

Biophysical Society  
2015 Annual Meeting  
Baltimore, MD

02/11/2015

Cell Mechanics,  
Mechanosensing, and Motility II  
10:00 AM, Room 307/308

\*\* Denotes movies played during talk

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# Dynamic Traction Forces of Human Neutrophil Adhesion

**Steven J. Henry,**

Christopher S. Chen, John C. Crocker, and Daniel A. Hammer

Funding:

NIH HL18208 to DAH

NSF GRFP to SJH

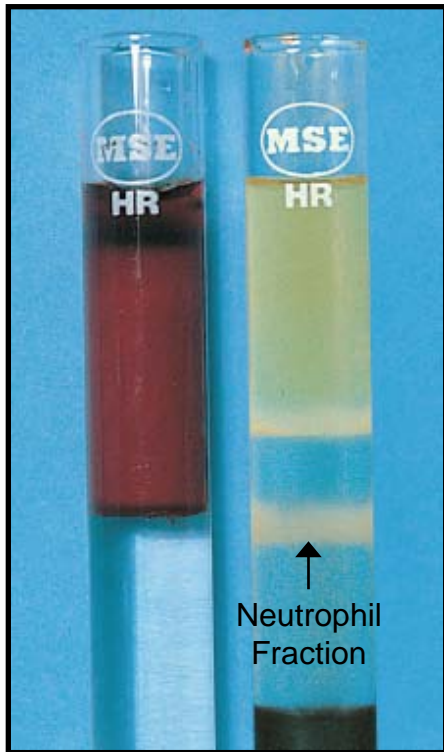
Note:

Manuscript submitted to *Biophysical Journal* for review

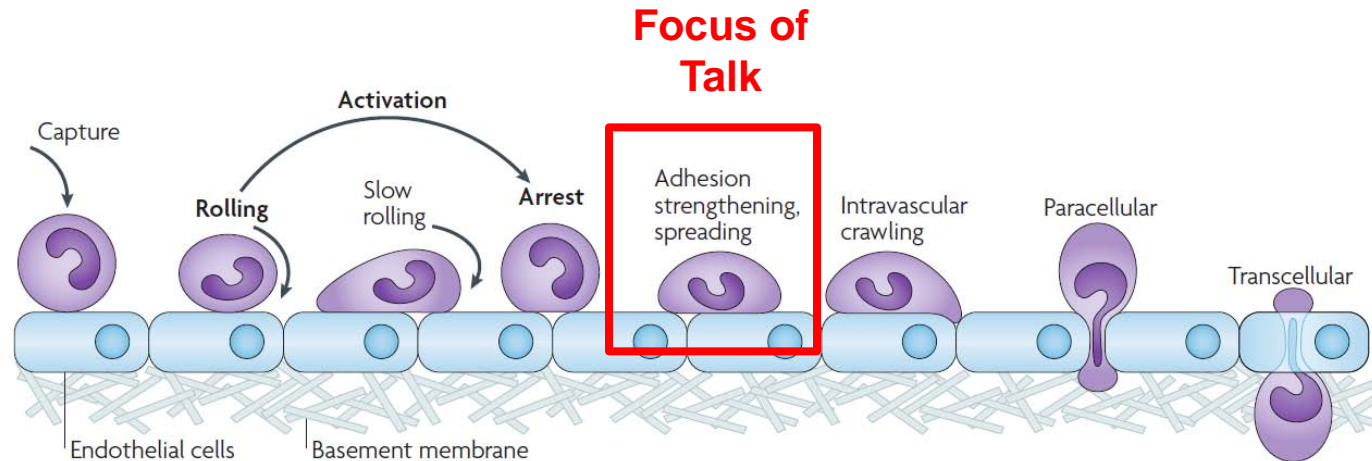


# Neutrophils: first responders to trauma and infection

Whole Blood    After Density  
                  Centrifugation

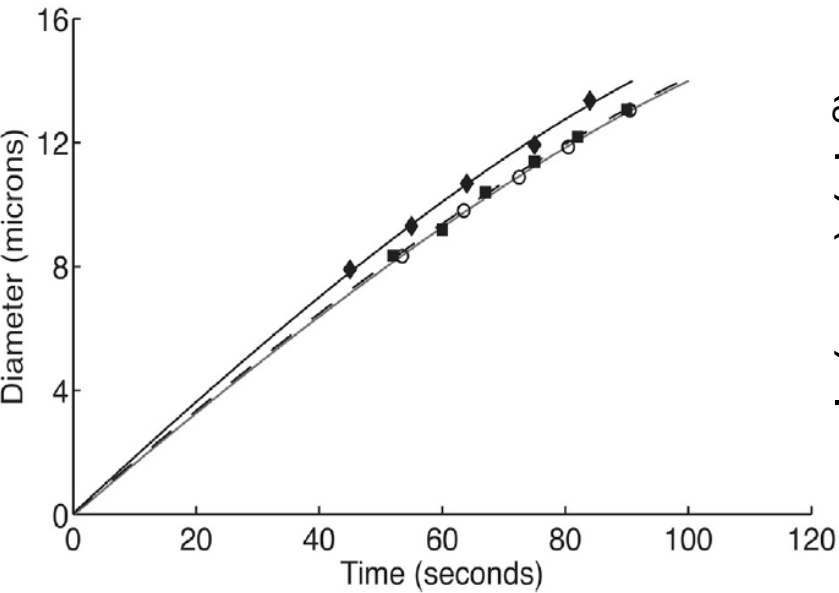
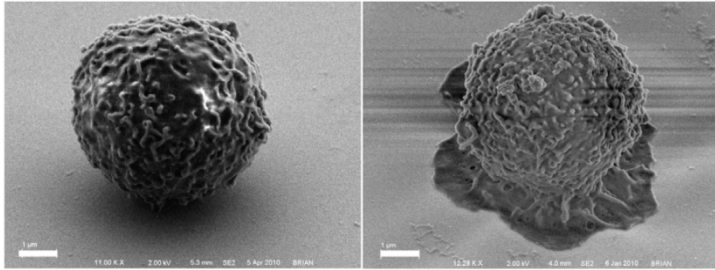


Axis-Shield News  
Bulletin

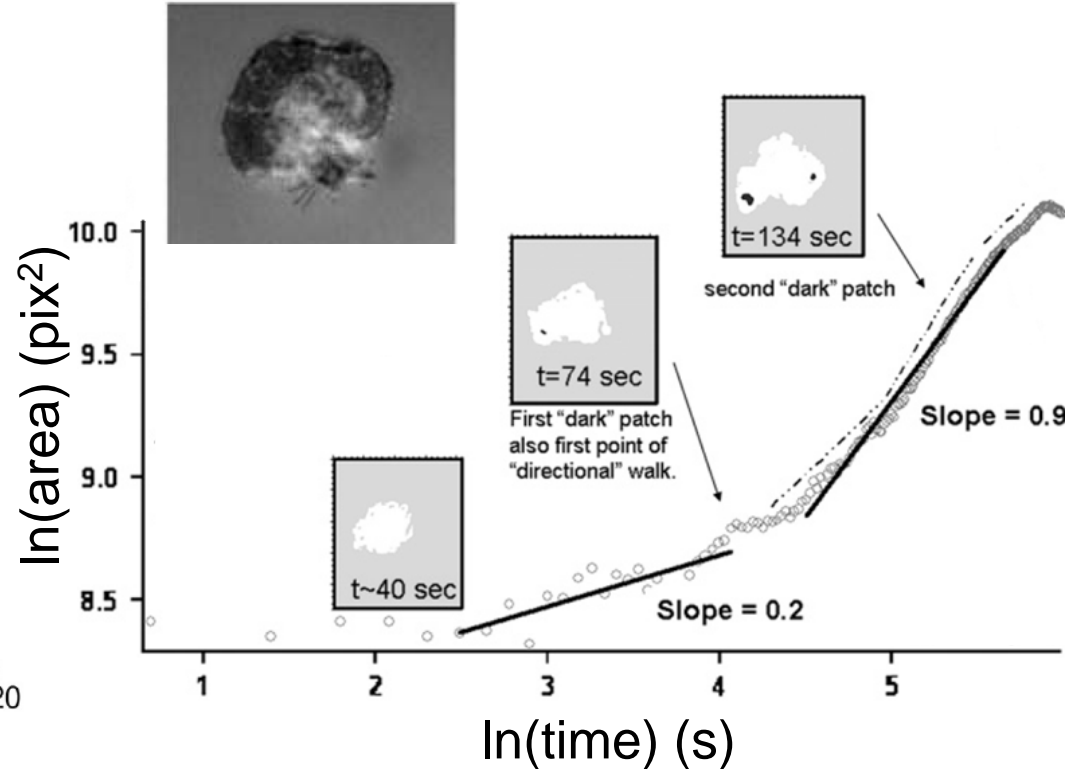


Ley et al. 2007. *Nat Rev Immunol.*

# Neutrophil spreading is fast. Can we measure the associated forces?

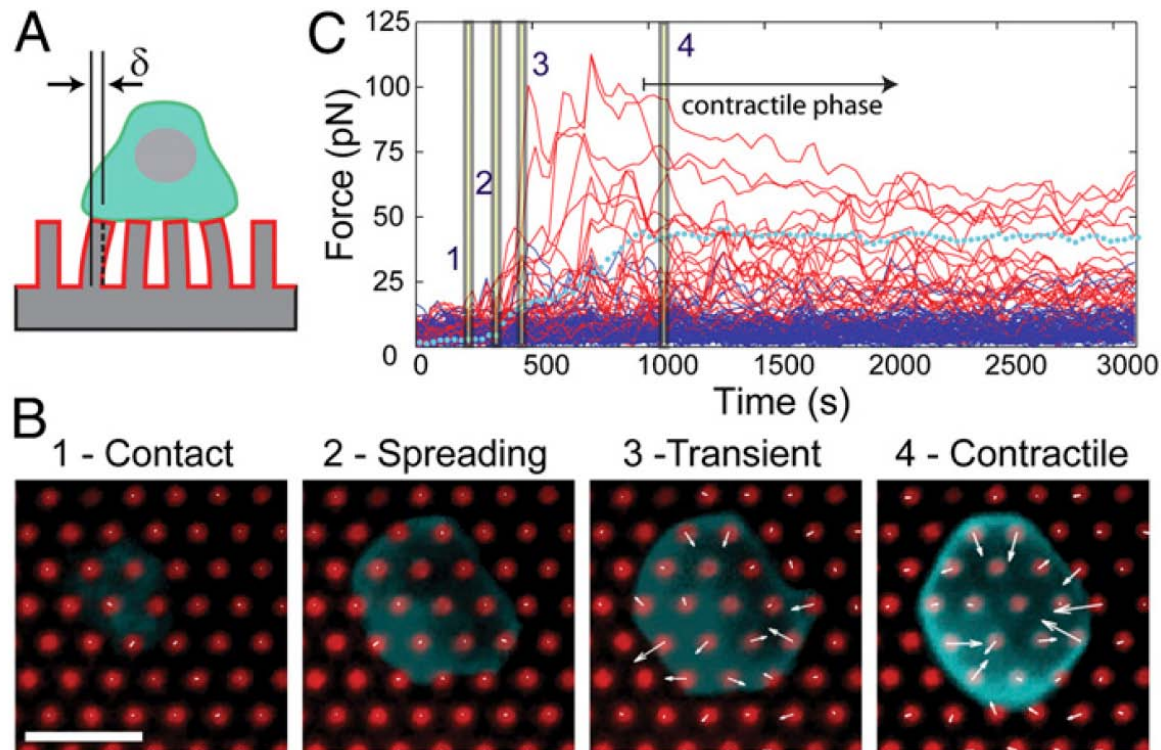


Lomakina et al. 2014. *Biophys J.*  
(Waugh Lab)



Sengupta et al. 2006. *Biophys J.*  
(Hammer Lab)

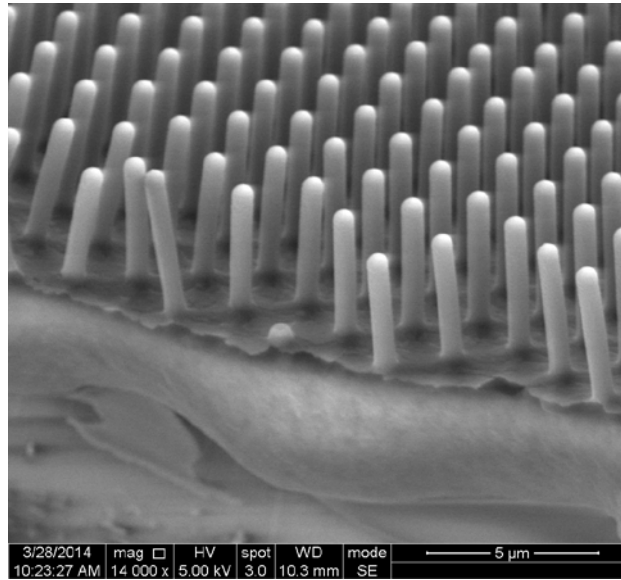
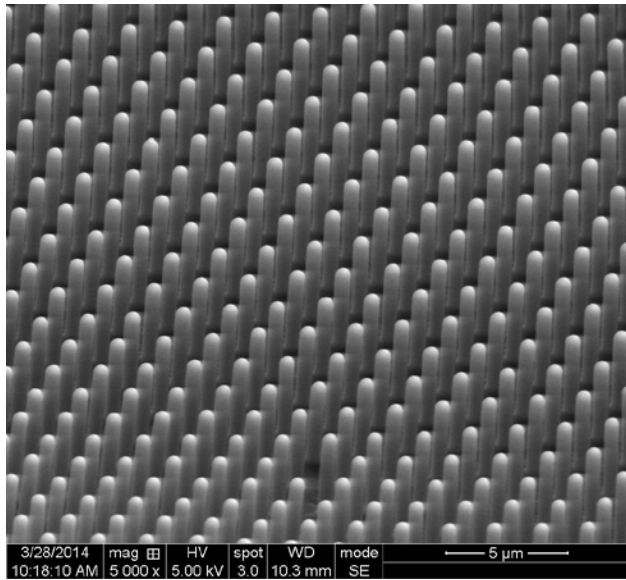
# Traction forces of T-lymphocyte activation on microfabricated Post-Array-Detectors (mPADs)



Bashour et al. 2014. *PNAS* (Kam Lab)

Dynamics of protrusive vs. contractile processes?  
Role of the actin cytoskeleton?

# microfabricated Post-Array-Detectors (mPADs)



$$k_{\text{spring}} = 0.28 \pm 0.07 \text{ nN/um}$$

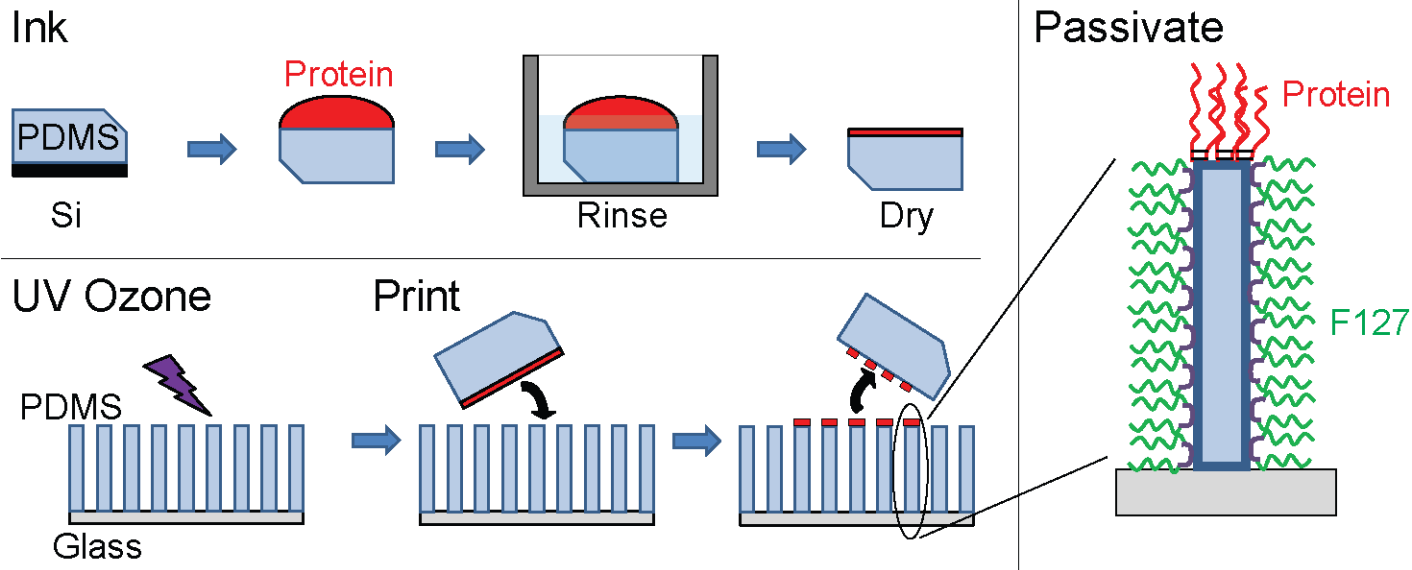
$$G \sim 5 \text{ kPa}$$

Schoen correction = 0.93

$$k_{\text{spring}}^* = (0.93)(k_{\text{spring}})$$

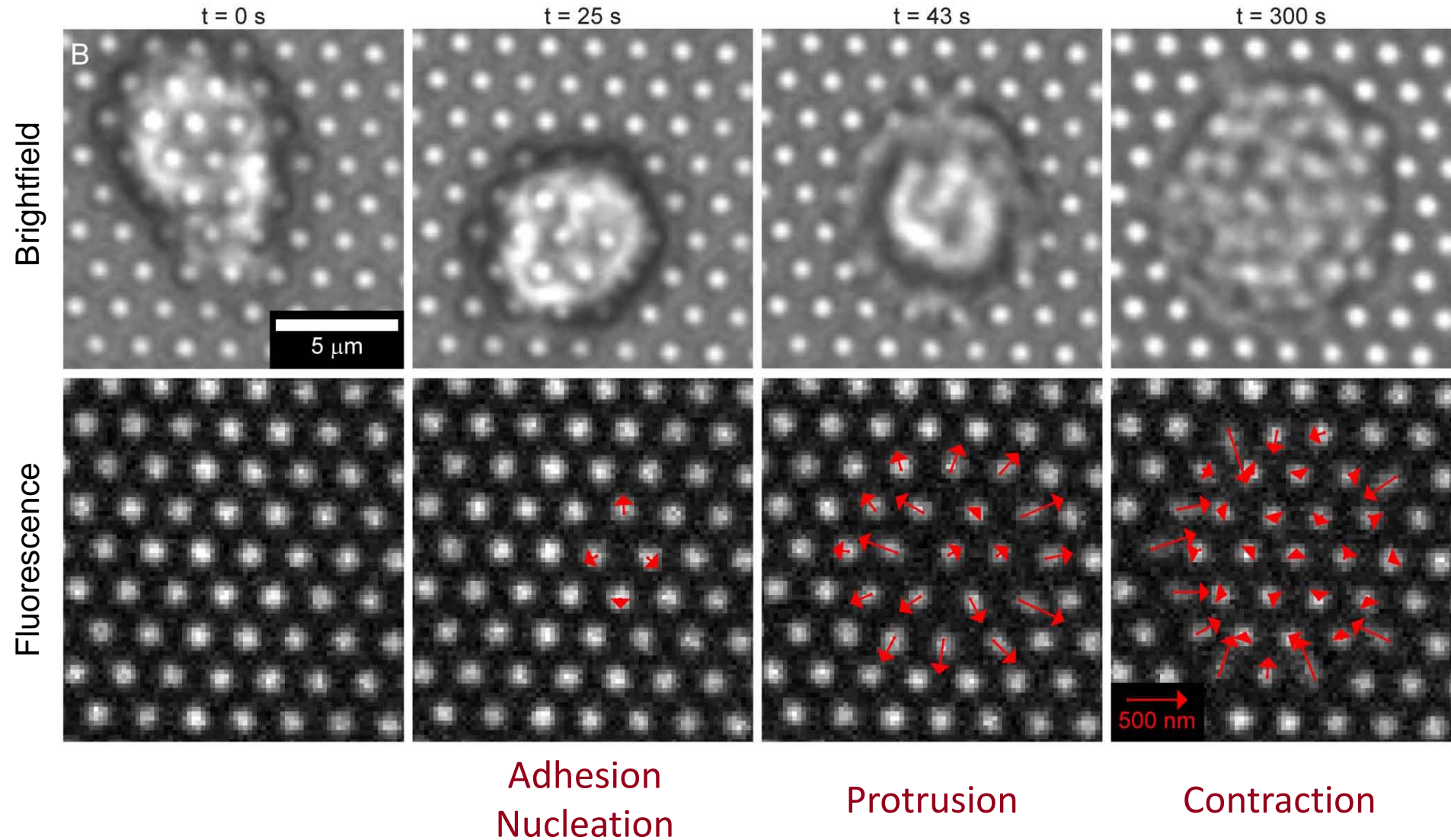
$$k_{\text{spring}}^* = 0.26 \text{ nN/um}$$

Schoen et al. 2010. *NanoLett.*



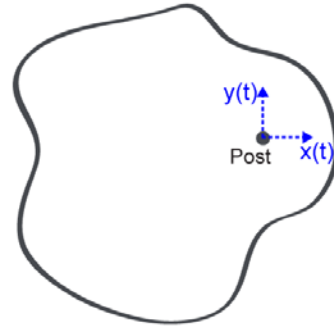


# \*\*Neutrophil spreading on mPADs

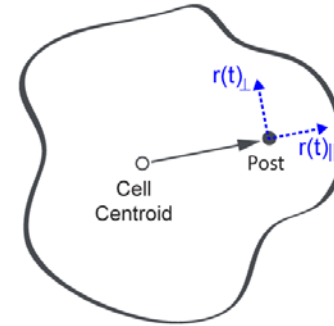


# Plotting force trajectories in the cell reference frame

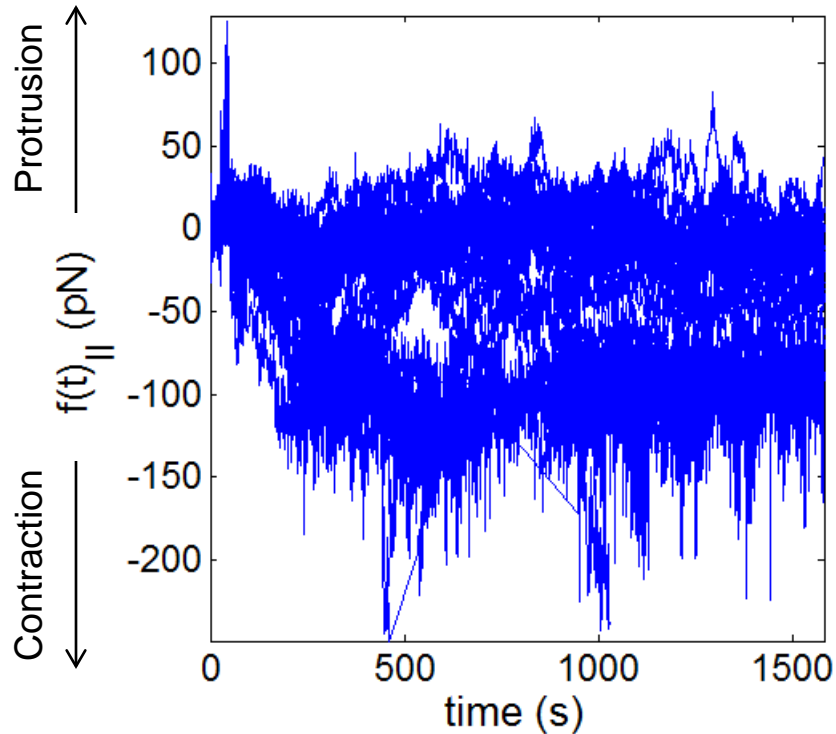
Lab Reference Frame



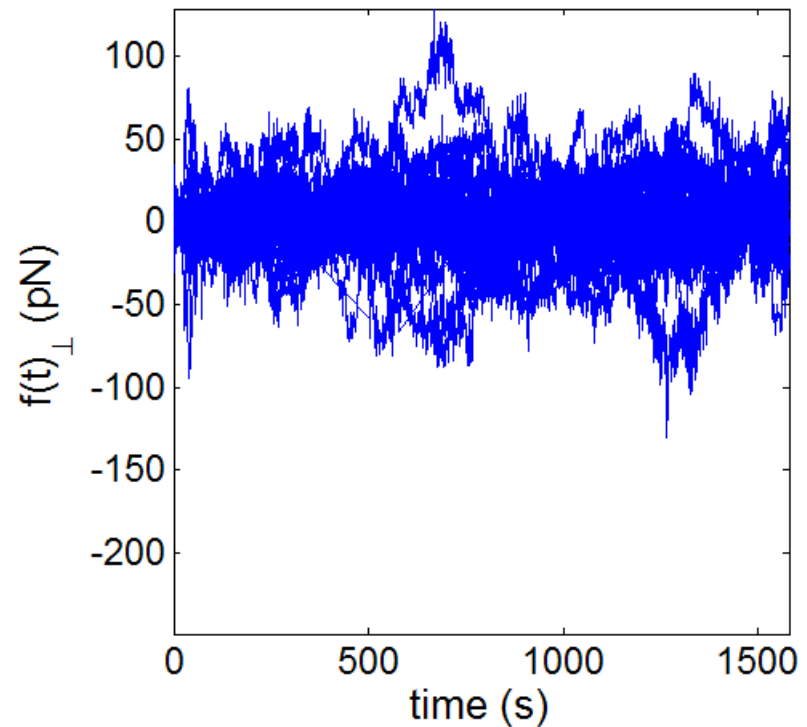
Cell Reference Frame



Radial

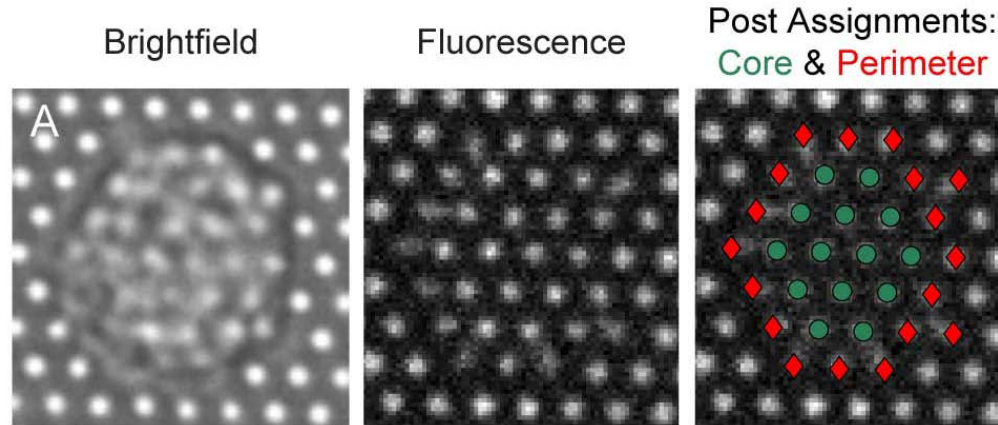


Tangential



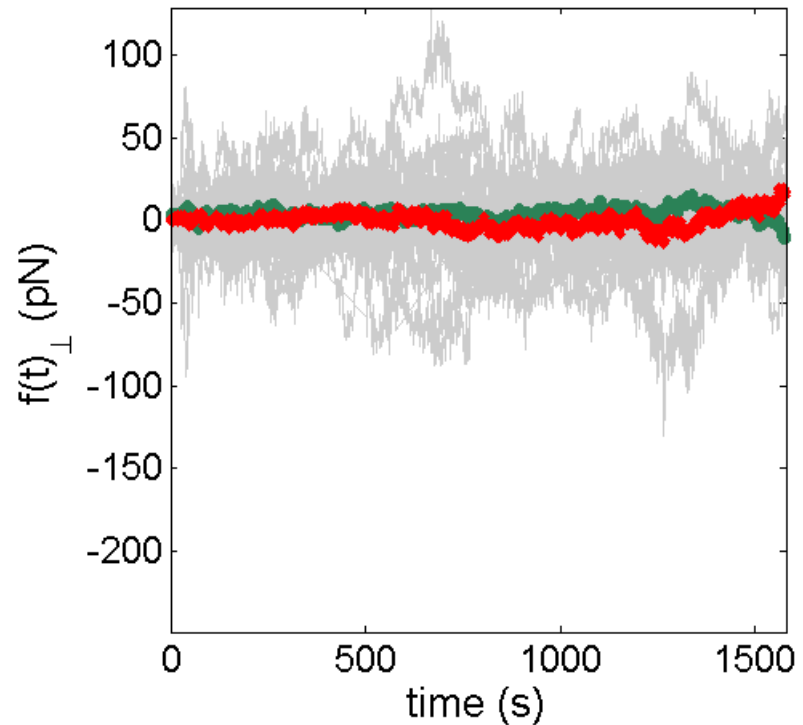
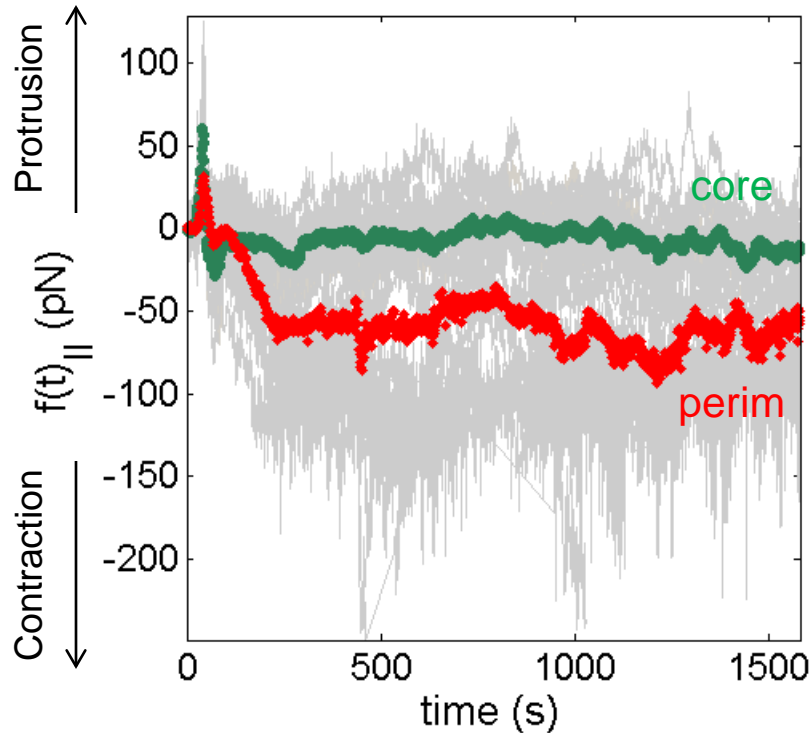


# Dichotomizing data on geometric location



Radial

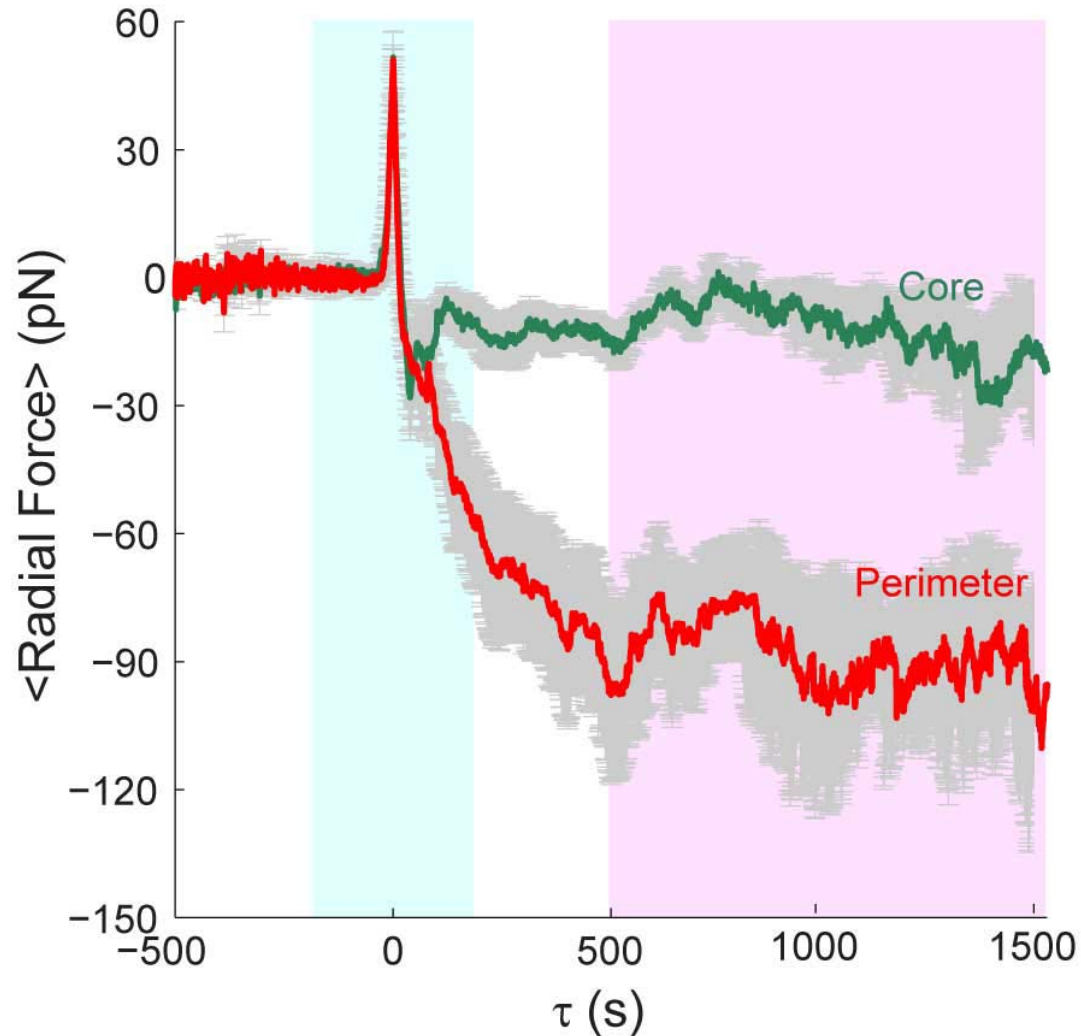
Tangential



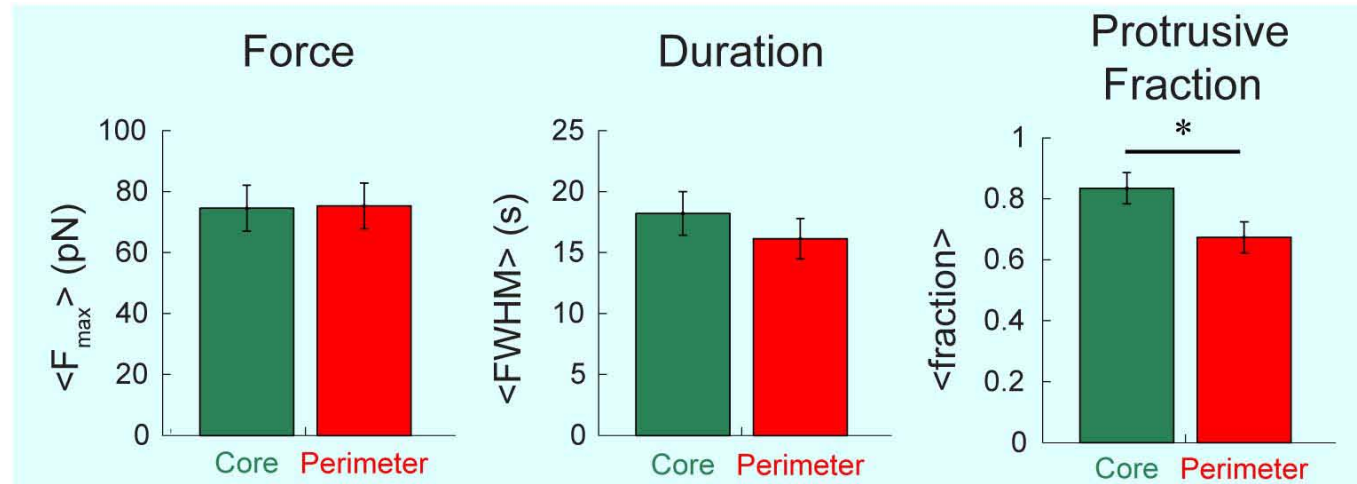
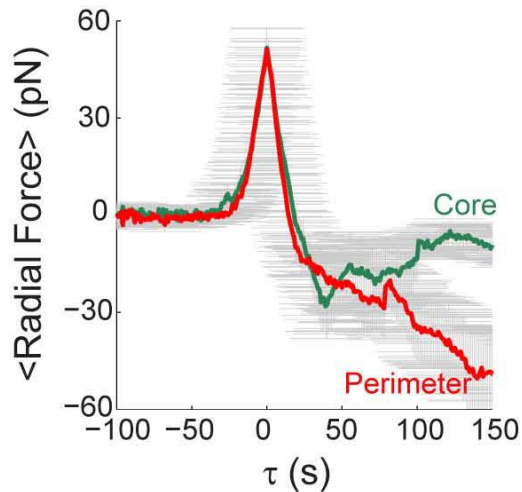
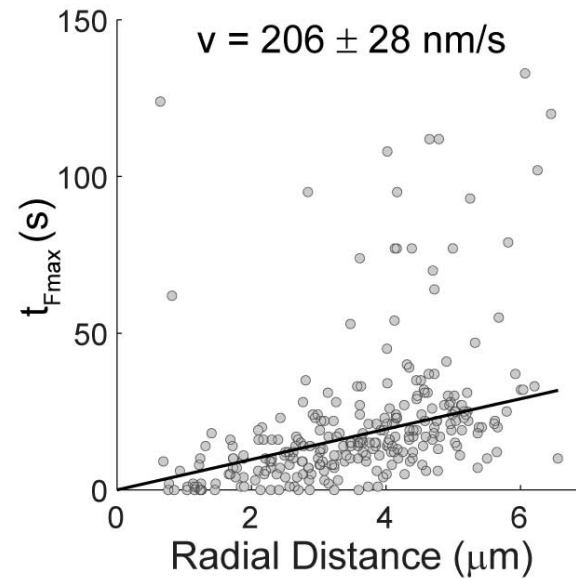
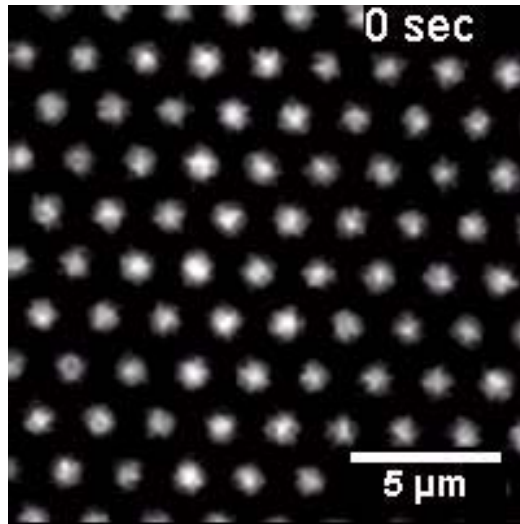
# Distinct mechanical regimes are apparent

Transient Protrusion

Steady State Contraction

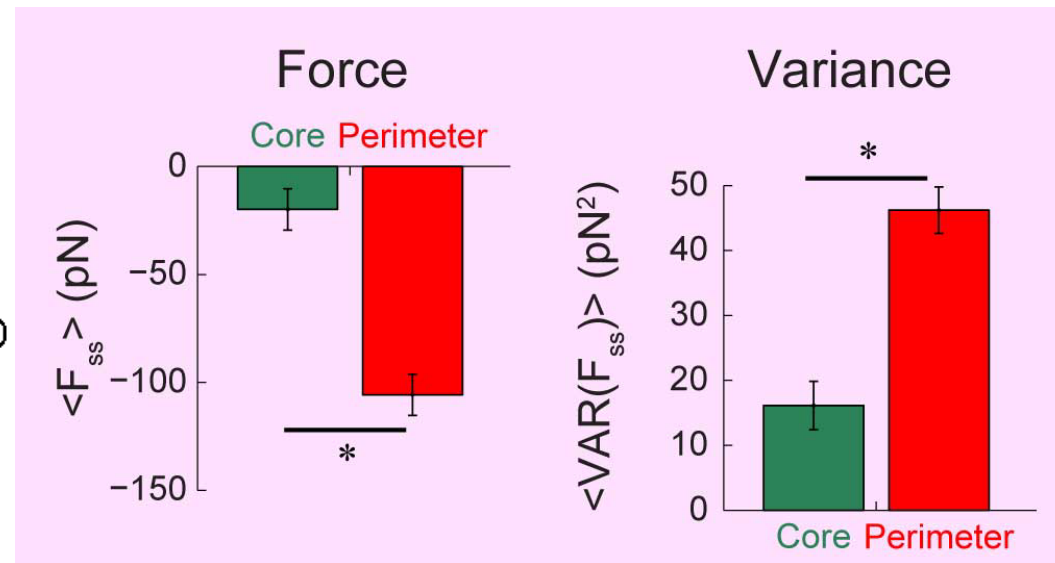
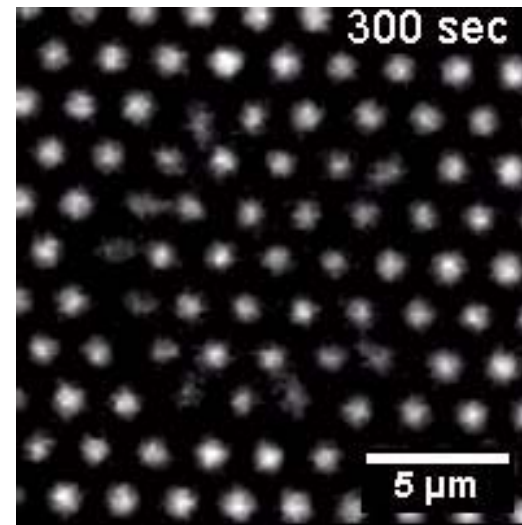
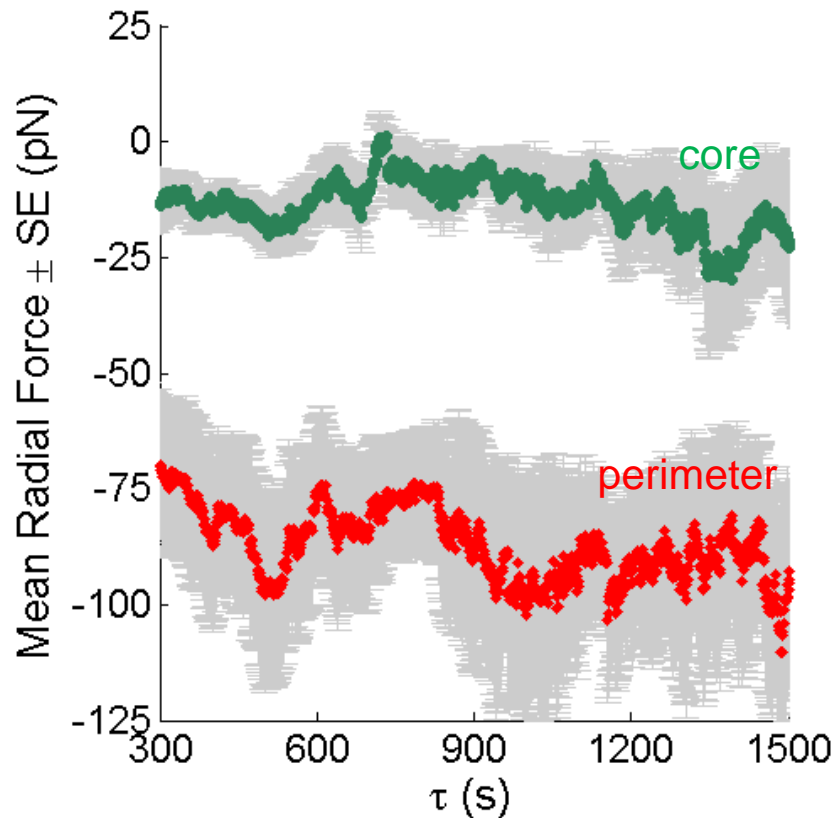


# \*\*Characterizing the protrusive wave



\*  $p < 0.05$ , Tukey LSD method,  $n = 14$  cells

# \*\*Characterizing the Steady State Contractile Regime



\*  $p < 0.05$ , Tukey LSD method,  $n = 14$  cells

# Are protrusion and contraction biochemically distinct?

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Hypothesis: Contraction is canonical  
RhoA/Rock and Myosin Mediated

Y27632 (1  $\mu$ M)

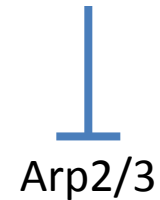


Blebbistatin (5  $\mu$ M)

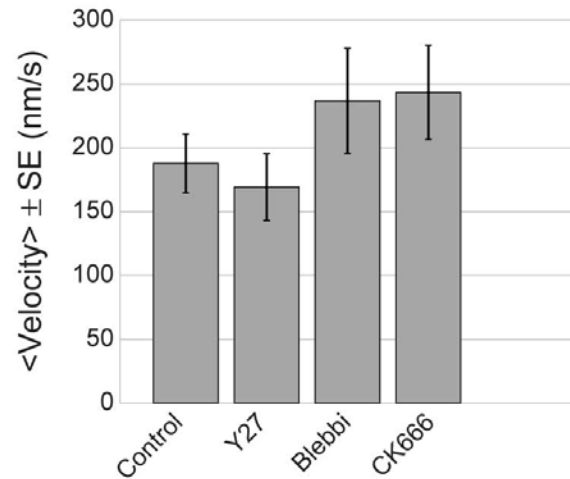
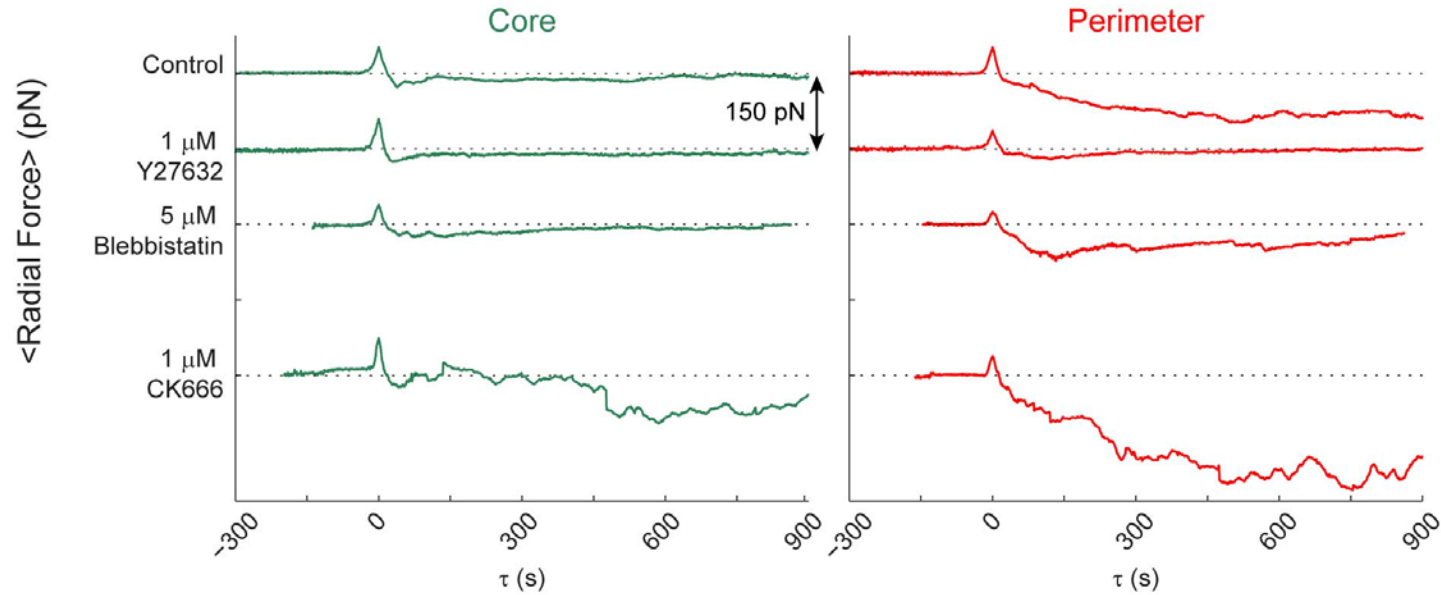


Hypothesis: Protrusion is  
lamellipodium formation

CK666 (1  $\mu$ M)



# Sustained contractility ROCK and Myosin II mediated but spreading **not** analogous to lamellipodium formation



		Y27	Blebbi	CK666
n cells		10	4	5
n posts		325	117	161
Protrusion	<Fmax>	Core		
		Perimeter		
	<FWHM>	Core		
		Perimeter		
	<VAR(Fmax)>	Core		
		Perimeter		
	<Participation>	Core		
		Perimeter		
	<Velocity>	All Posts		
Contraction	<Fss>	Core		
		Perimeter	* ↓	* ↓
	<VAR(Fss)>	Core		* ↑
		Perimeter	* ↓	

□ = no sig. diff.

\* p < 0.05, Tukey-Kramer multi. comp.



# Spreading is not analogous to lamellipodium formation

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Hypothesis: Contraction is canonical  
RhoA/Rock and Myosin Mediated

Y27632 (1  $\mu$ M)



p160ROCK

Blebbistatin (5  $\mu$ M)



NM Myosin II

Hypothesis: Protrusion is  
lamellipodium formation

CK666 (1  $\mu$ M)

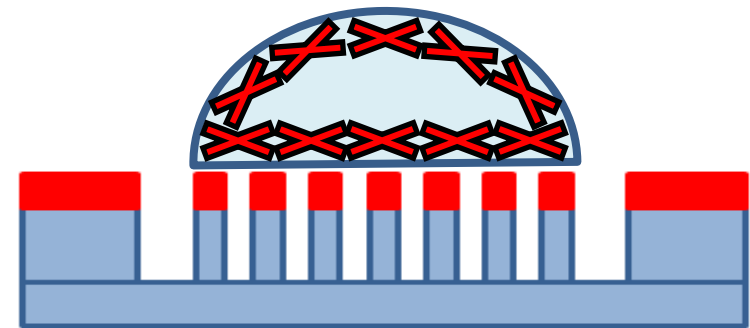
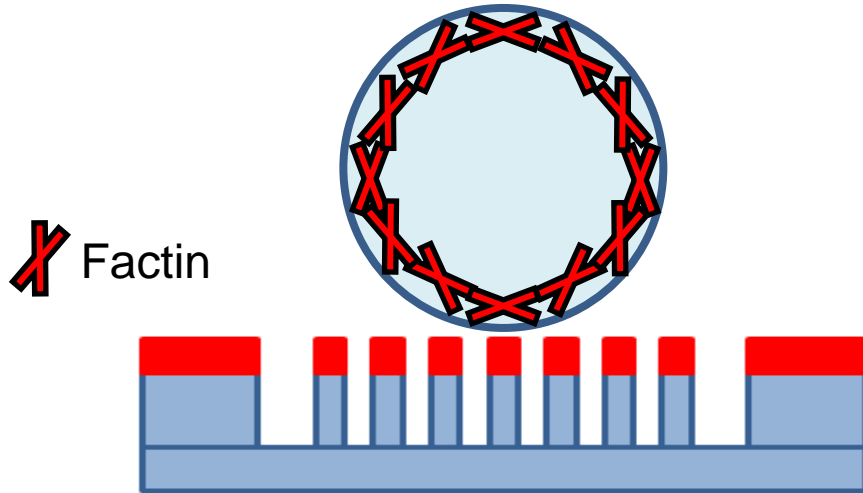


Arp2/3

# Competition b/n adhesive energy and cortical stiffness?

Tension  $>$  Adhesive Energy

Tension  $<$  Adhesive Energy

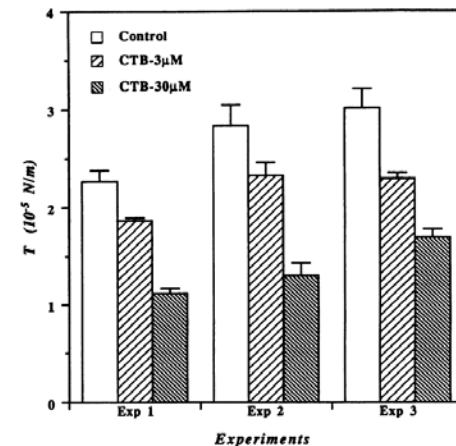
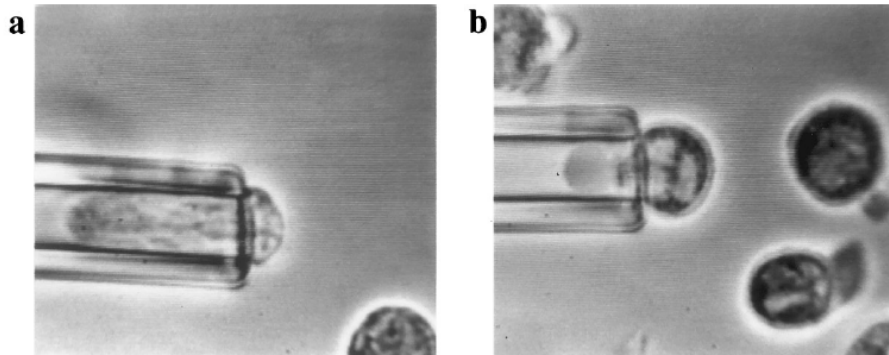


Jasplakinolide = stiffening

Cytochalasin B = softening

Control

10  $\mu$ M



# A revised hypothesis:

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Hypothesis: Contraction is canonical RhoA/Rock and Myosin Mediated

Y27632 (1 uM)



p160ROCK

Blebbistatin (5 uM)



NM Myosin II

Hypothesis: cortical tension resists spreading

Jasplakinolide (1 uM)



actin  
depolymerization;  
(↑ cortical stiffness)

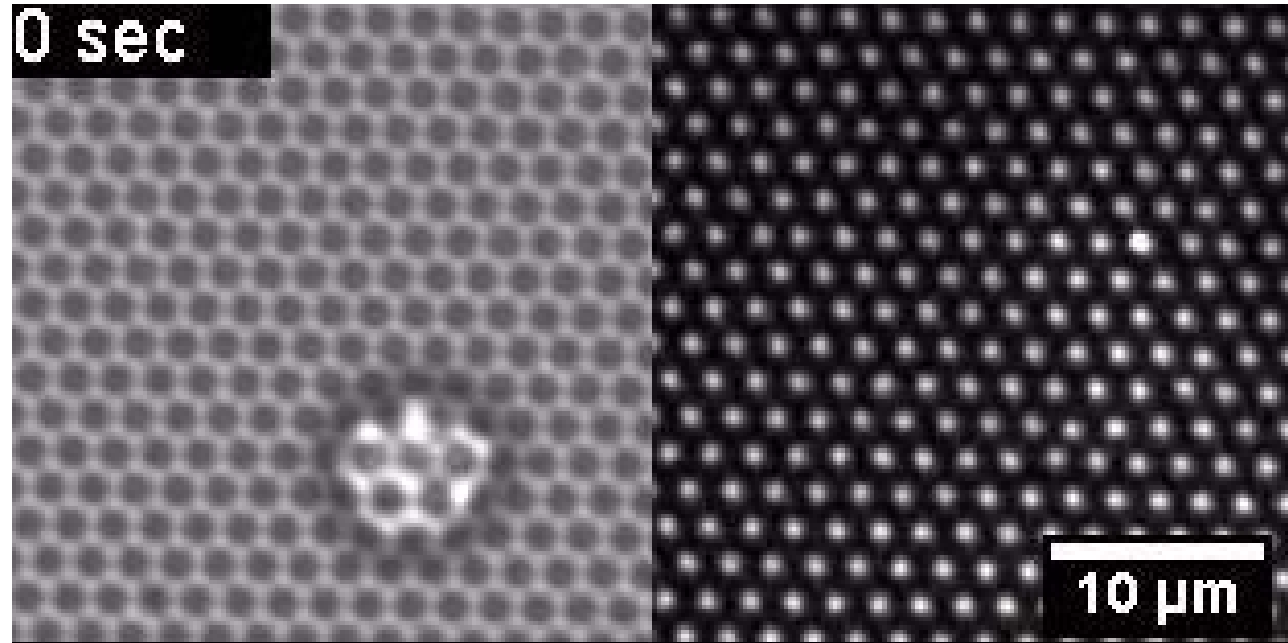
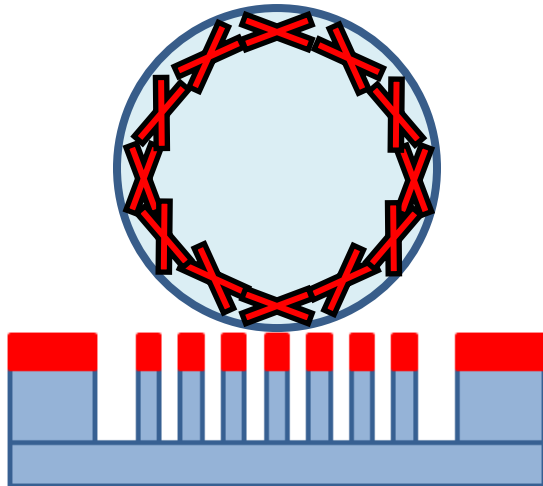
CytochalasinB (3 uM)



actin polymerization  
& filament interaction;  
(↓ cortical stiffness)

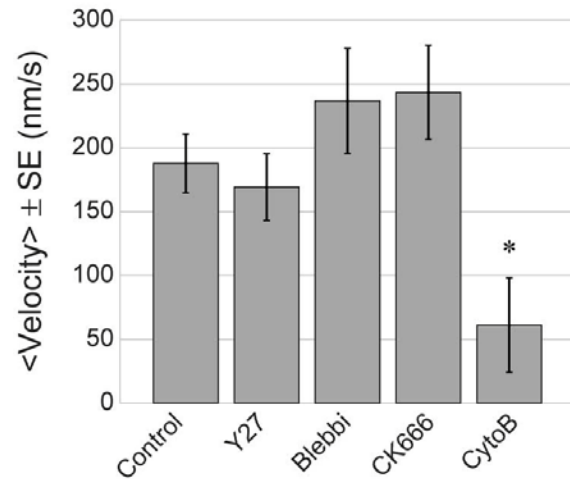
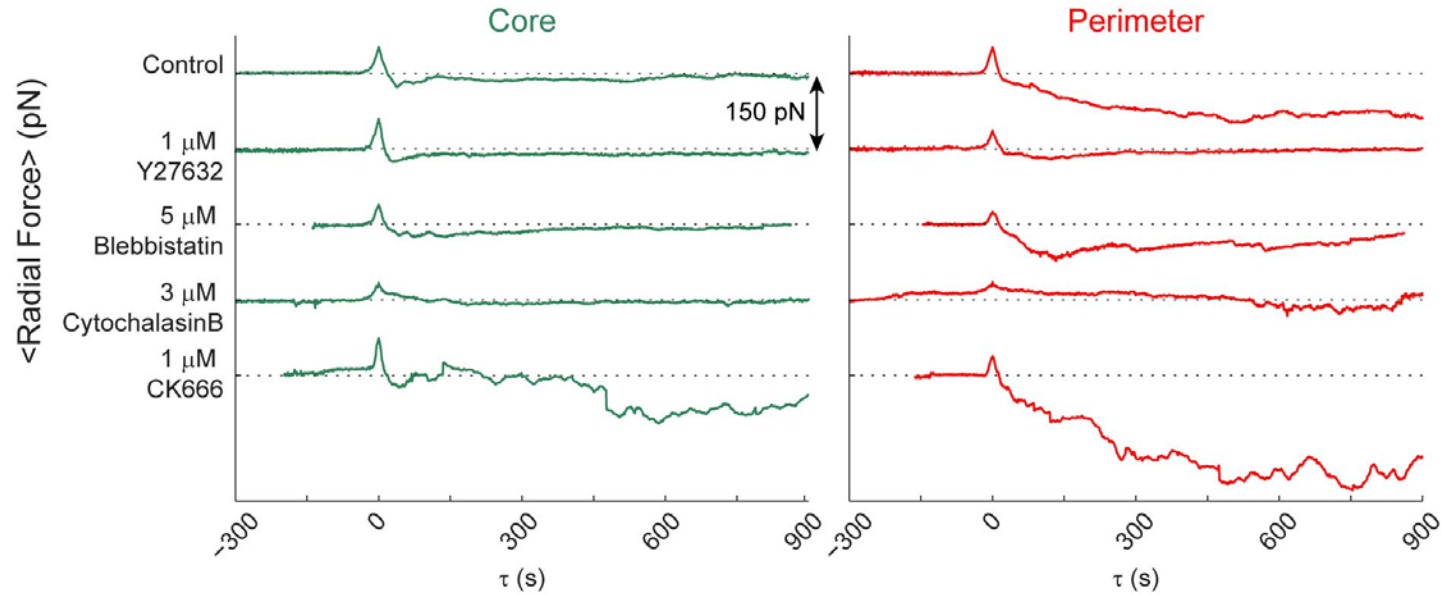
# \*\*Cortical stiffening via Jasplakinolide abrogates spreading

1  $\mu\text{M}$  Jasplakinolide inhibits cortical actin depolymerization



# Cortical softening **slows** spreading

...implies release of pre-stress is important



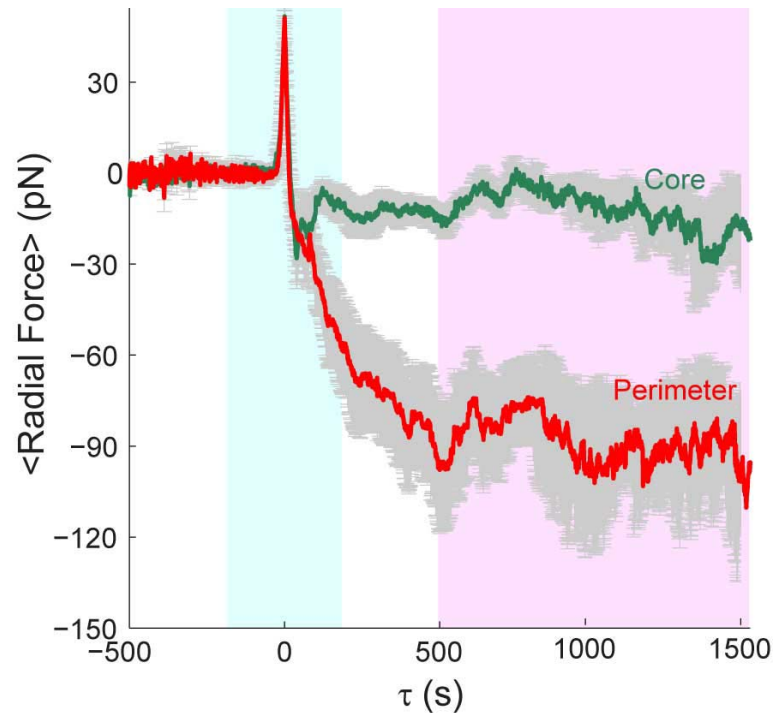
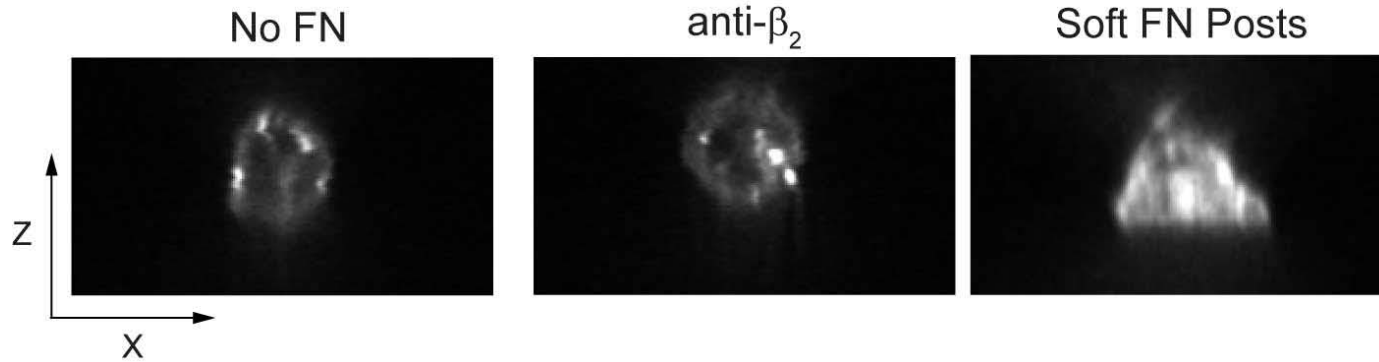
		Y27	Blebbi	CK666	Jasp	CytoB
		n cells	10	4	5	9
		n posts	325	117	161	n/a
Protrusion	<Fmax>	Core				
		Perimeter				
	<FWHM>	Core				* $\uparrow$
		Perimeter				* $\uparrow$
	<VAR(Fmax)>	Core				
		Perimeter				
Contraction	<Participation>	Core				
		Perimeter				
	<Velocity>	All Posts				* $\downarrow$
	<Fss>	Core				
	Perimeter	* $\downarrow$	* $\downarrow$			
	<VAR(Fss)>	Core				
		Perimeter	* $\downarrow$		* $\uparrow$	

No Spreading

= no sig. diff.

\*  $p < 0.05$ , Tukey-Kramer multi. comp.

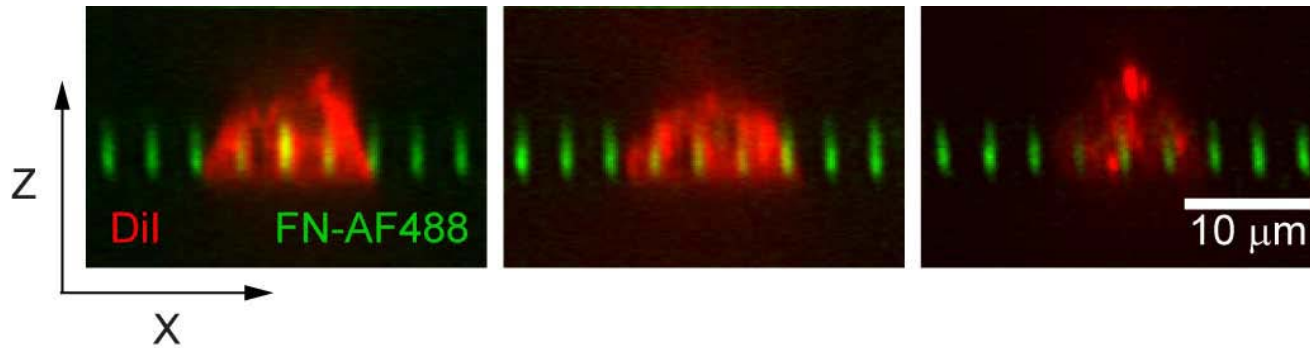
# Spreading is integrin-mediated but connection to the actomyosin substructure takes minutes to develop



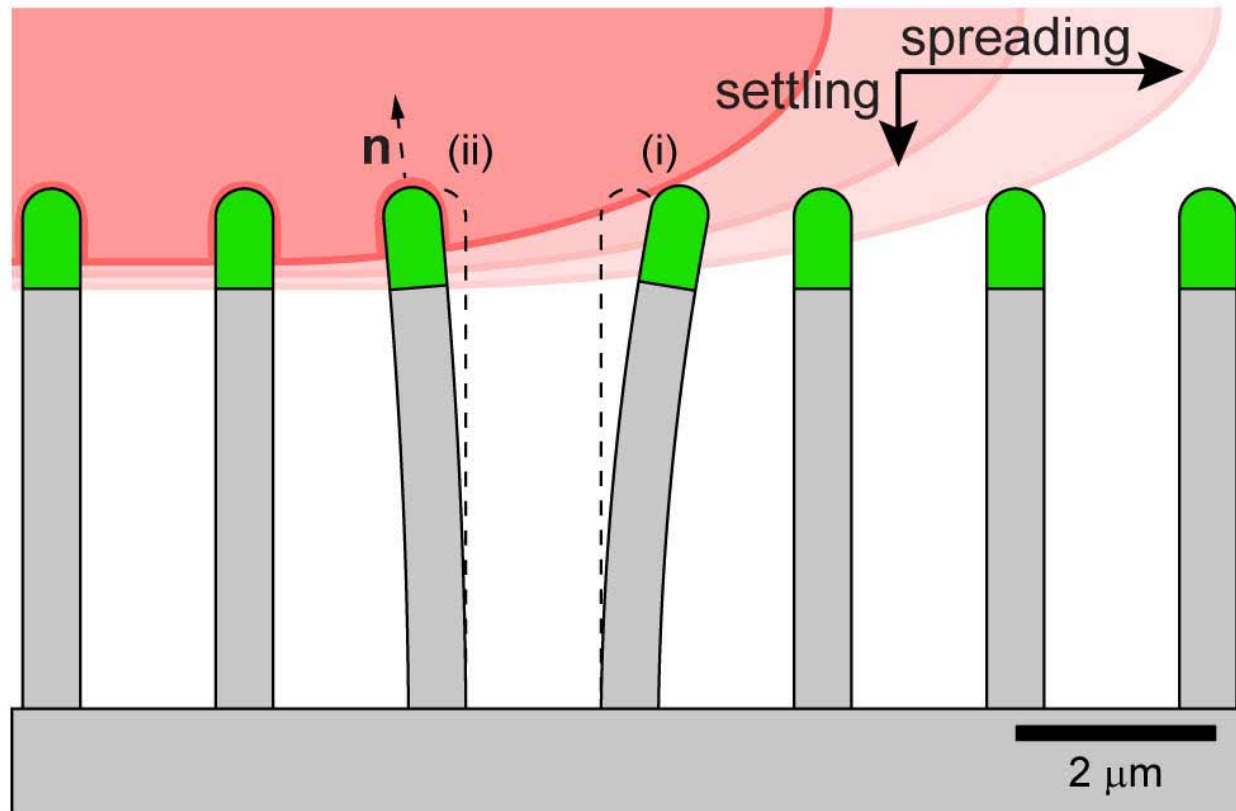
Why do we see protrusion at all?



# Invagination: a spreading neutrophil plows through posts

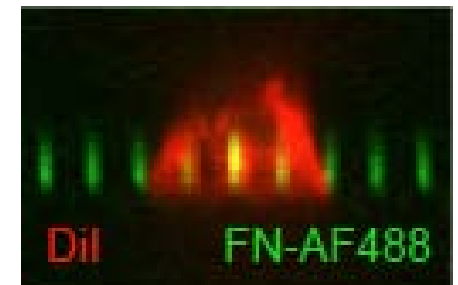
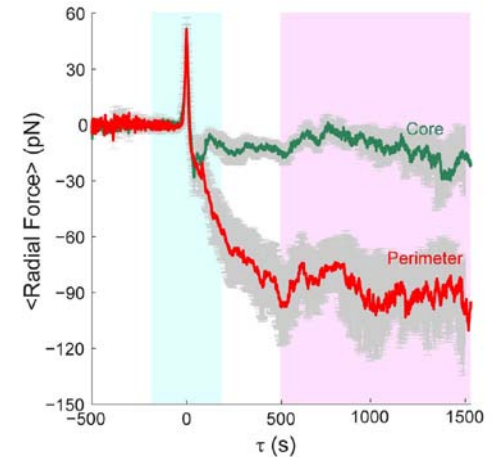
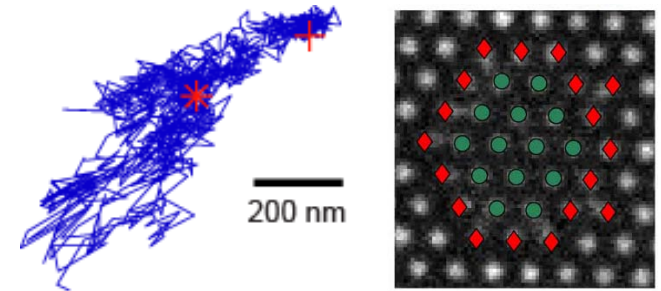


Schematic  
to scale



# Conclusions and Thank You!

- Two mechanical regimes:
  - Short time: transient protrusion
  - Long time: steady state contraction
- Contractility maintenance via canonical ROCK/myosin II pathways
- Resting cortical tension and release are critical to spreading
- Invagination of post arrays reports force of shape change



Manuscript submitted to *Biophysical Journal* for review