion is
lamellipodium formation

CK666 (1 μ M)

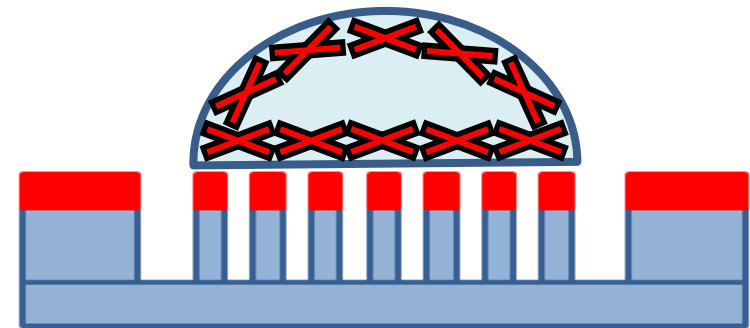
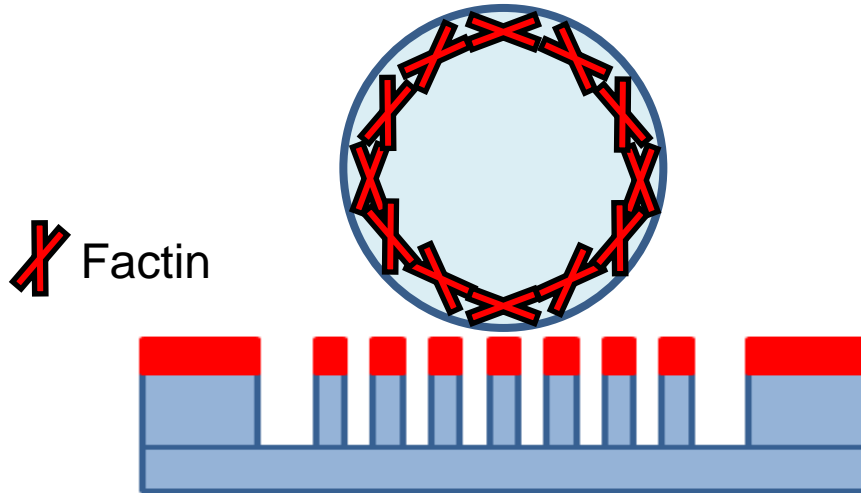


Arp2/3

Competition b/n adhesive energy and cortical stiffness?

Tension $>$ Adhesive Energy

Tension $<$ Adhesive Energy

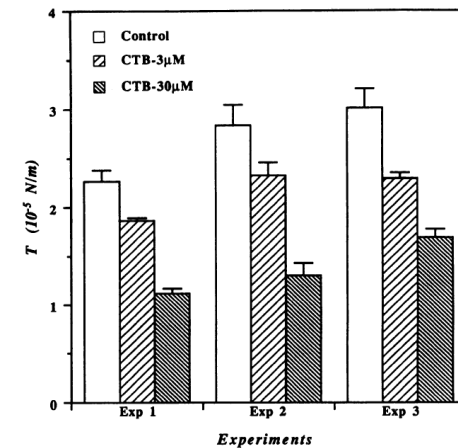
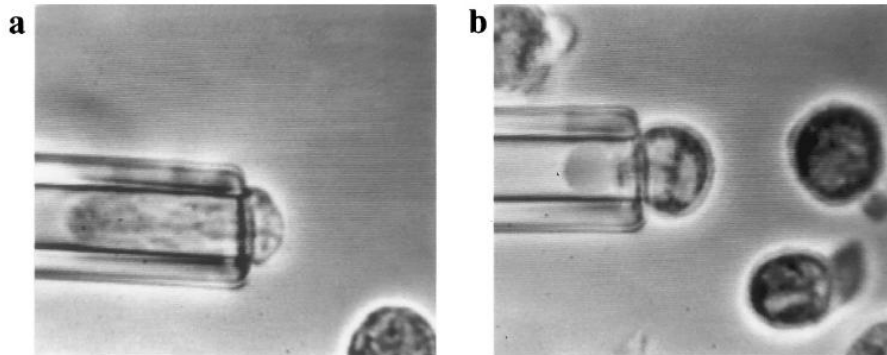


Jasplakinolide = stiffening

Cytochalasin B = softening

Control

10 μ M



A revised hypothesis:

Hypothesis: Contraction is canonical RhoA/Rock and Myosin Mediated

Y27632 (1 uM)



p160ROCK

Blebbistatin (5 uM)



NM Myosin II

Hypothesis: cortical tension resists spreading

Jasplakinolide (1 uM)



actin
depolymerization;
(↑ cortical stiffness)

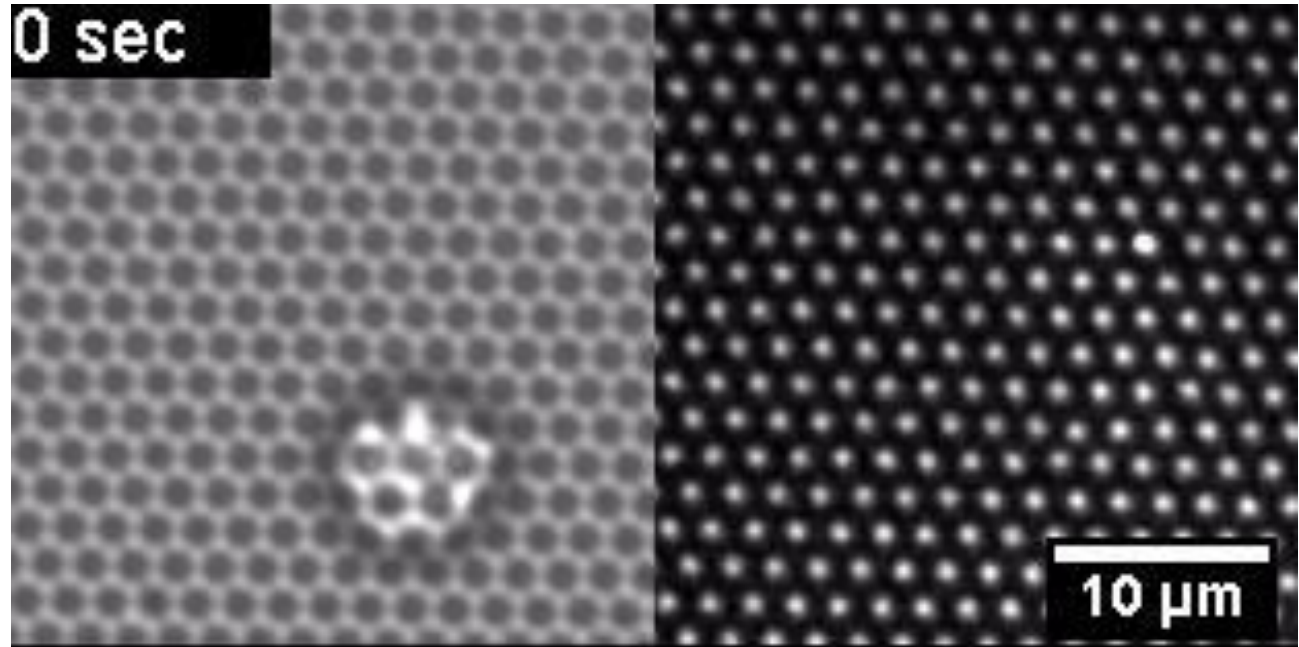
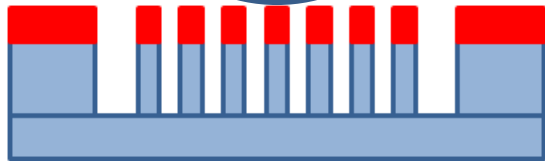
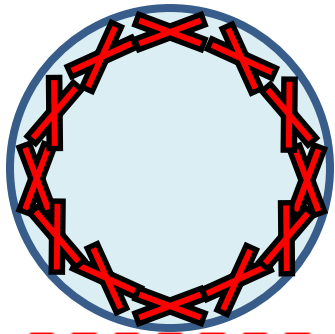
CytochalasinB (3 uM)



actin polymerization
& filament interaction;
(↓ cortical stiffness)

Cortical stiffening via Jasplakinolide abrogates spreading

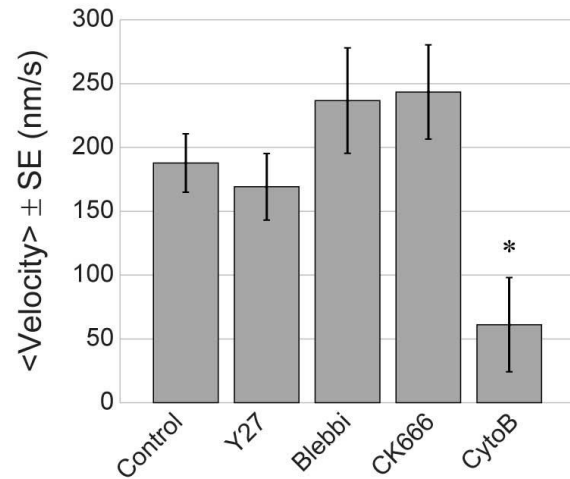
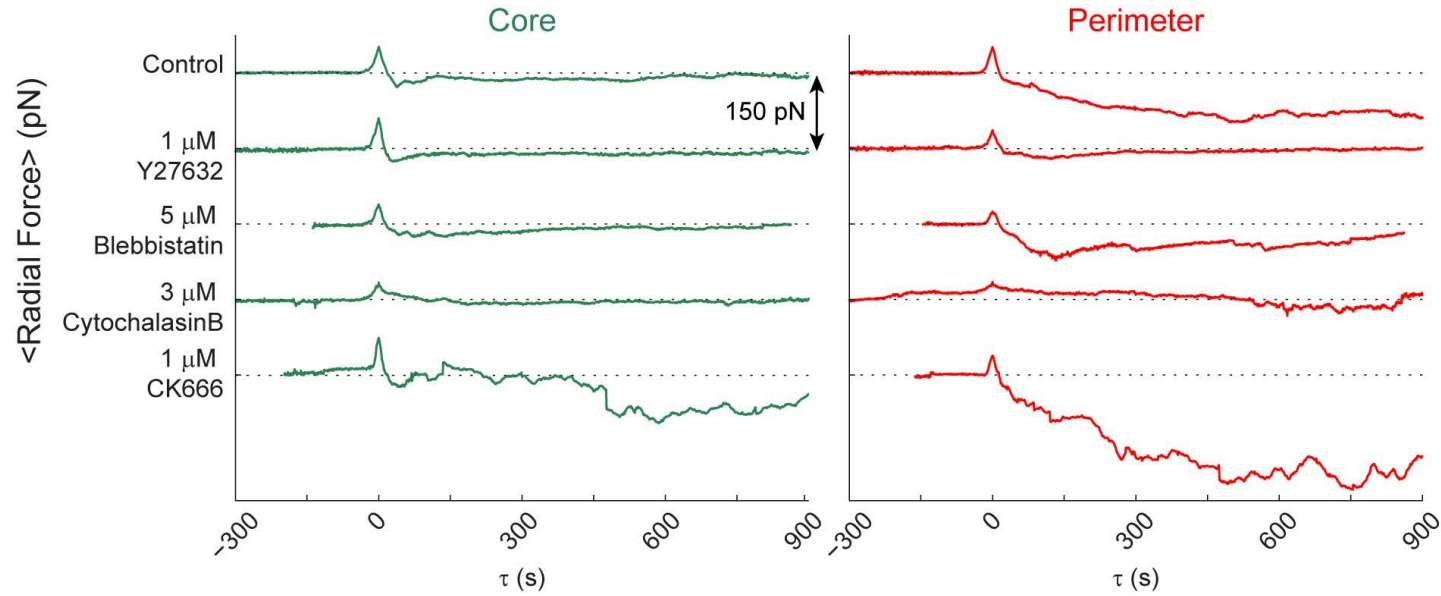
1 μM Jasplakinolide
inhibits cortical actin
depolymerization



 Cross-linked filamentous actin

Cortical stiffening eliminates spreading

Cortical softening slows spreading



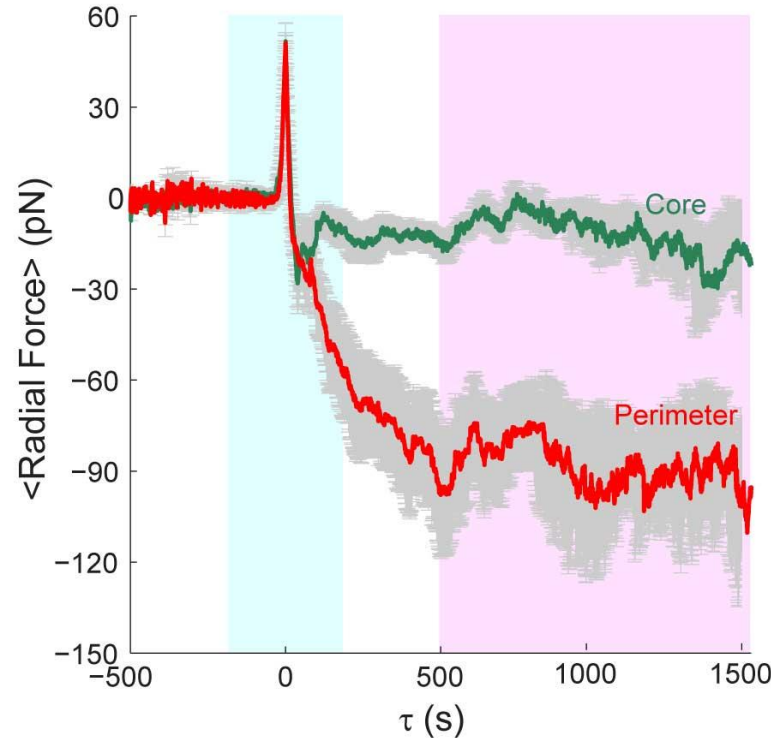
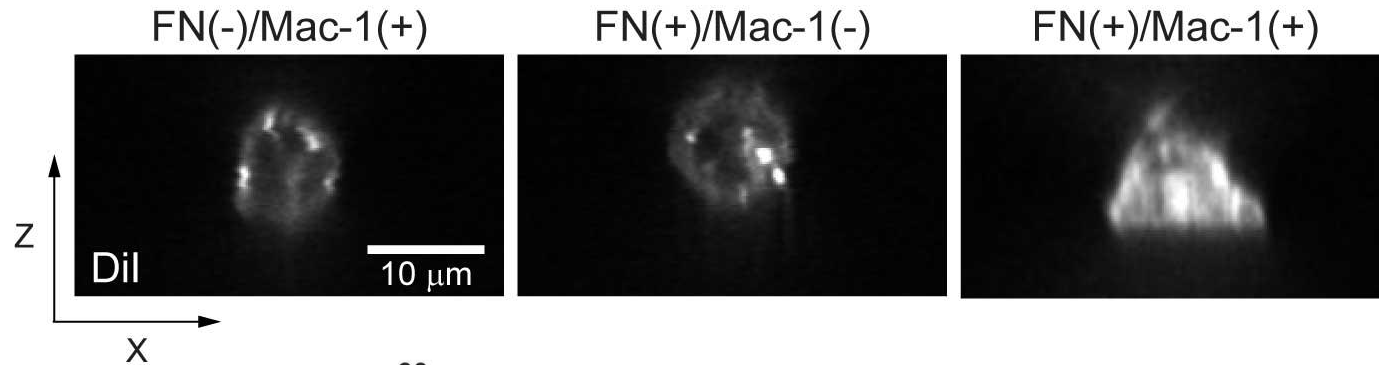
		Y27	Blebbi	CK666	Jasp	CytoB
		n cells	4	5	9	6
		n posts	117	161	n/a	177
Protrusion	Force	Core				
	Force	Perimeter				
	Duration	Core				* ↑
	Duration	Perimeter				* ↑
	Variance	Core				
	Variance	Perimeter				
Contraction	Participation	Core				
	Participation	Perimeter				
	Velocity	All Posts				* ↓
	Force	Core				
	Force	Perimeter	* ↓	* ↓		* ↓
	Variance	Core			* ↑	
Variance	Perimeter	* ↓				

□ = no sig. diff.

* p < 0.05, Tukey-Kramer multi. comp.

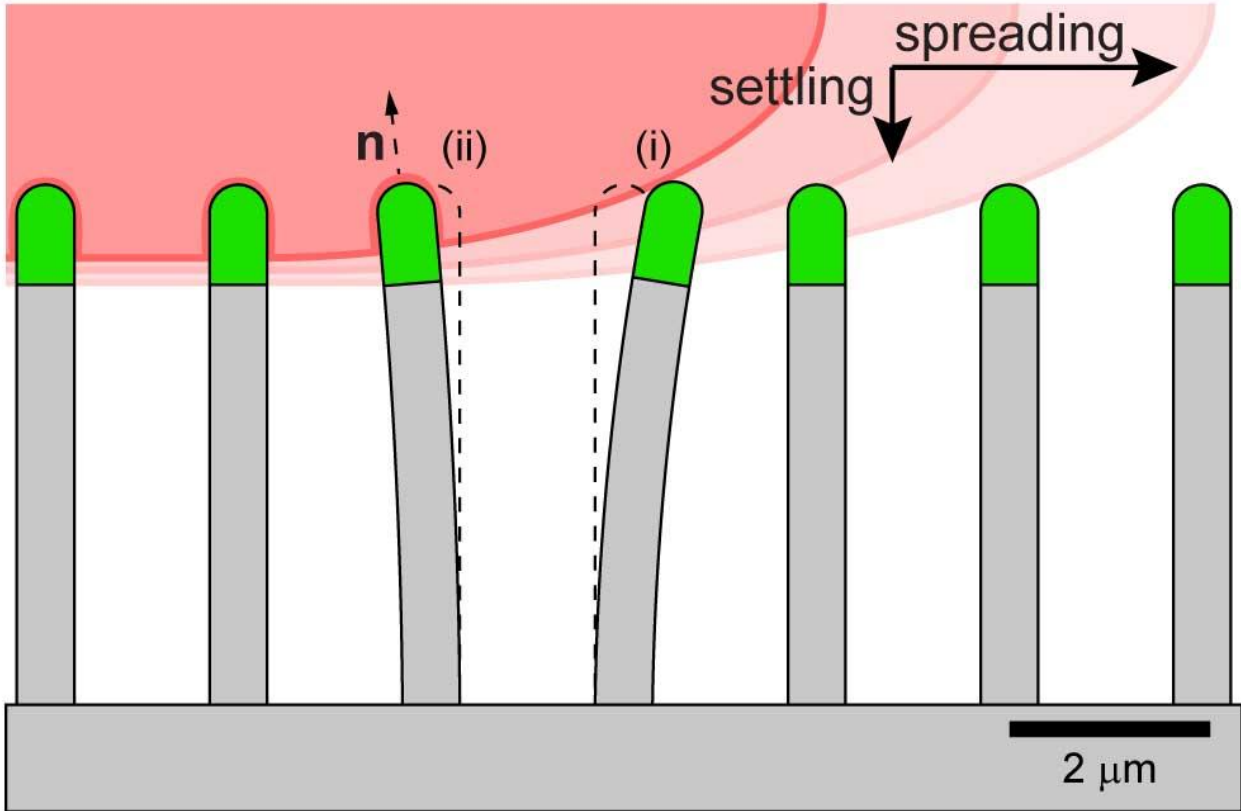
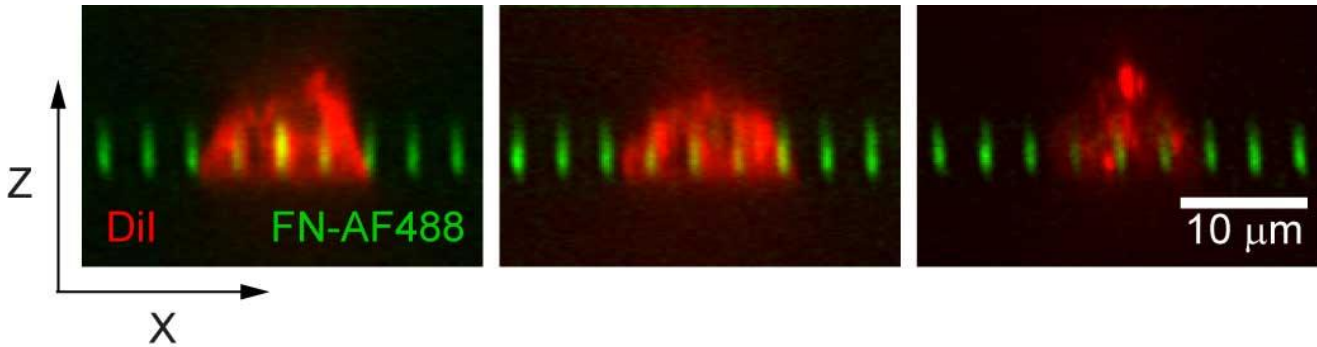
Henry et al. 2015. *Biophys J.* (Under Revision)

Spreading is integrin mediated **but** connection to the mature actomyosin substructure takes minutes to develop...



Why do we see protrusion at all?

Invagination: a spreading neutrophil pushing through post tips

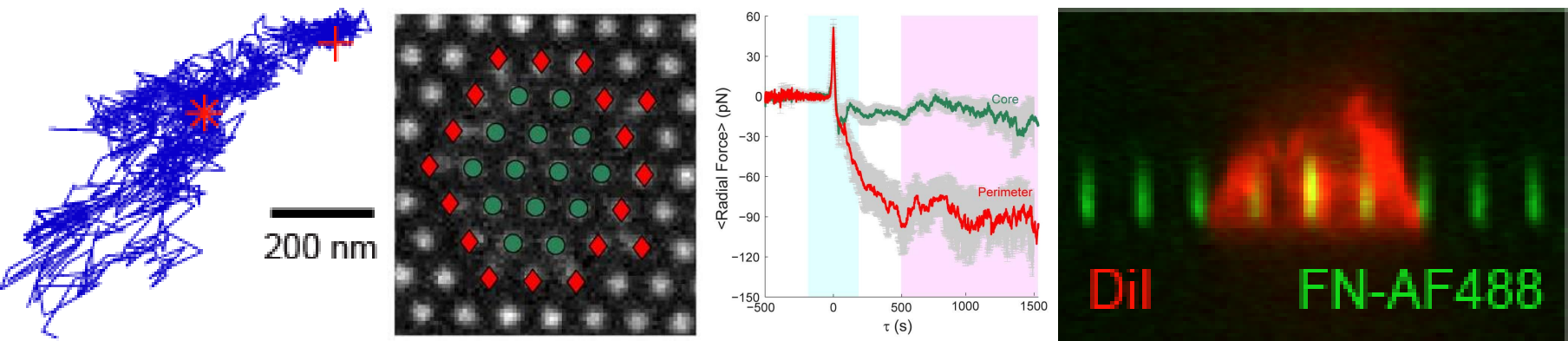


Calibrated
invagination
depth $\sim 1 \mu\text{m}$

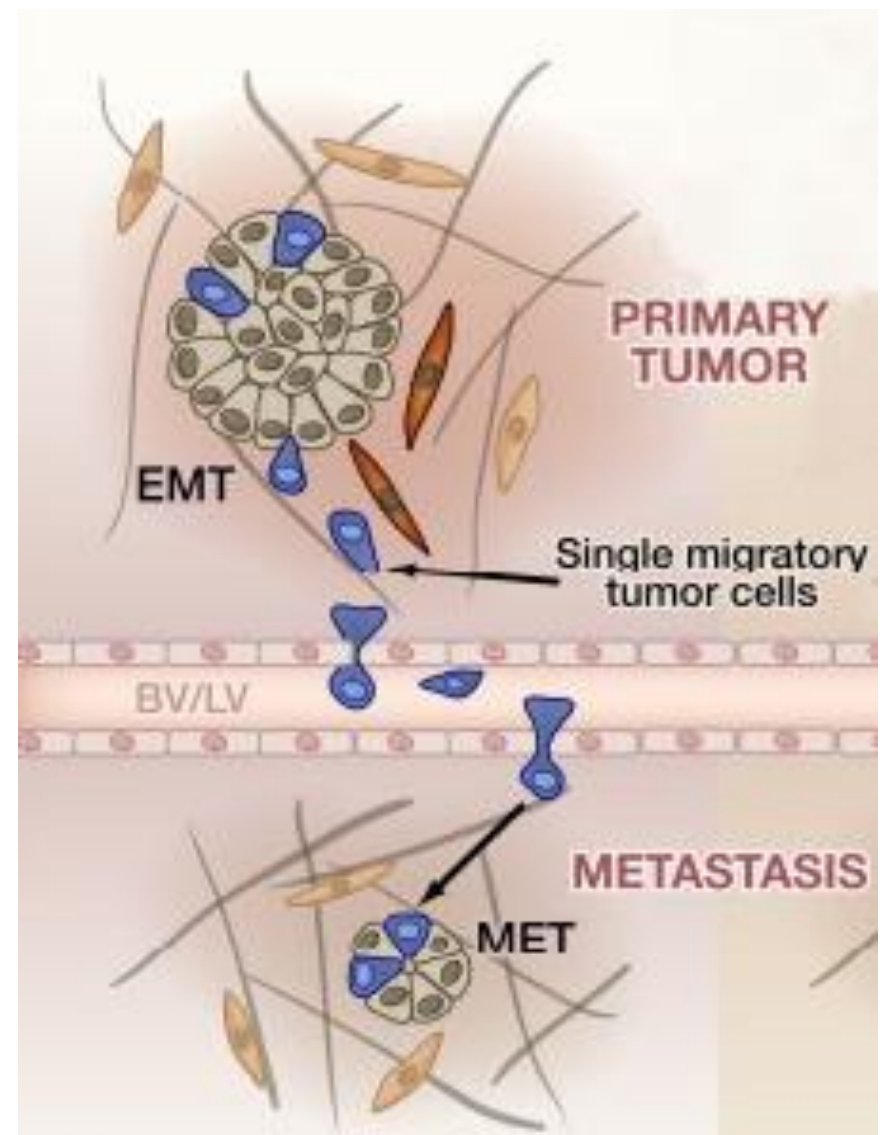
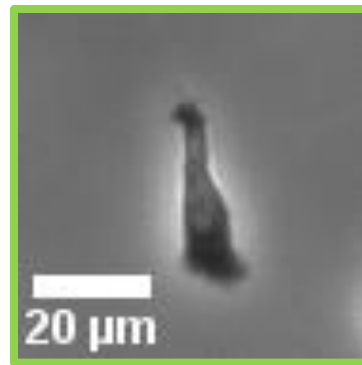
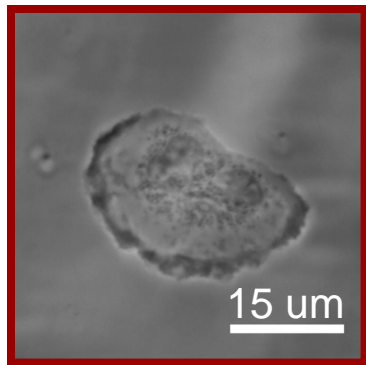
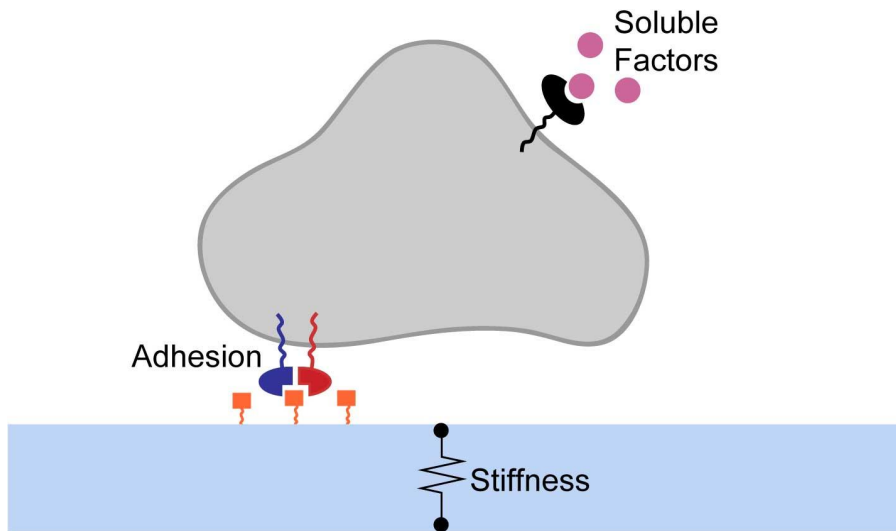
Schematic
to scale

Part III Summary

Neutrophil adhesion-driven spreading is itself a phenotypic switch triggered by decrease in resting cortical tension.



Role of adhesivity in cancer metastasis?



Thank you!

Advisor

Daniel A. Hammer, PhD

Committee

Scott L. Diamond, PhD (Chair)

John C. Crocker, PhD

Dongeun Huh, PhD

Scientific Collaborators

John C. Crocker, PhD

Christopher S. Chen, PhD

Neha P. Kamat, PhD

Daeyeon Lee, PhD

Hammer Lab

All members past and present

Funding

National Institutes of Health

(HL18208 to DAH)

National Science Foundation

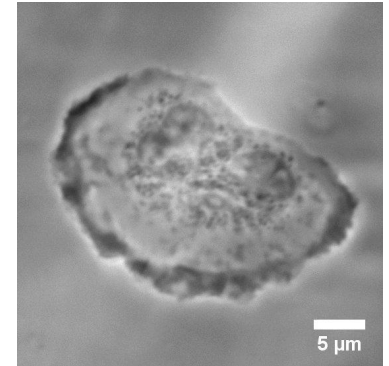
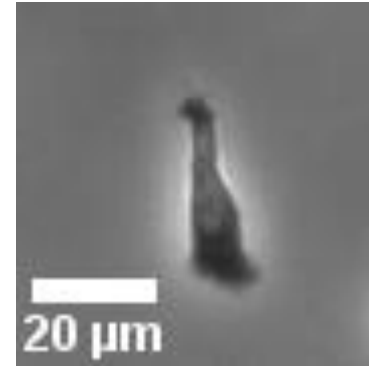
(GRFP to SJH)

Questions?

Shape and Motility

Ligand density elicits phenotypic switch in human neutrophils

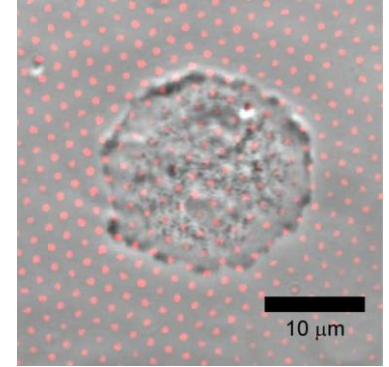
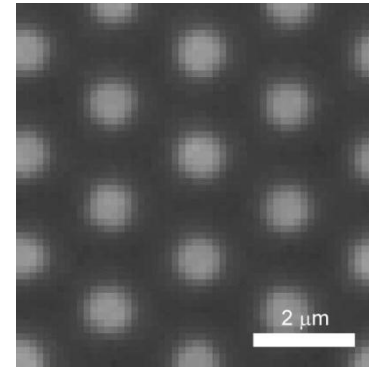
Henry, Crocker, Hammer. 2014. *Integr Biol.*



Density Sensing

Dynamic traction forces of spreading and adherent human neutrophils

Henry, Crocker, Hammer. 2015. *ABME* (In Prep)



Spreading Mechanics

Dynamic traction forces of spreading and adherent human neutrophils

Henry, Chen, Crocker, Hammer. 2015. *Biophys J.* (Under Revision)

